



Original Research Article

Molecular Detection of Some Virulence Genes in *Klebsiella pneumoniae* Isolates from Patients with UTI Infection.

Saba Saadoon Khazaal^{*}

College of Science, University of AL-Mustansiryiah, Baghdad, IRAQ *E-mail: sabask7@yahoo.com

Accepted 19 Oct, 2017

Abstract

Klebsiella pneumoniae causes a wide range of bacterial diseases like pneumonia, UTI and sepsis. Therefore, this study was done to assess the prevalence and molecular characteristics of *K. pneumoniae* in 24 samples of men and women isolated from Iraqi patients which was suffering from urinary tract infection (UTI) compared with (10) of healthy individuals. All samples were collected from Educational laboratories of Medical city/Iraqi Health Ministry. The samples were screened for the presence of kfu and k2A genes through PCR. The results of this study represented by (7 out of 24) isolates percent with (29.17%) positive for kfu gene and (9 out of 24) isolates (37.5%) were positive for k2A gene while the other (8) isolates were none of these genes. The two genes revealed significant differences between them (p< 0.5), so as with healthy group (p< 0.01). The study concluded that two genes kfu and k2A may affect on pathological of *K. pneumoniae*. The aim of this study represented by detection of kfu (iron uptake system gene) and k2A genes in patients with UTI which have a role in the pathogenicity of *K. pneumoniae* isolates.

Key Words: K. pneumoniae, kfu and k2A genes, UTI, Molecular detection, PCR.

التحري الجزيئي عن بعض جينات الضراوة في عزلات الكليبسيلا الرئوية من المرضى الذين يعانون التهاب المسالك البولية

<u>الخلاصة</u>

تسبب الكليبسيلا الرئوية مجموعة واسعة من الأمراض البكتيرية مثل الالتهاب الرئوي، التهاب المسالك البولية وخمج الدم. لذلك، فان هذه الدراسة قد بحثت الانتشار والخواص الجزيئية لـ (٢٤) عينة من النساء والذكور تم عزلها من المرضى العراقيين اللذين يعانون من إصابات المسالك البولية (UTI) مقارنة بـ (١٠) أفراد أصحاء. جميع العينات المرضية قد تم جمعها من المختبرات التعليمية لدائرة مدينة الطب / وزارة الصحة العراقية. تم فحص العينات عن وجود (١٠) أفراد أصحاء. جميع العينات المرضية قد تم جمعها من المختبرات التعليمية لدائرة مدينة الطب / وزارة الصحة العراقية. تم فحص العينات عن وجود جينات (kfu) و (K2A) من خلال تقنية تفاعل البلمرة المتسلسل PCR تمثلت نتائج هذه الدراسة بوجود (٧ من ٢٤) عزلة موجبة من جين (kfu) بنسبة جينات (١٠) أو (١٠) و (K2A) من خلال تقنية تفاعل البلمرة المتسلسل PCR تمثلت نتائج هذه الدراسة بوجود (٧ من ٢٤) عزلة موجبة من جين (kfu) بنسبة (١٠٩) و (١٠١٣) و (k2A) من خلال تقنية تفاعل البلمرة المتسلسل PCR تمثلت نتائج هذه الدراسة بوجود (٧ من ٢٤) عزلة موجبة من جين (kfu) بنسبة (١٩٥) و (١٧،٢٩) و (١٩ من ٢٤) عزلة موجبة من جين (kap بنسبة (٢٣٥، %) بينما تمثلت بقية العزلات الـ (٨) بعدم وجود هذه الجيات. اظهرت نتائج الدراسة عن وجود فروق معنوية بين وجود الجينين بنسبة (٤٥٩) و در٣٥، كن البينين في العزلات مقارنة مع الاصحاء. (الهمات عنوبي ولاد من الامي الموت نتائج الدراسة عن وجود فروق معنوية بين وجود الجينين بنسبة (kfu) و جود هذين الجينين في العزلات مقارنة مع الاصحاء. (٥٠٥) وكذلك بين وجود هذين الجينين في العزلات مقارنة مع الاصحاء. (٥٠٥) ولدراسة عن وجود الحينين بنسبة (لارسة ينه وجود هذين الجينين في المولين في العزلات مقارنة مع الاصحاء. (٥٠٥) وكذلك بين وجود هذين الجينين الموسيلين بالمجاري اليولية (المات عن وجود الجينين بنسبة (kfu) و جين (k24) في المرضى الحينين في العزلات مقارية معاوية (الهاي على ولارات. المولي و وردى ومال مال مولي في العربات معارية معان وليسليما عرفة عن وجود جين (kfu) و دولي مالماني و العربات معان و معن بكتريا الكليسيل الرئوية.

الكلمات المفتاحية: الكليبسيلا الرئوية، جينات kfu و k2A، التهاب المسالك البولية، التحري الجزيئي، تفاعل البلمرة المتسلسل.

Introduction

Kelebsiella is the oldest genus among their family, the normal habitat of this bacteria is the intestinal tract of human and animal, but may be transferred to another site causing a wide range of infectious diseases like in burn, wound, respiratory and urinary tract so as bacteremia. Klebsiella have a capsule that oppose host defenses which depend mainly on impair immune defenses and the bactericidal of effect serum mediated in large part by complement proteins [1].

They are facultative anaerobic, fermentative, They produce different virulence factors like adhesins, large capsule that are antiphagocytic, siderophores, and various endotoxins [2]. They are motile by peritrichous flagella, lactose fermenters. oxidase negative, and catalase positive. A common opportunistic pathogen of

community-acquired and nosocomial infections [1-3].

It is also linked with a distinguishing clinical characterized by syndrome communityacquired bacteremia with liver abscesses, and metastatic meningitis [4]. Serotype-specific genes like a chromosomal gene magA (mucoviscosity associated gene A) is restricted to gene cluster of K. pneumoniae capsule serotype K1 and the chromosomal K2 capsule associated gene A (k2A) for the K2 serotype [5-7] which isolates with capsule serotypes K1 and K2 are more resistant to phagocytosis than Non-K1/K2 strains [8, 9]. The k2A gene of K. pneumoniae could be used as a specific diagnostic technique to identify the cps of K. pneumoniae capsule K2 serotype, which matches to the magA region in the capsules gene clusters of K1 isolate [10]. While the kfu gene which codes for an iron uptake is a virulence gene, related with hyper-mucoviscosity phenotype and it is also linked to purulent infections of tissue caused by this potent pathogenic bacteria species [11].

The aim of this study was to detect the virulence genes kfu and k2A genes of K. *pneumoniae* in patients with UTIs that may have a role in the pathogenicity of this bacteria.

Materials and Methods: Samples collection

A total of (24) UTI samples from apparently sick patients were collected from established private laboratories and transported to the laboratory on ice. All specimens were managed for isolation of suspected bacterial isolates. Isolation and identification of bacterial isolates.

The isolation of bacteria was achieved using different standard techniques [1, 2] by culturing of urine specimens on blood agar and incubated for 24 hrs at 37°C. After that, the bacterial isolates were identified to the species level of using cultural and morphological characteristics of colonies grown on MacConkey agar. In addition to Gram's staining, and finally by biochemical tests according to the Cowan and Steels' manual for identification of medical bacteria [12]. The bacterial identification of isolates MJB-2017

was confirmed using API 20E system strips.

DNA Extraction

Template DNA from the colony was prepared with minor modifications. Genomic DNA extracted directly as leaflet kit (Geneaid company/Korea) for blood/culture. The assay was carried out following the instructions in the kit's leaflet from bacterial colonies grown on agar plates.

Amplification of virulence genes by PCR technique:

PCR was achieved using template DNA (3µl), primers (2µl) for two genes kfu F: (5-AGAACCTTCCTCGCTGAACA-3), R: (5 -ATAGTAGGCGAGCACCGAGA-3) and k2A gene F: (5-CAACCATGGTGGTCGATT AG-3), R: (5 -TGGTAGCCATATCCCTTTG G-3) and completed by 13ul DNase free water (Promega/ USA) in a total volume of 20 µl. The DNA for two genes were amplified using the modified cycling conditions (Applied-PCR/USA) Biosystem as in: Initial denaturation 95c° for 5 min. followed by 40 cycles consisting of 30 s of denaturing at 94°C, 30 Sec of annealing at 54°C, and 1 min of elongation at 72°C, followed by a final extension step at 72°C for 10 min. [13].

Results:

In the present study, the hyper mucoviscosity signs were positive in most isolates of all UTI patients and revealed the two genes kfu and k2A detected by conventional PCR technique (using specific primer sequences) yielded product sizes of 520 bp and 532 bp respectively Figure (1).

The study showed that out of a total of 24 isolates, nine (37.5%) were positive for k2A gene and (7) isolates (29.17%) were positive for kfu gene. The two genes revealed significant differences compared between them so as with negative control of healthy group represented by (p < 0.5) and (p < 0.01)respectively. While the other (No.= 8) isolates (33.3%) were negative for both genes and no significant differences were obtained compared with healthy negative of control group which had no bacterial growth (Tables 1 and 2).

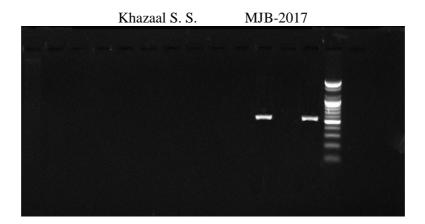


Figure (1) Ethidium bromide-PCR products separated in agarose gel (1.2%, Bio-Basic/Canada) at 75V, for 90 minutes, detected by UV transillumination. Amplified genes *kfu* and *k2A* with M.W. 520 and 532 respectively identified in *K. pneumoniae*, isolated from UTI (Lane M: 25bp/Bioneer ladder).

Table (1): Comparison between negative and positive of pathogenesis of K. pneumoniae growth in patients
with LITL infections

_	K. pneumonia growth			
Test	No.	%		
+	16	66.67		
_	8	33.33		
Total	24	100%		
P value		0.0001**		

Table (2): Comparison between two genes kfu and k2A affected on K. pneumoniae pathogenesis

Group	Total No.	Gene k2A		Gene kfu		
		+	-	+	-	P value
Patients	24	9 (37.5)	15 (62.50)	7 (29.17)	17 (70.83)	0.041*
Control	10	0 (0.00)	10 (100)	0 (0.00)	10 (100)	
P value		0.0001**		0.0001**		

 $(P\!\!<\!\!0.5)^*, (P\!\!<\!\!0.01)^{**}$

Discussion:

The majority of gene encoding capsule has been carried on bacterial plasmids, which the studies revealed that the rmpA gene responsible for produce of capsule protect bacteria from macrophage [14-16]. In the present study, the hyper mucoviscosity signs were positive in most isolates of all UTI patients. These results were in agreement with a study reported by Turton and his colleagues [17]. An extensive range of *K. pneumoniae* infections had been described worldwide,

Khazaal S. S.

including pneumonia, UTIs, meningitis, in addition to different types abscess at different sites.

The mechanism of infections in *Klebsiella* species involve production of various virulence factors such as hyper-mucoviscosity capsular serotype rmpA gene, particularly K1 or K2, and virulence related genes, like kfu and k2A [9, 18]. The study revealed the two genes kfu and k2A yielded product sizes of 520 bp and 532 bp respectively.

As a later of previous studies for, magA and k2A specific to K2 capsule serotype were reported to be specific to capsule gene clusters of K1 and K2 serotype, respectively [19, 20]. A specific to the capsule gene clusters of K1 and K2 of both kfu and k2A serotype were found in this study [13], which known to be related to virulence factors of *K. pneumoniae* [20].

The study done a huge number of virulence gene dissemination and clinical conditions caused by *K. pneumoniae*, which found that there was a statistically significant difference in the incidence of phenotype, *kfu* and *k2A* as explained above which corresponded with previous studies [18-21].

Nevertheless, comprehensive evidence about the spreading of capsular K serotypes of causative organisms and the clinical features of subjects with K. pneumoniae in UTI and their interrelationships were reported. The investigation of the laboratory data, disease outcomes and clinical relevance of patients acquired purulent K. pneumoniae infections was done, and the prevalence of virulence associated of kfu genes were found at the rate of 29%, which is in extremely contrast with a study of Yu and his colleagues who detected the prevalence at the rate of 35% in UTI disease from these defect of genes. Detection of these genes may specify the virulence impending of the isolates [5, 13].

However, until now, little data are available about the pathogenicity of this bacterium. Certain current clonal analyses of *K*. *pneumoniae* isolates show that there are diverse clonal groups, certain of which may be linked with definite disease conditions. Nevertheless, what render one clonal cluster more virulent and what change the disease outline are not so far clear and continue as a vital question for the upcoming days [22-24]. MJB-2017

Conclusion:

The genetic analysis of these genes kfu and k2A offers an opportunity for prompt diagnosis of the infection from predisposed patient.

References:

- 1. Doyle J. and Evans J. Klebsiella. Access Science. 2008;33: 100-300.
- Fang CT., Chuang YP., Shun CT., Chang SC. and Wang JT. A novel virulence gene in *K*. *pneumoniae* strains causing primary liver abscess and septic metastatic complications. J. Exp. Med. 2004;199:697–705.
- Al-Charrakh AH., Yousif SY. and Al-Janabi HS. Occurrence and detection of extendedspectrum-β-lactamases in Klebsiella isolates in Hilla, Iraq. Afr. J. Biotechnol. 2011;10 (4): 657-665.
- Ko WC., Paterson DL. and Sagnimeni, A.J. Community-acquired Klebsiella pneumoniae bacteremia: global differences in clinical patterns. Emerg. Infect. Dis. 2002;8:160–166.
- Yu W., Ko W., Cheng K., Lee H., Ke D., Lee C.;, Fung C. and Chuang Y. Association between rmpA and magA Genes and Clinical Syndromes Caused by *Klebsiella pneumoniae* in Taiwan. Clin. Infect. Dis. 2006;42:1351– 1358.
- Kenne L. and Linberg B. Bacterial polysaccharides, In G.O. Aspimal (ed.), The polysaccharides. Academic Press Inc., New York, pp.1983;287–363.
- Doud M., Zeppegno R., Molina E., Miller N., Balachandar D., Schneper L., Poppiti R. and Mathee K. A k2A-positive *Klebsiella pneumoniae* causes liver and brain abscess in a Saint Kitt's man, Int. J. Med. Sci. 2009;6(6): 301-304.
- 8. Wang L., Gu H. and Lu, X. A rapid low-cost real-time PCR for the detection of *Klebsiella pneumoniae* carbapenemase genes, Annals of Clin Microbiol Antimicrob. 2012; 11(9):1-6.
- Podschun R. and Ullmann U. *Klebsiella* spp. as Nosocomial Pathogens: Epidemiology, Taxonomy, Typing Methods, and Pathogenicity Factors. Clin. Microbiol. Rev. 1998; 11(4):589–603.
- Chuang Y., Fang C., Lai S., Chang S. and Wang J. Genetic determinants of capsular serotype K1 of *Klebsiella pneumoniae* causing primary pyogenic liver abscess. J Infect. Dis. 2006; 193:645-654.

Khazaal S. S.

- Ma LC., Fang CT., Lee CZ., Shun CT. and Wang JT. Genomic heterogeneity in *K. pneumoniae* strains is associated with primary pyogenic liver abscess and metastatic infection. J. Infect. Dis. 2005;192:117–128.
- Barrow GI. and Feltham RKA. Cowan and Steels' manual for identification of medical bacteria. 3rd edition. United Kingdom: Cambridge University Press.2003.
- Harald P., Kathrine B., Peter T., Yvonne P., Wolfgang W., Kate T. and Till TB. Direct Detection and Genotypingof *Klebsiella pneumoniae* Carbapenemases from Urine by Use of a New DNA Microarray Test. J Clin Microbial. 2012; 50(12):3990–3998.
- 14. Sharma RK., Boro BR. and Borachi PC. Incidence of Caprine Pneumonia and associated bacterial isolates. Ind. J. Anim. Sci. 1991; 61:54-55.
- 15. Yu WL., Ko WC., Cheng KC., Lee HC., Ke DS., Lee CC. et al. Association between rmpA and magA genes and clinical syndromes caused by *Klebsiella pneumoniae* in Taiwan. Clin Infect Dis. 2006;42:1351-8.
- Nassif X., Honoré N., Vasselon T., Cole ST., Sansonetti PJ. Positive control of colanic acid synthesis in Escherichia coli by rmpA and rmpB, two virulence-plasmid genes of *Klebsiella pneumoniae*. Mol Microbial. 1989; 3:1349-59.
- Turton JF., Englender H., Gabriel SN., Turton SE., Kaufmann ME. and Pitt, TL. Genetically similar isolates of *Klebsiella pneumoniae* serotype K1 causing liver abscesses in three continents. J. Med. Microbio. 2007;56:593– 597.
- Ko WC., Paterson DL., Sagnimeni AJ., Hansen DS., Von Gottberg A., Mohapatra, S. et al. Community-acquired *Klebsiella pneumoniae* bacteremia: global differences in clinical patterns. Emerg Infect Dis. 2002;8:160-6.
- 19. Struve C., Bojer M., Nielsen FM., Hansen DS., Krogfelt KA. Investigation of the putative virulence gene magA in a worldwide collection of 495 *Klebsiella* isolates: magA is restricted to the gene cluster of *Klebsiella pneumoniae* capsule serotype K1. J Med Microbiol. 2005;54:1111-3.
- 20. Yu WL., Fung CP., Ko WC., Cheng KC., Lee CC., Chuang YC. Polymerase chain reaction analysis for detecting capsule serotypes K1 and K2 of *Klebsiella pneumoniae* causing abscesses of the liver and other sites. J Infect Dis.2007; 195:1235-6.

MJB-2017

- 21. Ma LC., Fang CT., Lee CZ., Shun CT., Wang JT. Genomic heterogeneity in *Klebsiella pneumoniae* strains is associated with primary pyogenic liver abscess and metastatic infection. J Infect Dis. 2005; 192:117-28.
- Christopher A., Michelle P. and Virginia, LM. *Klebsiella*: a long way to go towards understanding this enigmatic jet-setter. F1000 Prime. 2014;6: 64.
- AL-Kadmy IMS., Musafer HK., Abd Ali MH. and Sajid, S. Detection of 16S Ribosomal RNA Methylation in Extended-Spectrum βlactamase-Producing *Klebsiella pneumoniae* Clinical Isolates from Baghdad Hospitals. Iraqi Journal of Biotechnology. 2015;14(1):53-62.
- 24. Al-Charrakh AH. Bacteriological and genetic study on extended-spectrum Beta-lactamases and bacteriocins of *Klebsiella* isolated from Hilla city. Ph.D Thesis, College of Science, Baghdad University.2005.