

DOI: <http://doi.org/10.32792/utq.jceps.09.01.18>

Detection of *clfA* Gene in *Staphylococcus aureus* Isolated From Tonsillitis Patients

Zainab D. Degaim¹, Abbas D. Mater², Esraa D. Taher¹

¹ College of Medicine, University of Thi-Qar, Thi-Qar, Iraq.

² College of Dentistry, University of Thi-Qar, Thi-Qar, Iraq.

Abstract

Staphylococcus aureus is an opportunistic pathogen and it was one of virulent causer of tonsillitis. The present investigation was aimed to the molecular detection of adhesion gene (*clfA*) was done by polymerase chain reaction (PCR) and DNA sequencing. Only 64 isolates (42%) were mannitol fermenter and recorded as *S. aureus*, these isolates recovered from 152 swabs were collected from tonsillitis patients in ENT department in Al-Habboby Teaching Hospital, Thi-Qar province, during the period from November 2016 to March 2017. Out of 64 isolates, 56 (87.5%) were harbored *clfA* gene. The sequencing of PCR products showed significant alignments identities (96-99%) to the *S. aureus* which are located in BLAST-NCBI Genbank. Phylogenetic analysis of *S. aureus* based upon the neighbour-joining of partial *clfA* gene sequences showed that these sequences were derived from *Staphylococcus* genes.

Keywords: *S. aureus*, tonsillitis, *clfA* gene, gene sequencing.

Introduction:

Staphylococcus aureus is an opportunistic pathogen with the ability to invade and persist in unprofessional phagocytes: fibroblasts, osteoblasts and different types of epithelial cells. The infectious potential of this bacterium is determined by a large number of cell-associated and extracellular virulence factors, some of which are implicated in the adhesion process and others in the bacterial invasion (Holban *et al.*, 2013). *S. aureus* can produce secreted virulence factors such as enterotoxins and surface-exposed virulence factors (fibrinogen, protein A, fibronectin binding proteins) (Zhang *et al.*, 2016). Fibrinogen is the most abundant host protein in endothelial lesions. Clumping factors A and B (*ClfA*, *ClfB*) are fibrinogen-binding proteins expressed by *S. aureus* on bacterial cells, promoting adherence to cell surfaces. The *ClfA* factor is expressed during the bacterial growth, whereas *ClfB* is present only during the early logarithmic phase (Peacock *et al.*, 2000). The clumping factor is very important for the virulence of *S. aureus*, and is thought to be essential for colonization and establishment of infections, It participates in the infection process by facilitating bacterial binding via soluble or immobilized fibrinogen as fibrinogen plays a significant role in platelet thrombus formation and almost all *S. aureus* strains have the *clfA* gene (Josefsson *et al.*, 2008; Karahan *et al.*, 2011; Delfani *et al.*, 2016). Microbial surface components recognizing adhesive matrix molecules on *S. aureus* surface, mediate staphylococcal adherence to components of the extracellular matrix of the host (Vazquez *et al.*, 2011). These components are attached covalently to peptidoglycan by sortase enzymes (Heilmann, 2011). Furthermore, these components participate in biofilm formation, in addition to the *ica* operon that produces the polysaccharide intercellular adhesion (PIA) (Mirzaee *et al.*, 2014). MSCRAMMs can bind to molecules such as collagen (mostly via Cna), fibronectin (via FnbAB), and fibrinogen (with ClfAB and Fib) and thus evade immune system, and then can develop infections (Foster *et al.*, 2014). The aim of this study was to screen the *clfA* gene among the isolates of *S. aureus* from tonsillitis patients. Likewise, to comparative genomic sequencing analysis and phylogenetic tree generating, allows for an epidemiological discrimination of closely related bacterial isolates.

Material and methods

Bacterial isolates

One hundred and fifty two swabs were collected from patients infected with tonsillitis whom admitted to ENT unit in AL-Habbuby Teaching Hospital of Thi-Qar province, during the period from November 2016 to March 2017 by moistened sterile swabs with normal saline, these swabs directly inoculated on mannitol salt agar (LAB/ United Kingdom) and incubated at 37°C for 24 h.

Identification of *S. aureus*

S. aureus was identified depending on the morphological properties on culture media and biochemical tests (Catalase test, Coagulase test, DNase production test) which done according to Bergeys manual (Harley & Prescott, 2002 ; Brooks *et al.*, 2007). API Staph system (BioMerieux, France) was used to identify a *Staphylococcus* and *Micrococcus*.

Bacterial DNA extraction

All isolates of *S. aureus* had been incubated on Brain Heart Infusion Broth (LAB/ United Kingdom) for 18–24 h at 37° C. Chromosomal DNA extraction was performed using Genomic DNA Extraction kit (Geneaid/Korea).

The specific primer pairs of *clfA* gene as following: forward: 5'- ATT GGC GTG GCT TCA GTG CT -3' and reverse: 5'- CGT TTC TTC CGT AGT TGC ATT TG -3'. The PCR cycling conditions of this gene: initial denaturation at 94°C for 5 min, followed by 25 cycles of denaturation at 94°C for 1 min, annealing at 55°C for 1 min, extension at 72°C for 1 min and final extension for 10 min after the last cycle (Tristan *et al.*, 2003). Electrophoresis of PCR product was carried out in 1% agarose gel and the presence of a 292bp band indicate a positive result for this gene.

Results and Discussion

The results of the present study showed that the incidence of *S. aureus* was 64 isolates (42%). *S. aureus* was one of the most common and virulent causer of tonsillitis (Jeong *et al.*, 2007).

The recent data differed from the local study in the same field which recorded higher percentages of *S. aureus* prevalence, such as Dakhil & Hamim, (2016) which revealed that the incidence of *S. aureus* was 62 (64.5 %) from which 60 isolates (96.77%) that recovered from tonsillitis.

The molecular analysis of biofilm-associated gene (*clfA* gene) revealed that 87.5% of isolates had this gene, and the size of this gene was approximately 292 bp, Figure(1). Clumping factor A was one of important virulence factors which critical for pathogenicity of invasion *S. aureus* and colonization in infection sites. Momtaz *et al.*, (2010) suggest that clinical strains of *S. aureus* may contain different frequencies of clumping factors, being essential for colonization. *S. aureus* has different mechanisms of virulence, pathogenicity and favors the development of antibiotic resistance and increases vulnerability to infection (Almeida *et al.*, 2007). Moreover, Gotz, (2002) suggested that infections related with biofilm production are generally frequent, because the antimicrobial treatment predominantly eliminates planktonic forms, leaving the sessile cells free to reproduce and propagate the biofilm after treatment, so, the pathogen in biofilms are more protected against the host immune system. The biofilm-associated diseases include infections caused by heart valve implants, catheters, and contact lenses.

The results of current data disagreed with results of Atshan *et al.*, (2012); Ghasemian *et al.*, (2015) and Omara *et al.*, (2016) documented that all *S. aureus* strains carried *clfA* gene, while Gowrishankar *et al.*, (2016) confirmed through their study that only 58.7% of isolates harbored this gene.

There was didn't expression of *cflA* gene between MRSA and MSSA strains (Souza *et al.*, 2014).

The occurrence of *clfA* gene may be abundant in clinical *S. aureus* isolates compared with that from animal sources. Momtaz *et al.*, (2010) reported that nearly 20% of *S. aureus* isolates causing mastitis contain *clfA* gene.

The evolutionary history was inferred using the Neighbor-Joining method Saitou and Nei, (1987). The optimal tree with the sum of branch length = 0.03635857 is shown. (above the branches). The evolutionary distances were computed using the Maximum Composite Likelihood method Tamura *et al.*, (2004), and were in the units of the number of base substitutions per site, Figure (2). The analysis involved 8 nucleotide sequences. Codon positions included were 1st+2nd+3rd+Noncoding. All ambiguous positions were removed for each sequence pair. There were a total of 503 positions in the final dataset. Evolutionary analyses were conducted in MEGA7 Kumar *et al.*, (2016).

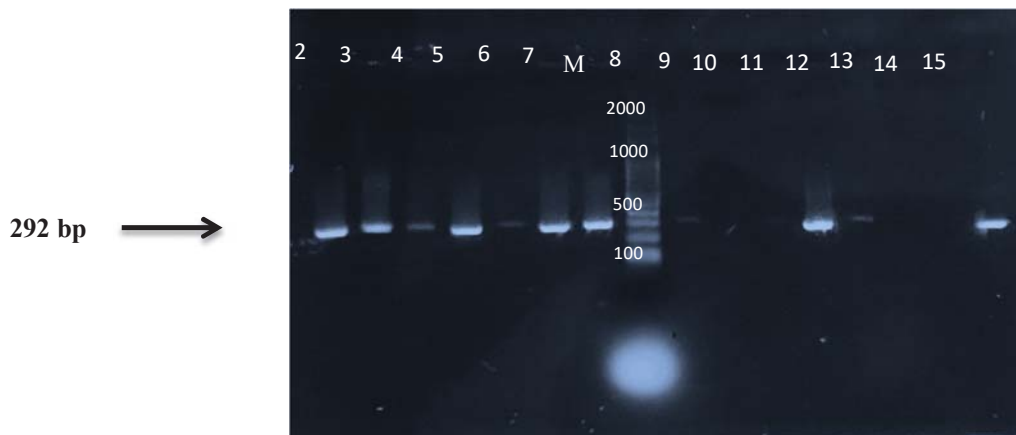


Figure (1): Agarose gel electrophoresis of *clfA* gene amplification, M: ladder, 1-8, 10-12, 15: positive results, 9,13-14: negative result.

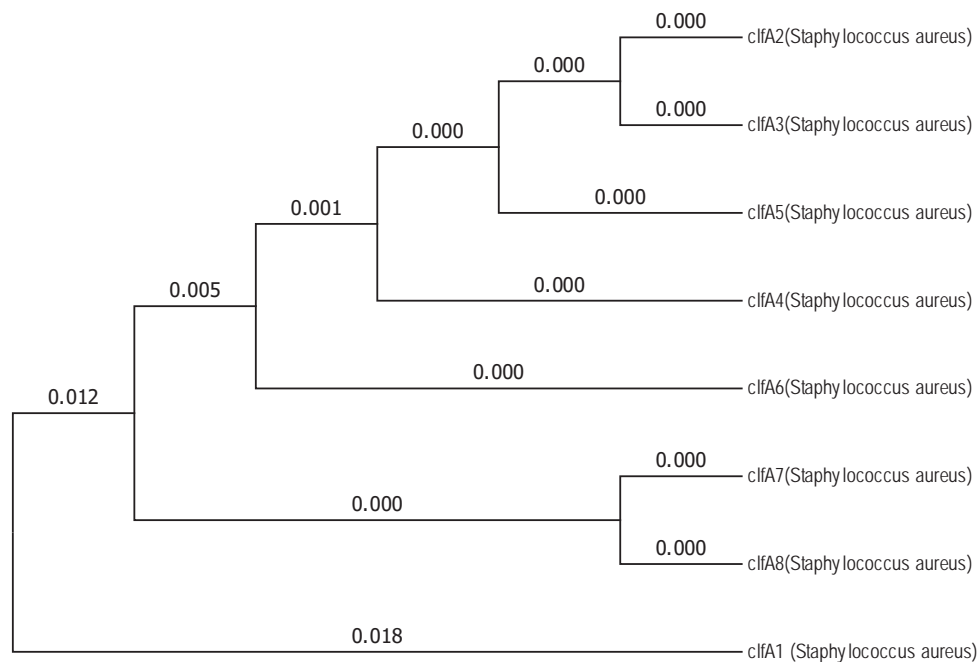


Figure (2): Evolutionary relationships of taxa, based on Clumping factors gene partial sequence that used for *S. aureus* detection from Human tonsillitis samples.

Conclusion:

The recent results recorded a high percentage of *clfA* gene which related with promoting adherence to cell surfaces and biofilm production that may be increased the pathogenicity of this bacteria which caused different human diseases.

References:

1. ♦ Almeida, M I.; Bedendo, J.; Cavasin, E D. and Tognim, M C. (2007). Prevalência e perfil de sensibilidade de amostras de *Staphylococcus aureus* isoladas de casos clínicos de infecções hospitalares. J. Revista. Eletrônica. de Enfermagem., 9: 489-495.
2. ♦ Atshan, S S.; Nor Shamsudin, M.; Sekawi, Z.; Lung, L T.; Hamat, R A.; Karunanidhi, A.; Ali, A.; Ghaznavi-Rad, E.; Ghasemzadeh-Moghaddam, H.; Seng, J S.; Nathan, J J. and Chong Pei Pei, C P. (2012). Prevalence of adhesion and regulation of biofilm-related genes in different clones of *Staphylococcus aureus*. J. Biomed. Biotechnol. Pp1-10.
3. ♦ Brooks, G F.; Caroll, K C. and Morse, S A. *Staphylococcus* (Jawetz). (2007). Melnick and Adelberg's, Medical Microbiology. (24th ed). The McGraw-Hill. New York.
4. ♦ Dakhil, B R. and Hamim, S S. (2016). Antibiotic susceptibility of *Streptococcus pyogenes* and *Staphylococcus aureus* isolated from pharyngitis and tonsillitis patients in Nasiriyah city, Iraq. World. J. Pharm. Sci., 4(4): 14-19.
5. ♦ Delfani, S.; Mobarez, A M.; Fooladi, A A I.; Amani, J. and Emaneini, M. (2016). Protection of mice against *Staphylococcus aureus* infection by a recombinant protein ClfA–IsdB–Hlg as a vaccine candidate. J. Med. Microbiol. Immunol., 205:47–55.
6. ♦ Foster, T J.; Geoghegan, J A. Ganesh, V K. and Höök, M. (2014). Adhesion, invasion and evasion: the many functions of the surface proteins of *Staphylococcus aureus*. J. Nat. Rev. Microbiol., 12(1):49-62.
7. ♦ Ghasemian, A.; Peerayeh, S N.; Bakhshi, B. and Mohsen Mirzaee, M. (2015). The microbial surface components recognizing adhesive matrix molecules (MSCRAMMs) genes among clinical isolates of *Staphylococcus aureus* from hospitalized children. Iran. J. Pathol., 10(4): 258 – 264.
8. ♦ Gotz, F. (2002). *Staphylococcus* and Biofilms. Molecular Microbiology., 43: 1367-1378.
9. ♦ Gowrishankar, S.; Kamaladevi, A.; Balamurugan, K. and Pandian, S. (2016). In vitro and in vivo biofilm characterization of methicillin-resistant *Staphylococcus aureus* from patients associated with pharyngitis infection. J. Bio. Med. Res. Intern., Pp:1-14.
10. ♦ Harley, J P. and Prescott, L M. (2002). Laboratory Exercises in Microbiology. (5th ed). The McGraw-Hill Companies, Inc., New York.
11. ♦ Heilmann, C. (2011). Adhesion mechanisms of staphylococci. J. Adv. Exp. Med. Biol., 715:105-23.
12. ♦ Holban, A.; Cotar, A C.; Chifiriuc, M C.; Bleotu, C.; Banu, O. and Lazar, V. (2013). Variation of virulence profiles in some *Staphylococcus aureus* and *Pseudomonas aeruginosa* stains isolated from different clinical patients, African. J. Microbiol. Res., 7: 3453-3460.
13. ♦ Jeong, J H.; Lee, DW.; Ryu, R A.; Lee, Y S.; Leeshh, L. and Kang, J O. (2007). Bacteriological comparison of tonsil core in recurrent tonsillitis and tonsillar hypertrophy. Laryngoscope, 117(12):2146-2151.
14. ♦ Josefsson, E.; Higgins, J.; Foster, T J. and Tarkowski, A. (2008). Fibrinogen binding sites P336 and Y338 of clumping factor A are crucial for *Staphylococcus aureus* virulence. J. PLoS. One., 3(5):e2206.

15. ♦ Karahan, M.; Nuri Aciki, M. and Cetinkaya, B. (2011). Investigation of virulence genes by PCR in *Staphylococcus aureus* isolates originated from subclinical bovine mastitis in Turkey. *J. Pak. Vet.*, 31(3): 249-253.
16. ♦ Kumar S.; Stecher, G. and Tamura, K. MEGA7: (2016). Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. *J. Mol. Bio. Evol.*, 33:1870-1874.
17. ♦ Mirzaee, M.; Najar Peerayeh, Sh. and Ghasemian, A M. (2014). Detection of *icaABCD* genes and biofilm formation in clinical isolates of methicillin resistant *Staphylococcus aureus*. *Iran. J. Pathol.*; 9 (4), 257-262.
18. ♦ Momtaz, H.; Rahimi, E. and Tajbakhsh, E. (2010). Detection of some virulence factors in *Staphylococcus aureus* isolated from clinical and subclinical bovine mastitis in Iran. *Afr. J. Biotechnol.*, 9 (25): 3753-3758.
19. ♦ Omara, S T.; Syame, S M. and Elgabry, E A. (2016). Molecular detection of the clumping factor (fibrinogen receptor) in the enterotoxigenic *S. aureus* isolated from raw milk and traditional cheese. *Intern. J. of Chem. Tech. Res. CODEN (USA): IJCRGG.*, 9(12): 923-933.
20. ♦ Peacock, S J.; Day, N P J.; Thomas, M G.; Berendt, A R. and Foster, T J. (2000). Clinical isolates of *Staphylococcus aureus* exhibit diversity in *fnb* genes and adhesion to human fibronectin. *J. Infect.*, 41(1):23-31.
21. ♦ Saitou, N. and Nei, M. (1987). The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution.*, 4:406-425.
22. ♦ Souza, S G.; Campos, G B.; Oliveira, P S.; Sousa, D S.; Da Silva, D C C.; Santos, V M.; Amorim, A T.; Santos, A M O.; Timenetsky, J.; Cruz, M P.; Yatsuda, R. and Marques, L M. (2014). Virulence factors in Methicillin-Resistant *Staphylococcus aureus* isolated from ICU units in Brazil. *J. Advanc. Microbol.*, 4:207-215.
23. ♦ Tamura, K., Nei, M. and Kumar, S. (2004). Prospects for inferring very large phylogenies by using the neighbor-joining method. *Proceedings of the National Academy of Sciences (USA).*, 101:11030-11035.
24. ♦ Tristan, A.; Ying, L.; Bes, M.; Etienne, J.; Vandenesch, F. and Lina, G. (2003). Use of multiplex PCR to identify *Staphylococcus aureus* adhesins involved in human hematogenous infections. *J. Clin. Microbiol.* 41(9): 4465–4467.
25. ♦ Vazquez, V.; Liang, X.; Horndahl, J K.; Ganesh, V K.; Smeds, E.; Foster, T J. and Hook, M. (2011). Fibrinogen is a ligand for the *Staphylococcus aureus* microbial surface components recognizing adhesive matrix molecules (MSCRAMM) bone sialoprotein-binding protein (Bbp). *J. Biol. Chem.*, 286 (34): 29797-29805.
26. ♦ Zhang, L.; Li, Y.; Bao, H.; Wei, R.; Zhou, Y.; Zhang, H. and Wang, R. (2016). Population structure and antimicrobial profile of *Staphylococcus aureus* strains associated with bovine mastitis in China. *J. Micro. Path.*, 97; 103-109.