#### Research Article

## **AL-ANBAR JOURNAL OF VETERINARY SCIENCES**

Vol. 13 Issue:2, (2020)

ISSN: P-1999:6527 E-2707:0603

# Effect of Monthly Variations on the Plasma Membrane, Acrosome and DNA Integrities of Spermatozoa in Friesian Holstein Bulls Born in Iraq

# **Qusay Mohammed Aboud\* and Saad Akram Hatif**

Department of surgery & obstetrics, College of Veterinary Medicine, University of Baghdad, Iraq

\*Corresponding author; qusaymoh988@uofallujah.edu.iq

# Doi: https://doi.org/10.37940/AJVS.2020.13.2.11

Received: 2/9/2020 Accepted:4/12/2020

This article is licensed under a CC BY (Creative Commons Attribution 4.0) http://creativecommons.org/licenses/by/4.0/.

#### **Abstract**

This study was aimed to evaluate the influence of months of the year on the quality of semen in Holstein bulls. A study carried out at artificial insemination center/ Abou-Ghareeb/ western of Baghdad. A total of 160 ejaculates were collected from 15 bulls born in Iraq via artificial vagina. The age of the bulls ranged between 4 to 5 years and the study period were December to March and September. The semen samples were diluted with Tris base extender. The semen were packed in a straw according to the program of artificial insemination center. Semen characteristics (plasma membrane, acrosome, and DNA integrities) were evaluated. The results revealed a significant decrease ( $P \le 0.01$ ) in plasma membrane and acrosome integrity in September as compared with December, January, February, and March. There was a significant decline ( $P \le 0.05$ ) in DNA integrity in September as compared with December, January, February, and March in fresh and frozen semen. In conclusion, the September month had a negative effect on the plasma membrane, acrosome, and DNA percentage in all bulls.

# Keywards: Holstein bulls, Semen, DNA, Acrosome, HOST

تأثير التغيرات الشهرية على غشاء البلازما ، الأكروسوم وسلامة المادة الوراثية في نطف ثيران الفريزيان هولشتاين المولودة في المعروبة المعروبة على المولودة في المعروبة ا

#### الخلاصة

هدفت هذه الدراسة إلى تقييم تأثير أشهر السنة على جودة السائل المنوي في ثيران الهولشتاين. تم اجراء هذه الدراسة في مركز التلقيح الاصطناعي / أبو غريب/ غرب بغداد. تم جمع 160 قذفة من 15 ثور مولود في العراق باستخدام المهبل الاصطناعي. كانت أعمار الثيران تتراوح بين 4-5 سنوات. اجريت الدراسة خلال الفترة من كانون الاول إلى أذار وأيلول. خفف السائل المنوي بمخفف الترس واعتمد برنامج تجميد السائل المنوي من قبل مركز التلقيح الاصطناعي. قيم السائل المنوي من خلال فحص سلامة كل من غشاء البلازما والاكروسوم والمادة الوراثية. أظهرت النتائج انخفاضًا معنويًا ( $P \leq 0.01$ ) في سلامة غشاء البلازما وسلامة الأكروسوم في أيلول مقارنة بشهر كانون الاول وكانون الأالي وشباط وأذار. كذلك انخفضت سلامة المادة الوراثية أيضًا بشكل ملحوظ ( $P \leq 0.05$ ) في أيلول مقارنة بشهر كانون الاول وكانون الثاني وشباط وأذار في السائل المنوي الطازج والمجمد. وقد استنتج من الدراسة ان لشهر أيلول تأثير سلبي على غشاء البلازما والأكروسوم ونسبة سلامة المادة الوراثية على نطف جميع الثيران.

Issue:2, (2020)

ISSN: P-1999:6527 E-2707:0603

#### Introduction

In most domestic animals, the veterinarians and veterinary specialists used artificial insemination (AI)(1). Characteristics of fresh and post-thaw bovine semen are affected by many factors such as age, genotype, breed and individual traits(2-9). However climate, relative humidity, season, daylight and ambient temperature can also affected quality(4,8,10-12). Ball and Peters(13) reports that the ambient temperature affect the characteristics of sperm at collection time, during spermatogenesis and during maturation of spermatozoa in the epididymis, Approximately 70 days prior to collection. It has been reported by several authors that seasonal and monthly changes in semen quality induced by disorders of thermoregulatory mechanisms due to heat stress (11,12,14-18) reducing conception and increasing embryo mortality (19-21). Garcia-Peniche et al. (22) observed that Holsteins are sensitive to heat stress which was also reported by several workers during 1968 to 2020 in Iraq (23-27).

Spermatozoa has a functional membrane needed for the Sperm fertilizing potential, since it is an essential part of sperm capacitation, acrosomal reactions, and attachment of sperm to the surface of the egg (28). The hypoosmotic swelling (HOS) test (29), acrosome integrity (30,31) and DNA integrity (32), it acts as a valuable measure of sperm fertility capacity (33). The current study was aimed to know the effect

of months of collection includes Dec. to March and September on fresh and thawed sperm characteristics in Holstein bulls born in Iraq.

#### Materials and methods

Fresh and thawed semen characteristics from 160 ejaculates were collected from fifteen Holstein bulls by artificial vagina during the period from December, January, February, March and September, presented at the Artificial Insemination Center, Abou-Ghareeb, Iraq. The animals aged between 4 to 5 years. Semen (n = 160) was collected from bulls weekly by artificial vagina method. At collection, fresh semen was kept in water bath at 37C° diluted with Tris-Fructose-Egg Yolk-Glycerol (TFEG) extender to yield 80 million sperms per ml, cooled to 5 C° 4hrs packed in 0.25 ml polyvinyl French straws (IMV, France) placed horizontally on a rack in 4cm above liquid nitrogen (LN2) for 9 min then dipped in LN2(34). Frozen straws were melted at 37 C° 30 seconds in a water bath. For fresh and thawed samples spermatozoa assessment, plasma membrane integrity by HOST as described by Ramu and Jeyendran (28). Sperm acrosomal integrity were calculated using Giemsa stain as described by Kovacs and Foote (35). Sperm DNA integrity by comet assay as described by Hughes et al. (36) and Tarozzi et al. (37). According to SAS (38), statistical analysis was conducted, followed by the Duncan test to detect important (p < 0.05) variations.

Issue:2, (2020)

ISSN: P-1999:6527 E-2707:0603

# **Results and discussion**

The values of fresh and post-thaw semen characteristics at different months were presented in table (1 and 2). Results revealed a significant decrease (P≤0.01) in plasma membrane integrity in September (51.8 $\pm$ 1.64 and 42.2 $\pm$ 1.87) as compared with December (62.6±2.36 and 61.4±2.28), January (66.2±2.36 and 56.3±2.35), February (63.3±1.81 and 55.8±2.38) and March (60.1±2.24 and 52.1±3.53) in fresh and frozen semen. This monthly variation might be due to heat stress that affect sperm cells lead to production of reactive oxygen species (ROS), this will affect Poly Unsaturated Fatty Acid (PUFA) on sperm plasma membrane and due to very low antioxidant enzymes in sperm cells makes it susceptible to oxidative stress leads to loses of plasma membrane fluidity that decreased sperm function and motility (39-41).

Table (1): Monthly variation in fresh semen characteristics of Friesian Holstein bulls (Mean  $\pm$  SE).

Months	Semen parameters (Mean ± S.F.			
	Plasma	Acrosome	DNA	
	Membrane	Integrity	integrity	
	integrity	%	%	
	%			
December	62.6±2.36	86.4±0.97	90.0±0.50	
	a	a	a	
January	66.2±2.36	88.3±1.14	88.3±0.88	
	a	a	а	
February	63.3±1.81	85.0±1.14	87.6±1.02	
	a	a	a	
March	60.1±2.24	82.5±1.70	87.8±1.13	
	a	a	a	
September	51.8±1.64	69.0±1.36	78.9±1.34	
	ь	ь	b	
Sig.	0.0075 **	0.0042 **	0.0374 *	
Means having with the different letters in same column differed significantly.				
* (P<0.05), ** (P<0.01),				

\* (P≤0.05), \*\* (P≤0.01).

Also, the acrosome integrity decreased significantly (P $\le$ 0.01) in September (69.0 $\pm$ 1.36 and 59.8 $\pm$ 1.72) compared to December (86.4 $\pm$ 0.97 and 80.1 $\pm$ 0.93), January (88.3 $\pm$ 1.14 and 79.0 $\pm$ 1.18), February (85.0 $\pm$ 1.14 and

Issue:2, (2020)

ISSN: P-1999:6527 E-2707:0603

 $76.7\pm1.31$ ) and March (82.5±1.70 and 73.3±1.88) in fresh and frozen semen, respectively. Al madaly et al. (42) suggested that heat stress causes an increase in abnormal spermatozoa and damaged acrosomes.

Furthermore, the DNA integrity also decline significantly (P < 0.05) in September  $(78.9\pm1.34)$ and  $75.8\pm1.24$ ) compared December (90.0 $\pm$ 0.50 and 83.9 $\pm$ 0.76), January  $(88.3\pm0.88 \text{ and } 82.1\pm1.01)$ , February  $(87.6\pm1.02)$ and 82.1±1.09) and March (87.8±1.13 and 82.1±1.41) in fresh and frozen semen, respectively. This variation in DNA damage might be due to heat stress. Borg et al. (43) stated that at the time of heat stress, spermatids present within the testis have a higher degree of DNA damages. When the ambient temperature continued to rise may leads to decreased blood flow and the testicular tissues becomes hypoxic lead to extreme production of ROS and the thermoregulatory mechanism disturbed (40,44).

#### Conclusion

Thus, it was concluded from this study that the September month adversely affect the various bio-physical characteristics of semen in Friesian Holstein bulls. December, January, February and March were the most favorable months for good quality semen production.

Table (2): Monthly variation in post-thaw semen characteristics of Friesian Holstein bulls (Mean  $\pm$  SE).

Months	Semen parameters (Mean ± S.E)			
	Plasma	Acrosome	DNA	
	Membrane	Integrity	integrity	
	integrity	%	0/0	
	%			
December	61.4±2.28	80.1±0.93	83.9±0.76	
	a	a	a	
January	56.3±2.35 ab	79.0±1.18	82.1±1.01	
January	30.3-2.33	a	ab	
February	55.8±2.38 ab	76.7±1.31	82.1±1.09	
		ab	ab	
March	52.1±3.53	73.3±1.88	82.1±1.41	
	b	ь	ab	
September	42.2±1.87	59.8±1.72	75.8±1.24	
	c	С	ь	
Sig.	0.0068 **	0.0001 **	0.0492*	
Means having with the different letters in same column differed significantly .				
* (P≤0.05), ** (P≤0.01).				

# References

- 1. Heise A. Artificial insemination in veterinary science. A Bird's-Eye View Vet Med. 2012;17-33.
- 2. Bryant J, López-Villalobos N, Holmes C, Pryce J. Simulation modelling of dairy cattle performance based on knowledge of genotype, environment and genotype by environment interactions: current status. Agric Syst. 2005;86(2):121-43.
- 3. Fuerst-Waltl B, Schwarzenbacher H, Perner C, Sölkner J. Effects of age and environmental

Issue:2, (2020)

ISSN: P-1999:6527 E-2707:0603

- factors on semen production and semen quality of Austrian Simmental bulls. Anim Reprod Sci. 2006;95(1–2):27–37.
- Hofírek B, Dvořák R, Němeček L, Doležel R, Pospíšil Z. Nemoci skotu. 1. vyd., Brno: Noviko as, Česká buiatrická společnost, 1149 s. 2009;
- 5. Beran J, Stádník L, Bezdíček J, Louda F, Čítek J, Ducháček J. Effect of sire and extender on sperm motility and share of live or dead sperm in bulls' fresh ejaculate and in AI doses after thawing. Arch Anim Breed. 2012;55(3):207–18.
- 6. Hafez ESE, Hafez B. Reproduction in farm animals. John Wiley & Sons; 2013.
- 7. Beran J, Stádník L, Ducháček J, Toušová R, Louda F, Štolc L. Effect of bulls' breed, age and body condition score on quantitative and qualitative traits of their semen. Acta Univ Agric Silvic Mendelianae Brun. 2014;59(6):37–44.
- 8. Stádník L, Rajmon R, Beran J, Šimoník O, Doležalová M, Šichtař J, et al. Influence of selected factors on bovine spermatozoa cold shock resistance. Acta Vet Brno. 2015;84(2):125–31.
- 9. AL-Badry KI. Effect of magnetically treated water on enzymes and total protein in seminal plasma of Holstein bulls born in Iraq. Iraqi J Vet Med (ISSN-P 1609-5693 ISSN-E 2410-7409). 2016;40(2):82–8.
- 10. Falah H, Al-Ghetaa K. Effect of Environmental High Temperature on the Reproductive activity of Awassi Ram Lambs. Iraqi J Vet Med. 2012;36(2):244–53.
- 11. Bhakat M, Mohanty TK, Gupta AK, Abdullah M. Effect of season on semen quality of crossbred (Karan Fries) bulls. Adv Anim Vet Sci. 2014;2(11):632–7.
- 12. Alragubi SM. Factors affecting on semen production in Friesian bulls in Libya. Int J Biopharm. 2015;6(1):32–6.
- 13. Ball PJH, Peters AR. Reproductive problems. Reprod Cattle. 2004;172–5.

- 14. Haugan T, Reksen O, Gröhn YT, Kommisrud E, Ropstad E, Sehested E. Seasonal effects of semen collection and artificial insemination on dairy cow conception. Anim Reprod Sci. 2005;90(1–2):57–71.
- 15. Argov-Argaman N, Mahgrefthe K, Zeron Y, Roth Z. Season-induced variation in lipid composition is associated with semen quality in Holstein bulls. Reproduction. 2013;145(5):479–89.
- 16. Snoj T, Kobal S, Majdic G. Effects of season, age, and breed on semen characteristics in different Bos taurus breeds in a 31-year retrospective study. Theriogenology. 2013;79(5):847–52.
- 17. Pileckas V, Siukscius A, Razamaite V. Survival Effect Of Keap Period On BullSemen Properties. Vet Irzootechnika. 2013;64(86):76–81.
- 18. Nardone A, Ronchi B, Lacetera N, Ranieri MS, Bernabucci U. Effects of climate changes on animal production and sustainability of livestock systems. Livest Sci. 2010;130(1–3):57–69.
- 19. Mathevon M, Buhr MM, Dekkers JCM. Environmental, Management, and Genetic Factors Affecting Semen Production in Holstein Bulls. J Dairy Sci [Internet]. 1998;81(12):3321–30. Available from: http://dx.doi.org/10.3168/jds.S0022-0302(98)75898-9
- 20. Nichi M, Bols PEJ, Züge RM, Barnabe VH, Goovaerts IGF, Barnabe RC, et al. Seasonal variation in semen quality in Bos indicus and Bos taurus bulls raised under tropical conditions. Theriogenology. 2006;66(4):822–8.
- 21. Hansen PJ. Exploitation of genetic and physiological determinants of embryonic resistance to elevated temperature to improve embryonic survival in dairy cattle during heat stress. Theriogenology. 2007;68:S242–9.
- 22. Garcia-Peniche TB, Cassell BG, Pearson RE, Misztal I. Comparisons of Holsteins with Brown Swiss and Jersey cows on the same farm for age at first calving and first calving

## **AL-ANBAR JOURNAL OF VETERINARY SCIENCES**

Research Article

Vol. 13

Issue:2, (2020)

ISSN: P-1999:6527 E-2707:0603

- interval. J Dairy Sci. 2005;88(2):790-6.
- 23. Dessouky F, Juma KH. Seasonal variation in semen characteristics of Friesian bulls in Iraq. J Agric Sci. 1968;71(1):37–40.
- 24. Juma KH, Dessouky F. Deterioration in some semen characters of Friesian bulls raised in Iraq. Trop Agric. 1969;46(1):63.
- 25. Al-Janabi AS, Abdul-Latif BM, Al-Badry KI, Ibrahim FF. Seasonal changes in certain seminal attributes of Friesian bulls in Iraq. Bulg J Agric Sci. 1999;5(5):797–800.
- 26. Al-badry KI. Monthly Changes in Libido and Semen Characteristics for Holstein Bulls Born in Iraq of Different Reproductive. 2013;2(1):67–74.
- 27. Al-Badrany QM, Al-Badry KI, Zalzala SJ, Ibrahim FF, Lateef WY. Effect of different concentration of trehalose or sucrose to tris diluents on some properties of sperms for holstein bulls during dilution, cooling and freezing. Plant Arch. 2020;20(April):1953–60.
- 28. Ramu S, Jeyendran RS. The hypo-osmotic swelling test for evaluation of sperm membrane integrity. In: Spermatogenesis. Springer; 2013. p. 21–5.
- 29. Zubair M, Ahmad M, Jamil H. Review on the screening of semen by hypo-osmotic swelling test. Andrologia. 2015;47(7):744–50.
- 30. Neild DN, Gadella BM, Agüero A, Stout TAE, Colenbrander B. Capacitation, acrosome function and chromatin structure in stallion sperm. Anim Reprod Sci. 2005;89(1–4):47–56.
- 31. Esteves SC, Sharma RK, Thomas AJ, Agarwal A. Evaluation of acrosomal status and sperm viability in fresh and cryopreserved specimens by the use of fluorescent peanut agglutinin lectin in conjunction with hyposmotic swelling test. Int Braz J Urol. 2007;33(3):364–76.
- 32. Balić IM, Milinković-Tur S, Samardžija M, Vince S. Effect of age and environmental factors on semen quality, glutathione peroxidase activity and oxidative parameters

- in simmental bulls. Theriogenology. 2012;78(2):423–31.
- 33. Agarwal A, Nallella KP, Allamaneni SSR, Said TM. Role of antioxidants in treatment of male infertility: An overview of the literature. Reprod Biomed Online [Internet]. 2004;8(6):616–27. Available from: http://dx.doi.org/10.1016/S1472-6483(10)61641-0
- 34. El-Sheshtawy RI, Sisy GA, El-Nattat WS. Effects of different concentrations of sucrose or trehalose on the post-thawing quality of cattle bull semen. Asian Pacific J Reprod. 2015;4(1):26–31.
- 35. Kovacs A, Foote RH. Viability and acrosome staining of bull, boar and rabbit spermatozoa. Biotech Histochem. 1992;67(3):119–24.
- 36. Hughes CM, Lewis SEM, McKelvey-Martin VJ, Thompson W. Reproducibility of human sperm DNA measurements using the alkaline single cell gel electrophoresis assay. Mutat Res Mol Mech Mutagen. 1997;374(2):261–8.
- 37. Tarozzi N, Bizzaro D, Flamigni C, Borini A. Clinical relevance of sperm DNA damage in assisted reproduction. Reprod Biomed Online. 2007;14(6):746–57.
- 38. SAS JMP. Statistical Analysis System, v. 10.0. 2. Cary, North Carolina USA. 2012;
- 39. Mandal DK, Nagpaul PK, Gupta AK. Motion characteristics of Murrah buffalo bull spermatozoa in various seasons and its relationship with functional integrity of the plasmallema.

  Theriogenology. 2003;60(2):349–58.
- 40. Nichi M, Bols PEJ, Züge RM, Barnabe VH, Goovaerts IGF, Barnabe RC, et al. Seasonal variation in semen quality in Bos indicus and Bos taurus bulls raised under tropical conditions. Theriogenology. 2006;66(4):822–8.
- 41. Kothari S, Thompson A, Agarwal A, du Plessis SS. Free radicals: Their beneficial and detrimental effects on sperm function. Indian J Exp Biol. 2010;48(5):425–35.

# **AL-ANBAR JOURNAL OF VETERINARY SCIENCES**

Research Article

Vol. 13 Issue:2, (2020)

ISSN: P-1999:6527 E-2707:0603

- 42. Almadaly E, Farrag F, Shukry M, Murase T. Plasma Membrane Integrity and Morphology of Frozen-Thawed Bull Spermatozoa Supplemented with Desalted and Lyophilized Seminal Plasma. 2014;13(5):753–66.
- 43. Borg KE, Lunstra DD, Christenson RK. Semen characteristics, testicular size, and reproductive hormone concentrations in mature Duroc, Meishan, Fengjing, and Minzhu boars. Biol Reprod. 1993;49(3):515–21.
- 44. Trisini AT, Singh NP, Duty SM, Hauser R. Relationship between human semen parameters and deoxyribonucleic acid damage assessed by the neutral comet assay. Fertil Steril. 2004;82(6):1623–32.