

### BASRAH JOURNAL OF VETERINARY RESEARCH, 2023, 22(4):95-109 https://bjvr.uobasrah.edu.iq/

### Methicillin resistance *Staphylococcus aureus* (MRSA) and Vancomycin Resistant *Staphylococcus aureus* (VRSA) problem in human and livestock and solutions

Marwan M. Mohammed <sup>1</sup>, Mohammed H. Khudor<sup>2</sup> Hanaa K. Ibraheim <sup>2</sup>.

- 1 Department of Medical laboratory Technologies, Basrah college of sciences and technology.
- 2 Department of Microbiology, College of Veterinary Medicine, University of Basrah, Iraq.

Corresponding Author Email Address: marwan ismailiya@yahoo.com

**ORCID ID:** https://orcid.org/0009-0006-6259-5224

**DOI:** <u>10.23975/bjvetr.2023.142835.1042</u>

Received: 23 August Accepted: 7 November 2023.

#### Abstract

Staphylococcus aureus is an opportunistic bacterium of humans and other mammals that is becoming more clinically and veterinary important due to its fast development of antibiotic resistance. Some of these S. aureus varieties are methicillin-resistant S. aureus (MRSA), which is common in healthcare organizations, community settings, and livestock farms across the world. Beyond humans, MRSA has the potential to live in other animal species, which could result in the emergence and spread of antimicrobial agent resistance in various animal species. Vancomycin is a type of antibiotic classified as a final resort option, employed to address severe infections instigated by Gram-positive bacteria. Vancomycin remains among the primary choices for the initial treatment of MRSA infections. During the past few years, there has been an emergence of Staphylococcus aureus strains exhibiting strong resistance to vancomycin. The vanA gene cluster, obtained from vancomycin-resistant enterococcus, eases the transfer of vancomycin resistance in S. aureus. In terms of likely transfer routes, underlying mechanisms, and consequences of methicillin and vancomycin resistance from animals to humans and vice versa, this review aims to highlight the Methicillin resistance Staphylococcus aureus (MRSA) and vancomycin resistance Staphylococcus aureus (VRSA) issues

**Key words:** *Staphylococcus aureus*, MRSA, VRSA, human, animal.

### Introduction

### Staphylococcus

Staphylococcus is a genus of the Grampositive cocci family staphylococcaceae, which contains more than 30 species of coagulase-positive and coagulase -negative staphylococci that culminate in a range of clinical symptoms. Staphylococcus aureus (S. aureus) is the most clinically significant opportunistic species of pathogen, colonizing between 30% and 50% of the human population as part of the typical commensal flora of humans and livestock (1). S. aureus is one of the principal organisms in hospital and community infections and can cause numerous infectious disorders, such as minor skin and soft tissue infections. Bacteremia, deadly pneumonia, osteomyelitis and infectious endocarditis (2) . Alexander Ogston, a surgeon in Aberdeen, Scotland, made the initial discovery of Staphylococcus aureus in 1880 while managing patients with open wounds. Under microscope, a Staphylococcus aureus appears structured like a "string of grapes" and is an element of the Staphylococcus genus of Firmicutes. It is Gram-positive, grows best around 37 °C, and has a pH of 7.4 (3). Staphylococcus aureus has a wide range of metabolic capabilities and pharmic resistance, which let it adapt to a variety of situations. S. aureus colonizes the epidermis nasopharyngeal membranes of around 25-30% of healthy people; this bacterium is a natural component of the human microbiota and does not harm individuals (4). S. aureus can result in a wide range of dangerous illnesses in people and cattle, from severe

systemic infections to minor skin and soft tissue infections. (2). Six biotypes of S. aureus have been identified: human, hemolytic human, bovine, caprine, avianabattoir, and non-host specific. Animal isolates of S. aureus have been found to have distinctive phenotypic characteristics that differ depending on the host of origin (5). These biotypes have, for the most part, weathered the use of advanced characterization techniques; isolates from various hosts that are identified by multilocus enzyme electrophoresis group together (2). An arsenal of different toxins and virulence factors produced by S. aureus are thought to be responsible for its pathogenesis (6). According to the WHO's most recent global priority pathogens list (global PPL) of antibiotic-resistant bacteria, S. aureus is one of the most clinically significant multidrug-resistant globally (7). Second on the priority list of the bacteria for the development of new antibiotics are methicillin-resistant S. aureus (MRSA), vancomycin-intermediate, resistant S. aureus strains (8). This paper focus on MRSA & VRSA issues in human and animals with suggestion for solution.

### Clinical significant of S. aureus

S. aureus infections can range from simple, superficial skin infections harmful. fundamental illnesses like. osteomyelitis and pneumonia that can enter the bloodstream and create septicemia in human. (9). S. aureus causes various infections in human, including superficial skin diseases, surgical and trauma wound infections, urinary tract infections, and infections pneumonia, organ like

osteomyelitis, endocarditis. phlebitis, mastitis, and meningitis. People with chronic illnesses. diabetes. traumatic injuries, burns, or immunosuppression are more susceptible to these infections (10). Hospital infections caused by S. aureus, particularly is currently recognized as the common cause of most infective endocarditis in the industrialized world. have generated serious concerns in addition to food-borne diseases (11). S. aureus has been discovered in a variety of human physiological circumstances, including livestock, pets, food, and manufacturing processes. Since 1884, consuming tainted cheese has resulted in food-borne diseases. (12). A decade later, it was determined that a family experienced health issues brought on by eating meat from a cow that had passed away from a pyogenic staphylococci fever (8). Additional cases of food poisoning attributed to enterotoxin-producing aureus strains have been documented (7). As a result of these outbreaks, inspecting food items and production processes for bacterial contamination, particularly S. aureus, is more important than ever. (9).

The pathogen has also infected livestock resulting in the contamination of food products. This was demonstrated when a mastitis epidemic was noticed in a dairy herd that was very congested and was later determined to be brought on by a specific strain of *S. aureus* (13). In an examination of a variety of species of animals, including cows, goats, lambs, rabbits, chickens, and cats, *S. aureus* isolates carrying different staphylococcal enterotoxin genes was found. The incidence of outbreaks and the discovery of new genes predominating in *S.* 

aureus from animal hosts show that cattle constitute maior reservoir staphylococcal infections (14). S. aureus is a challenging microorganism with capacity to override humoral immune responses, release proteins that neutralize antimicrobial peptides, and avoid immune system (15).To 1imit prevalence incidence and of highly transmissible strains of the bacteria, strict cleanliness during milking, segregation of any cattle with S. aureus infections, and extensive culling of those infected might prove necessary (16). Regardless of the host, S. aureus isolates produce a variety of toxins that enable the bacterium to be a seriously dangerous pathogen (17).

### **Staphylococcal Toxins**

When microbes make direct contact with a host, they discharge toxins, which are harmful substances. Panton-Valentine leukocidin (PVL), a notable toxin generated by S. aureus, is connected with numerous severe infections, which has been linked to a substantial number of S. aureus isolates that result in necrotic skin lesions, severe necrotizing pneumonia, and white blood cell destruction (10). To understand what makes S. aureus isolates with PVL positivity, several research was carried out. In healthy children and young adults, PVL-producing S. aureus strains manufactured. hemorrhagic, necrotizing pneumonia with a high mortality rate (18). Another toxin generated by S. aureus, identified as an alpha toxin ( $\alpha$ -toxin), plays a role in its pathogenesis as a virulence factor because it causes tissue necrosis and invasion while modifying macrophages' ability to eliminate bacteria (19).The alpha-toxin discovered to be a key component of the pathogenicity of opportunistic bacterial lung infections, where it significantly boosted bacterial growth and inhibited acidification of bacteria-containing macrophages (phagosomes), lowering the efficiency of S. aureus destruction (20). Additional toxins are involved in the pathogenesis of S. aureus in addition to the main toxins. Food poisoning and toxic shock are caused whenever the pyrogenic-toxin superantigens bind to the major histocompatibility complex II protein. This promotes substantial T-cell proliferation and the release of cytokines (18). The pathogen also makes a variety of other enzymes, such as proteases, lipases, and hyaluronidases, which break down host tissue and can assist in an infection spread through different enzyme beta-lactamase tissues. The inactivating penicillin (PBP2a), in particular, results in S. aureus that is resistant to Transpeptidase antibiotics, penicillinbinding proteins (PBPs) that Staphylococci manufacture decrease the action of -lactam antibiotics. (18).Reduced-affinity penicillin-binding protein 2a (PBP2a), which is produced by the mecA gene, was first discovered in 1981. Despite the presence of fatal doses of methicillin, the creation of the peptidoglycan cell wall of MRSA was allowed to continue because of the poor binding affinity of this PBP2a to lactam antibiotics (12). It's interesting to note that a variety of virulence factors, such as toxins, cell-surface-associated adhesions, and secreted exo-proteins, are necessary for the pathogenesis of livestock-associated S.

aureus (LA-SA), which is comparable to human-associated *S. aureus* (HA-SA) (21).

### **Antimicrobial resistance**

Antibiotics are medications used to suppress or eliminate bacterial development, produced by living cells, by preventing the growth of challenging bacteria (22). However, more lately, Antimicrobial have been referred to be antibiotics that are also created entirely or in portion using synthetic methods (23). Several antibiotics have been approved for use in both humans and animals, categorized into five groups based on their actions (24):

- 1. Beta-lactam antibiotics (penicillins, cephalosproins, carbapenems, and monobactams) and vancomycin are examples of cell wall synthesis inhibitors with a bactericidal effect.
- 2. Protein synthesis inhibitors involve antibiotics that bind to either the 30S or 50S ribosomal subunits of bacteria (tetracyclines and aminoglycosides, for example).
- 3. The folate pathway inhibitors that hinder the synthesis of folic acid.
- 4. The DNA breakage-reunion stepinterrupting quinolones, the RNA polymerase activity-inhibiting rifampicin, the DNA-gyrase or topoisomerase II and topoisomerase IV-binding inhibiting agents of nucleic acid synthesis.
- 5. The polymers and polypeptides that hinder the membrane of cells permeability.

The healthcare system faces challenges due to overuse and negligent prescribing of antibiotics, often for viral illnesses that cure on their own. (17). Research indicates a lack understanding about responsible of antibiotic use and poor prescribing practices, leading to widespread, severe consequences in many nations. (16). Antibiotic resistance increases with increased usage, primarily due to novel selection, gene epidemics, and strain epidemics. These are caused by promoting entire species, mutations, and transferring resistance genes to mobile DNA (21). Antimicrobial resistance is a growing global concern, particularly in Europe, where it is becoming a significant public health issue due to the increasing use of broad-spectrum antibiotics. (25). Between 2000 and 2010, the global consumption of antibiotics increased significantly (35%) with 76% of the growth coming from Brazil, Russia, India, China, and South Africa (25). Multiple resistant proportions to various antibiotics were observed, including 87.1% of Enterococcus faecium to vancomycin, Staphylococcus 56.8% of aureus to oxacillin-methicillin, 39.7% ofStaphylococcus aureus to clindamycin, 32.6% of Pseudomonas aeruginosa to fluoroquinolones, 31.3% of E. faecium to daptomycin (26).

Another significant factor contributing to antibiotic resistance, besides patients and doctors using medicines inappropriately, was the use of antimicrobials in animals (27). The use of these antimicrobial drugs, particularly antibiotics, is widespread utilized in livestock production systems to protect their health and enhance their growth performance (28). Antimicrobial compounds in animal feed help reduce costs of meat, milk, eggs, and

animal products for human other consumption, meeting the world's nutritional needs. (29). 78% of antibiotics in the USA are used on animals for food production, potentially leading to inaccurate use as preventative measures and nontherapeutic Antibiotic resistance will uses. (30). propagate across different cultures of bacteria when healthy animals are exposed to antibiotics for an extended period of period. It's possible that antimicrobial resistance cannot entirely be prevented and avoided.

## Methicillin-Resistant Staphylococcus aureus (MRSA)

Fleming launched the era of antibiotics for the management of infections in the 1940s with the discovery of penicillin (Figure 1). S. aureus-related infectious illnesses were under control at the time, but as penicillin became more widely used in the 1950s, penicillin-resistant S. aureus started growing up in medical setting (31). Methicillin was used in the clinic in 1959 and was effective in managing penicillin-resistant S. aureus infections (32). Only two years after the use of methicillin, in 1961, British scientist Jevons reported the isolation of an MRSA strain. This resistance was triggered by a gene (mecA) containing the penicillinbinding protein 2a or 2' (PBP2a or PBP2') which was integrated into the chromosomal element (SCCmec) of methicillin-sensitive S. aureus (33). Additionally, MRSA is growing rapidly as the most prevalent resistant infection found around the world, including Europe, the United States, North Africa, the Middle East, and East Asia (34). MRSA is divided into two categories,

community-acquired MRSA (CAMRSA) and hospital-acquired MRSA (HA-MRSA), according to its initial source (34). Livestock associated MRSA (LA-MRSA) was first described in 2005 (35). The fatality rate of

MRSA infection exceeds that of AIDS and Parkinson's disease, according to the Centers for Disease Control (CDC) in the US (34).

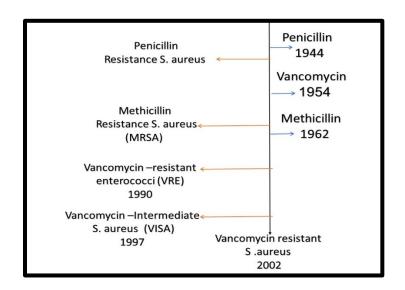


Figure 1: Demonstration the evolution of drug resistance in *S. aureus*.

### Methicillin-Resistant and Beta-Lactamase mechanisms

Methicillin resistance is caused by the creation of a novel penicillin-binding protein (PBP) called PBP2a, which has low affinity for methicillin along with other Betalactams and precludes the antibiotic from accessing its target site, leading to resistance. Methicillin resistance is mostly caused by the mecA gene (32,36). The genomic region known as staphylococcal cassette chromosome тес (SCCmec) encodes this gene. Two genes, mecR1, which controls transcription, and mecI, which encodes the repressor protein, are required for the expression of the mecA gene. The most frequent PBP2a-encoding gene is the *mecA* gene; nevertheless, a novel mec gene has just been discovered. *mec C*, a component of the SCCmec type XI, has been found in samples from mammals and the environment in both *S. aureus* and CoNS. Although the *mec B* gene has just been discovered in *S. aureus*, two fewer *mec* genes (33).

Beta -lactamase is a transferring enzyme that is conveyed by bacterial chromosomal genes that hydrolyzes an assortment of -lactam drugs, including broad-spectrum antibiotics like carbapenem (37). The studies revealed that beta-lactam antibiotics primarily eliminate bacteria through two main pathways: first, by binding to penicillin-binding protein (PBPs), which inhibits the

production of cell wall mucin synthase, disrupts the cell wall, and causes bacterial expansion and lysis; and second, by inducing the activity of the bacteria's autolytic enzyme, which causes autolysis and death (34). Antibiotic effectiveness is primarily decreased by excessive MRSA Beta-lactamase release via two processes, which results in MRSA resistance. The first is the hydrolysis process, in which Betalactam antibiotics were hydrolized and rendered inactive by Beta-lactamase; the second is the pinching mechanism, in which a significant concentration of -lactamase attaches quickly and permanently to target molecules. Two important processes through which MRSA's excessive -lactamase impact secretion diminishes the antibiotics and results in extracellular drugs that are resistant to MRSA antibiotics are unable to enter the intracellular space and so cannot reach the target site (34).

MRSA may be eradicated from hospitals by enforcing tight cleanliness environmental controls. In several countries. search and destroy tactics were used to reduce the spread of both HA- and CA-MRSA in healthcare settings (38). Patients and health-care personnel are screened for MRSA transmission in these techniques. Following that, all MRSA-positive persons are separated and treated, wherever possible in order to eradicate MRSA carriage. However, since 2012, the rate of reduction in HA-MRSA cases has slowed (3). MRSA may infiltrate a farm through a variety of means. The most serious of these include the migration of MRSA-infected animals from one farm to another, direct contact with infected humans, and animal interaction

with contaminated transport vehicles (39). During quarantine, animals should be checked for MRSA and only permitted to rejoin the main herd if they are MRSA-negative, before handling animals' farms should use a minimum of antimicrobials as possible. When MRSA first enters a farm, antibiotic usage aids in the selection of MRSA (40).

### Vancomycin discovery and action mechanism

One of the earliest antibiotics, vancomycin has been utilized in healthcare for more than sixty years. In the tropical rainforests of Borneo, in 1957, Dr. Kornield, an organic for Eli Lilly. chemist discovered vancomycin from Streptomyces orientalis (3). Vancomycin acts successfully against Gram-positive bacteria such as Listeria, Corynebacterium, Staphylococci, Enterococci, Pneumococci, and Streptococci. Vancomycin is currently used to treat individuals who suffer from allergies semi-synthetic penicillin or cephalosporins in addition to infections caused by MRSA (7). Its use was restricted due to the presence of chemicals that produced toxicities in previous formulations (3). Vancomycin's use grew starting in the 1980s when purer compositions were created in the late 1970s, but it was reconsidered after methicillin-resistant Staphylococci emerged in the 1970s (3). Vancomycin has supplanted conventional injectable medications as the preferred option for treating methicillin-resistant Staphylococcus aureus and drug-resistant Enterococcus species (41). Vancomycin eliminates bacteria by preventing the correct creation of their cell walls in receptive bacteria. A cell wall structure that coats the majority of bacterial membranes prevents cells from growing and bursting owing to intracellular excessive osmolality (42). The peptidoglycan-containing cell wall structure has to be reinforced. Penicillin-binding proteins (PBPs) transglycosylate transpeptide the precursor lipid II in order to incorporate it into the developing peptidoglycan. Vancomycin's hydrophilic molecule can engage via hydrogen bonds with the precursor lipid II's terminal Dalanyl-D-alanine (D-Ala-D-Ala) moieties. Vancomycin binding causes conformational change that precludes the inclusion of the precursor to the developing peptidoglycan chain and the subsequent transpeptidation, resulting in bacterial lysis and cell wall breakdown (43. Vancomycin's intricate structure prevents it from traversing Gram-negative bacteria's outermost membrane, and it barely has a moderate bactericidal effect on bacteria (44).

The clinical and laboratory standards Institute divides the *S. aureus* isolates into three categories in accordance with their lower susceptibility to vancomycin. The three variants are vancomycin-susceptible *S. aureus* (VSSA) with MIC 2 μg/ml, vancomycin-intermediate *S. aureus* (VISA) with MIC 4-8 μg/ml, and VRSA with MIC 16 μg/m (43). Molecular techniques should be used to establish the presence of *vanA* or other van resistance determinants to determine when an isolate belongs to VRSA (43).

The heterogeneous vancomycinintermediate S. aureus (hVISA), which is referred to as a *S. aureus* strain with a vancomycin MIC within the susceptible range (2  $\mu$ g/ml) identified using conventional methods, and a cell subpopulation is in the vancomycinintermediate range (4  $\mu$ g/ml), has been proposed to be the origin of VISA strains (44).

VISA was believed to be generated by the progressive mutation accumulation of VISA-associated genes. Particularly significant are the genes for several-component regulatory systems, including WalKR, GraSR, and VraSR (43). Although their genetic lineages differed, VISA strains shared common phenotypes such as thickened cell walls, reduced autolytic activity, and decreased virulence (44).

The first vancomycin-resistant enterococci (VRE) were discovered in Europe and quickly spread to health care medical units. Vancomycin resistance in VRE mediated by transposons determined mostly on plasmids, raising significant concerns about the possibility of vancomycin-resistant determinants spreading to generally susceptible bacteria of medical interest, particularly S. aureus. The effective transfer of the van element from Enterococcus faecalis to an MRSA strain in mix-infected rats validated this concern (46). In 2002, The first VRSA strain has been identified in Michigan, the second in Pennsylvania, and a total of 52 VRSA strains carrying van genes have since been noticed, including 14 isolated in the USA, 16 in India, 11 in Iran, 9 in Pakistan, 1 in Brazil, and 1 in Portugal (46).

### Mechanism of VRSA resistance

Vancomycin resistance is classified into several gene clusters based on the DNA sequence of the ligase van gene homologues that encode the key enzyme for the synthesis of D-alanyl-D-lactate (D-Ala-D-Lac) or Dalanyl-D-serine (D-Ala-D-Ser). At least 11 van gene clusters confer vancomycin resistance, responding for VanA, VanB, VanD, VanF, VanI, VanM, and VanN phenotypes (47. Genes encoding D-Ala:D-Lac ligases, such as vanA, vanB, vanD, vanF, vanI, and vanM 9Figure 2), frequently result in high-level vancomycin resistance with MICs higher than 256 mg/ml, while genes encoding D-Ala:D-Ser ligases, such as vanC, vanE, vanG, vanL, and vanN, generally result in low-level resistance with MICs of 8-16 mg/ml (48).

### **Control of Resistance for Vancomycin**

The development of preventive measures to control current resistance and prevent the development of resistant bacteria is critical for preserving the efficacy of antibiotics in both human and veterinary medicine. Studying the epidemiology of antibiotic resistance will allow us to devise preventative strategies to reduce current resistance and prevent the introduction of new strains of resistant bacteria (49. Hvgiene methods to avoid cross contamination are used to control the emergence of resistance. and a reduction in antibiotic use are desirable. Vancomycin use in animals should be limited to diseases that respond just to vancomycin and for which no other plausible options are available; when used in animals, it should be administered at the

right dosage, dosing interval, and treatment duration (44).

### **Conclusions**

Staphylococcus aureus is a bacterium that is found everywhere and is frequent in humans and animals. S. aureus may cause infections in hospitals and communities and has become the most common pathogen in hospitals across the world. The widespread use of antibiotics has resulted in an increase in bacterial resistance, beginning with the emergence of multidrug resistant strains such as MRSA, which has been regarded as a clinically important problem and has attracted extensive attention from domestic and international research. MRSA has become an obstacle in clinical therapy due to its features of simple infection, high mortality, and antibiotic resistance. Modern research has concentrated on how to successfully prevent and control MRSA. Science and technology have advanced time. medicine throughout and has continued to evolve.

For decades, vancomycin has been an effective treatment for MRSA infections. It is expected to maintain dominance as long as vancomycin resistance is under control and new antibiotics with higher performance are not available. Although the number of cases of VRSA infection is small, VRSA remains a potential hazard to public health. In health-care settings, intensive surveillance of vancomycin resistance, correct antibiotic usage, and adherence to infection control standards are critical.

The greatest strategy to control resistance in big groups of animals is to reduce the demand for antibiotics. This can be achieved through proper vaccination against infectious diseases, the adoption of good hygienic practices in animal husbandry, the discontinuation of the use of antibiotics as feed additives for growth promotion in animals bred for food, the appropriate use of antibiotics for food animals, and the development of guidelines, codes of practice, and policies on the appropriate use of antibiotics.

**Conflict of Interest**: The authors revealed that there is no conflict of interest.

### References

- 1.Al-Mussawi, A. A. (2016). Detection of Biofilm Coding icaD Gene in Methicillin Resistant *Staphylococcus aureus* (MRSA) Isolated from Patients undergoing Tonsillectomy in Basra City. *International Journal of Current Microbiology and Applied Sciences*, 5(6):1171–7.
- 2.Asgin, N. & Otlu, B. (2020). Antibiotic resistance and molecular epidemiology of vancomycin-resistant enterococci in a tertiary care hospital in Turkey. *Infection and Drug Resistance*, 13(2) 356–8.
- 3. Chambers, H. F. & Deleo, F. R. (2009). Waves of resistance: *Staphylococcus aureus* in the antibiotic era. *Nature Reviews Microbiology* . 7 (9):84–92.
- 4. Chatzopoulou, M. & Reynolds, L. (2020). Role of antimicrobial restrictions in bacterial resistance control: a systematic literature review. *Journal of Hospital Infection* . 104, (2):44–51.

- 5. Chatzopoulou, M. & Reynolds, L. (2020). Role of antimicrobial restrictions in bacterial resistance control: a systematic literature review. *Journal of Hospital Infection*. 104 (2): 2797–807.
- 6. de Jong, N. W. M., van Kessel, K. P. M. & van Strijp, J. A. G. (2019). Immune Evasion by *Staphylococcus aureus*. *Microbiology Spectrum*. 7(2):65–72.
- 7.Denayer, S., Delbrassinne, L., Nia, Y. & Botteldoorn, N. (2017). Food-borne outbreak investigation and molecular typing: High diversity of *Staphylococcus aureus* strains and importance of toxin detection. *Toxins*. 9(12):20–26.
- 8.Deng, Y., Liu, Y., Jiang, Z., Wang, J., Zhang, Q., Qian, Y., Yuan, Y., Zhou, X., Fan, G. & Li, Y. (2019). A multiplex loop-mediated isothermal amplification assay for rapid detection of Bacillus cereus and *Staphylococcus aureus*. *BioScience Trends*. *13*(6): 61-68.
- 9.DuPont, H. L. & Steele, J. H. (1987). The human health implication of the use of antimicrobial agents in animal feeds. *The Veterinary quarterly* . 9 (4): 261-267.
- 10.Gerada, C. & Ryan, K. M. (2020). Autophagy, the innate immune response and cancer. *Molecular Oncology* .14(9): 34-39.
- 11.González-Martín, M., Corbera, J. A., Suárez-Bonnet, A. & Tejedor-Junco, M. T. (2020). Virulence factors in coagulase-positive staphylococci of veterinary interest other than *Staphylococcus aureus*. *Veterinary Quarterly*. 40(1) 102-111.

- 12.Grima, L. Y. W., Leliso, S. A., Bulto, A. O. & Ashenafi, D. (2021). Isolation, Identification, and Antimicrobial Susceptibility Profiles of *Staphylococcus aureus* from Clinical Mastitis in Sebeta Town Dairy Farms. *Veterinary Medicine International*. 113(8): 87-91.
- 13. Khudor, M.H. (2010). The influence of heavy metals and antimicrobial on *Staphylococcus aureus* and *Pseudomonas aeroginosa* isolates. *Basrah Journal of Veterinary Research*. 9(2); 22-38.
- 14. Khudor. M. H., Basil ,B.A. and Idbeis, H. I.(2012) Detection of enterotoxin genes of *Staphylococcus aureus* isolates from raw milk. *Basrah Journal of Veterinary Research*. *11*(1): 254-264.
- 15.Hoque, M. N., Das, Z. C., Rahman, A. N. M. A., Haider, M. G. & Islam, M. A. (2018). Molecular characterization of *Staphylococcus aureus* strains in bovine mastitis milk in Bangladesh. *International Journal of Veterinary Science and Medicine*. 6(1):113-119.
- 16.Houri, H., Samadpanah, M., Tayebi, Z., Norouzzadeh, R., Malekabad, E. S. & Dadashi, A. R. (2020). Investigating the toxin profiles and clinically relevant antibiotic resistance genes among *Staphylococcus aureus* isolates using multiplex-PCR assay in Tehran, Iran. *Gene Reports*, 19 (5):34-37.
- 17.Hu, Q., Peng, H. & Rao, X. (2016). Molecular events for promotion of vancomycin resistance in vancomycin intermediate *Staphylococcus aureus*. *Frontiers in Microbiology*. *12* (7): :89–94.

- 18.Ibrahim, M. A. A., Abdeljawaad, K. A. A., Abdelrahman, A. H. M., Alzahrani, O. R., Alshabrmi, F. M., Khalaf, E., Moustafa, M. F., Alrumaihi, F., Allemailem, K. S., Soliman, M. E. S., Paré, P. W., Hegazy, M. E. F. & Atia, M. A. M. (2021). Non-β-lactam allosteric inhibitors target methicillin-resistant *Staphylococcus aureus*: An in silico drug discovery study. *Antibiotics.* 10(8): 73-80.
- 19.Imanishi, I., Nicolas, A., Caetano, A. C. B., Castro, T. L. de P., Tartaglia, N. R., Mariutti, R., Guédon, E., Even, S., Berkova, N., Arni, R. K., Seyffert, N., Azevedo, V., Nishifuji, K. & Le Loir, Y. (2019). Exfoliative toxin E, a new *Staphylococcus aureus* virulence factor with host-specific activity. *Scientific Reports*. *9*(1):132-138.
- 20.Iseppi, R., Di Cerbo, A., Messi, P. & Sabia, C. (2020). Antibiotic resistance and virulence traits in vancomycin-resistant enterococci (Vre) and extended-spectrum β-lactamase/ampc-producing (ESBL/ampc) enterobacteriaceae from humans and pets. *Antibiotics.* 9(4):223-231.
- 21. Jackson, K. A., Gokhale, R. H., Nadle, J., Ray, S. M., Dumyati, G., Schaffner, W., Ham, D. C., Magill, S. S., Lynfield, R. & See, I. (2020). Public Health Importance of Invasive Methicillin-sensitive Staphylococcus aureus Infections: Surveillance in 8 US Counties, 2016. Clinical Infectious Diseases. 70(6):88-96.
- 22. Jokinen, E., Laine, J., Huttunen, R., Rahikka, P., Huhtala, H., Vuento, R., Vuopio, J. & Syrjänen, J. (2017). Comparison of outcome and clinical characteristics of bacteremia caused by

methicillin-resistant, penicillin-resistant and penicillin-susceptible *Staphylococcus aureus* strains. *Infectious Diseases*. 49(7): 168-174.

23.Kashid, M. N., Kiwi-Minsker, L., Gehr, R., Chen, D., Moreau, M., Kikutani, Y., Horiuchi, T., Uchiyama, K., Hisamoto, H., Tokeshi, M., Kitamori, T., ZHAO, X., ZHANG, T., Zhou, Y., LIU, D. D., Ebrahimi, F., Kolehmainen, E., Oinas, P., Hietapelto, V., and Sin, Y. K. (2011). Peracetic Acid (CAS No. 79-21-0) and its Equilibrium Solutions. *Chemical Engineering Science*. 2(1): 236-242.

24.Kulhankova, K., Kinney, K. J., Stach, J. M., Gourronc, F. A., Grumbach, I. M., Klingelhutz, A. J. & Salgado-Pabón, W. (2018). The superantigen toxic shock syndrome toxin 1 alters human aortic endothelial cell function. *Infection and Immunity*, 86(3): 84-89.

25.Kumar, P. (2020). A review on quinoline derivatives as anti-methicillin resistant *Staphylococcus aureus* (MRSA) agents. *BMC Chemistry*. *14* (1): 141-148.

26.Lessa, F. C., Mu, Y., Ray, S. M., Dumyati, G., Bulens, S., Gorwitz, R. J., Fosheim, G., Devries, A. S., Schaffner, W., Nadle, J., Gershman, K. & Fridkin, S. K. (2012). Impact of USA300 methicillinresistant *Staphylococcus aureus* on clinical outcomes of patients with pneumonia or central line-associated bloodstream infections. *Clinical Infectious Diseases*. 55 (2): 315-328.

27.Liu, X., Zhang, Y., Li, Z., Zhang, P., Sun, Y. J. & Wu, Y. J. (2021). Paeoniflorin Derivative in Paeoniae Radix Aqueous

Extract Suppresses Alpha-Toxin of Staphylococcus aureus. Frontiers in Microbiology, (12):66-78.

28.Loubet, P., Ranfaing, J., Dinh, A., Dunyach-Remy, C., Bernard, L., Bruyère, F., Lavigne, J. P. & Sotto, A. (2020). Alternative Therapeutic Options to Antibiotics for the Treatment of Urinary Tract Infections. *Frontiers in Microbiology*. (11):187-198.

29.MacDougall, C., Johnstone, J., Prematunge, C., Adomako, K., Nadolny, E., Truong, E., Saedi, A., Garber, G. & Sander, B. (2020). Economic evaluation of vancomycin-resistant enterococci (VRE) control practices: a systematic review. *Journal of Hospital Infection* .105 (1):198-214.

30. Magiorakos, A. P., Srinivasan, A., Carey, R. B., Carmeli, Y., Falagas, M. E., Giske, C. G., Harbarth, S., Hindler, J. F., Kahlmeter, G., Olsson-Liljequist, B., Paterson, D. L., Rice, L. B., Stelling, J., Struelens, M. J., Vatopoulos, A., Weber, J. T. & Monnet, D. L. (2012). Multidrug-resistant, extensively drug-resistant pandrug-resistant and bacteria: An international expert proposal for interim standard definitions for acquired resistance. Clinical Microbiology and Infection. 18(3):316-328.

31.Matsuo, M., Yamamoto, N., Hishinuma, T. & Hiramatsu, K. (2019). Identification of a Novel Gene Associated with High-Level - Lactam Resistance in Heterogeneous Vancomycin-Intermediate *Staphylococcus aureus* Strain Mu3 and Methicillin-Resistant *S. Aureus* Strain N315. *Antimicrobial Agents and Chemotherapy*. 63(2):423-435.

- 32.Mdegela, R. H., Mwakapeje, E. R., Rubegwa, B., Gebeyehu, D. T., Niyigena, S., Msambichaka, V., Nonga, H. E., Antoine-Moussiaux, N. & Fasina, F. O. (2021). Antimicrobial use, residues, resistance and governance in the food and agriculture sectors, tanzania. *Antibiotics*. 10(4):378-389.
- 33.Nataf, Y., Yaron, S., Stahl, F., Lamed, R., Bayer, E. A., Scheper, T. H., Sonenshein, A. L., Shoham, Y., Zuroff, T. R., Xiques, S. B., Curtis, W. R., Gu, W., Fore, R. L., Leschine, S. B., Curtis, W. R., Zhang, X. Z., Zhang, Z., Zhu, Z., Sathitsuksanoh, N.&Tardif, C. (2013). The Rnf Complex of. Applied and Environmental Microbiology. 8(1).
- 34. Nikaido, H. (2009). Multidrug resistance in bacteria. *Annual Review of Biochemistry*. 78 (11):189-197.
- 35.Oliveira, D., Borges, A. & Simões, M. (2018). *Staphylococcus aureus* toxins and their molecular activity in infectious diseases. *Toxins*. *10* (6):234-242.
- 36.Idbeis, H. I..& Khudor, M. H (2019). Detection Of Intracellular Adhesion Gene (Icaa and Icad) and Biofilm Formation *Staphylococcus aureus* Isolates From Mastitis Milk of Sheep and Goat. *Basrah Journal of Veterinary Research*. 18 (2): 306-327.
- 37.Patel, R. (2000). Enterococcal-type glycopeptide resistance genes in non-enterococcal organisms. In FEMS Microbiology Letters. 185(1): 254-263.
- 38. Patel, R., Piper, K., Cockerill, F. R., Steckelberg, J. M. & Yousten, A. A. (2000).

- The biopesticide Paenibacillus popilliae has a vancomycin resistance gene cluster homologous to the enterococcal VanA vancomycin resistance gene cluster. *Antimicrobial Agents and Chemotherapy.* 44 (3):199-211.
- 39.Pérez, V. K. C., Costa, G. M. da, Guimarães, A. S., Heinemann, M. B., Lage, A. P. & Dorneles, E. M. S. (2020). Relationship between virulence factors and antimicrobial resistance in *Staphylococcus aureus* from bovine mastitis. *journal of Global Antimicrobial Resistance* .4(22):315-327.
- 40.Radostits, O. M. & Done, S. H. (2010). Veterinary Medicine A Textbook of the Diseases of Cattle, Horses, Sheep, Pigs and Goats, 10th edition. *The Canadian Veterinary Journal*. *51*(5):224-233.
- 41.Rasigade, J. P. & Vandenesch, F. (2014). *Staphylococcus aureus*: A pathogen with still unresolved issues. Infection, *Genetics and Evolution*, 21:161-172.
- 42. Rubinstein, E. & Keynan, Y. (2014). Vancomycin revisited 60 years later. *Frontiers in Public Health*. 2(13):87-95.
- 43. Ruffing, U., Alabi, A., Kazimoto, T., Vubil, D. C., Akulenko, R., Abdulla, S., Alonso, P., Bischoff, M., Germann, A., Grobusch, M. P., Helms, V., Hoffmann, J., W. V., Kremsner, Kern, Р. Mandomando, I., Mellmann, A., Peters, G., Schaumburg, F., Schubert, S.& Herrmann, M. Community-associated (2017).Staphylococcus aureus from Sub-Saharan Africa and Germany: A cross-sectional

- geographic correlation study. *Scientific Reports*. 7(1): 286-293.
- 44.Smith, T. H., Fox, L. K. & Middleton, J. R. (1998). Outbreak of mastitis caused by one strain of *Staphylococcus aureus* in a closed dairy herd. *Journal of the American Veterinary Medical Association*. 212(4); 133-142.
- 45. Anad, I., A. Abbas, B. & Khudaier, B.Y. (2014). Isolation of *Staphylococcus aureus* from buffalo milk in basrah governorate and detection of their antibiotic susceptibility. *Basrah Journal of Veterinary Research*. *13*(1): 235-245.
- 46.Tacconelli, E., Carrara, E., Savoldi, A., Harbarth, S., Mendelson, M., Monnet, D. L., Pulcini, C., Kahlmeter, G., Kluytmans, J., Carmeli, Y., Ouellette, M., Outterson, K., Patel, J., Cavaleri, M., Cox, E. M., Houchens, C. R., Grayson, M. L., Hansen, P., Singh, N. & Zorzet, A. (2018). Discovery, research, and development of

- new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis. *The Lancet Infectious Diseases. 18*(3): 312-318.
- 47. Taylor, A. & Unakal, C. G. (2020). *Staphylococcus aureus* StatPearls NCBI Bookshelf. *StatPerals*. 4:65-72.
- 48. World Health Organization. (2013). WHO News Release. *Saudi Medical Journal*, *34*(11): 113-121.
- 49.Yip, C. H., Mahalingam, S., Wan, K. L. & Nathan, S. (2021). Prodigiosin inhibits bacterial growth and virulence factors as a potential physiological response to interspecies competition. *PLoS ONE*. *16*(6): 76-83.
- 50.Zrnčić, S. (2020). European union's action plan on antimicrobial resistance and implications for trading partners with example of national action plan for Croatia. *Asian Fisheries Science*. *33*(1): 321-331.

# مشكلة مقاومة المكورات العنقودية للميثيسيلين (MRSA) والفانكومايسين (VRSA) في الانسان وحلولها.

مروان میثم محمد  $^{1}$ , محمد حسن خضر  $^{2}$ , هناء خلیل ابر اهیم  $^{2}$ .

1-قسم تقنية المختبرات الطبية/كلية البصرة الجامعة للعلوم والتكنولوجيا/البصرة/العراق

2-فرع الاحياء المجهرية/كلية الطب البيطري/جامعة البصرة/العراق

### الخلاصة

المكورات العنقودية الذهبية هي بكتيريا انتهازية تصيب البشر والثديبات والتي أصبحت أكثر أهمية من الناحية السريرية والبيطرية بسبب تطورها السريع لمقاومة المضادات الحيوية. بعض أصناف المكورات العنقودية الذهبية هذه مقاومة للميثيسيلين (MRSA)، وهو أمر شائع في منظمات الرعاية الصحية، والبيئات المجتمعية، ومزارع الماشية في جميع أنحاء العالم. وتعتبر MRSA ذات قدره على العيش في أنواع حيوانية أخرى، مما قد يؤدي إلى ظهور وانتشار مقاومة العوامل المضادة للميكروبات في مختلف الأنواع الحيوانية. الفانكومايسين هو نوع من المضادات الحيوية يُصنف من الحلول الأخيرة لمكافحة هذه البكتريا، يستخدم لمعالجة الالتهابات الشديدة التي تحرضها البكتيريا إليجابية الصابغة. يعتبر استخدام الفانكومايسين من بين الخيارات الأساسية للعلاج الأولي لعدوى (MRSA) خلال السنوات القليلة الماضية، كان هناك ظهور لسلالات المكورات العنقودية الذهبية التي تظهر مقاومة قوية للفانكومايسين. تعمل مجموعة الجينات (vanA) التي تم الحصول عليها من المكورات المعوية المقاومة الفانكومايسين، على تسهيل نقل مقاومة الفانكومايسين في الميوانات المكورات العنقودية الذهبية المكورات العنقودية المقاومة الميثيسيلين والفانكومايسين من الحيوانات إلى البشر والعكس، تهدف هذه المراجعة إلى تسليط الضوء على مشكلة المكورات العنقودية الذهبية المقاومة الميثيسيلين. (MRSA) للمكورات العنقودية الذهبية المقاومة الميثيسيلين.

الكلمات المفتاحية: المكورات العنقوديه الذهبيه، مقاومة البنسلين، مقاومة الفانكومايسين ، الانسان ، الحيوان.