

Molecular Identification of *Geobacillus* WCH 70 Isolate according to Nitrate reductase gene sequence

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Abstract

This study aimed to provide the molecular identity of *Geobacillus* WCH70 according to Nitrate reductase gene. DNA was extracted the purity was 1.8 ng\ μ l, then Nitrate reductase (NR) gene was amplified by a specific primer pairs, the results showed that the size of NR gene was 1626 bp after use a 1% gel electrophoresis. Then Nitrate reductase gene was sequenced according to the chain termination method. The sequence results were showed the nucleotide similarity 100% percentage of identical nitrate reductase *Geobacillus* WCH70, on the other hand the sequence similarity was ranged between 77% and 88% with sequence of other microorganisms.

Key Words: Nitrate reductases NR, *Geobacillus* WCH 70

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التحديد الجزيئي للعزلة المحلية *Geobacillus* WCH 70 بالاعتماد على التابع الجيني لجين

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الخلاصة

هدفت هذه الدراسة الى التشخيص الوراثي للعزلة المحلية *Geobacillus* WCH70 بالاعتماد على وجود جين النايتريت ريدكتيز. استخلص الدنا بنقاوة 1.8 نانوغرام/ مايكروليتر، ثم تمت مضاعفة الجين المسؤول عن انتاج النايتريت ريدكتيز باستخدام زوج من البوادى المتخصصة، لوحظ من النتائج ان حجم جين النايتريت ريدكتيز كان 1626 bp بعد الترحيل الكهربائي على هلام الاكاروز تركيز 1% بعدها تم تحديد التابع الجيني لجين النايتريت ريدكتيز باستخدام طريقة ايقاف السلسلة اظهرت نتائج تحديد التشابه للقواعد النروجينية للنموذج قيد الدراسة متطابق مع التابع لجين النايتريت ريدكتيز للعزلة WCH70 بنسبة، 100% من ناحية اخرى كانت نسبة التشابه للنموذج قيد الدراسة مع الانواع البكتيرية الاخرى من حيث التشابه بالمحتوى الوراثي تتراوح بين 77-88%.

الكلمات المفتاحية: النايتريت ريدكتيز NR جيوباسلس WCH70.

Introduction

Nitrogen is a basic element for life because it is a component of the two preeminent biological macromolecules: proteins and nucleic acids. Nitrate reduction plays a key role in the nitrogen cycle and has important agricultural, environmental, and public health implications (1). As assimilatory nitrate reduction, performed by bacteria, fungi, algae, and higher plants (2, 3, 4). Some organisms have more than one type of nitrate reductase for example *E.coli* consists of three types of NRs. The availability of more than one type of NRs indicates the preference for anaerobic respiration. Thermophilic bacteria are described as aerobes although the low availability of oxygen in high temperature environments (5, 6). Nitrate reductase primary reaction is catalyzing nitrate to nitrite. Based on the electron donors the nitrate reductases are classed into three groups (7). Thermophilic organisms have a potential to produce thermostable enzymes with higher stability and longer life (8, 9). Using thermostable bacteria in industrial processes reduces the probability of contamination, increase diffusion rate and reduces

the cost of external cooling (10). Nitrite reductase is the key enzyme in the dissimilatory nitrification process. The reduction of nitrite to NO can be catalyzed by the products of two different nitrite reductase genes: one product contains copper (the *nirK* product), and the other contains cytochrome cd1 (the *nirS* product). The two genes seem to occur mutually exclusively in a given strain, but both types have been found in different strains of the same species (11, 12). Although structurally different, both enzyme types are functionally and physiologically equivalent. *nirS* is more widely distributed; *nirK* is found in only 30% of the denitrifiers studied so far. However, *nirK* is found in a wider range of physiological groups (13,14). Eukaryotic and prokaryotic assimilatory nitrate reductase share no sequence similarity and have little in common beyond their physiological function (15, 16). The aim of this study, we report on the application of new primer pairs for nitrite reductase genes to determine the NR gene of denitrifying *Geobacillus* WCH70 by amplifying and sequences NR fragments successfully, then detect the similarity and identity of query with other different populations of denitrifying bacteria.

Material and Methods

- **Bacterial Strain:** *Geobacillus* WCH 70 Isolate was provide by the Biotechnology department\ Collage of Science\ Baghdad University.
- **DNA Extraction:** *Geobacillus* WCH 70 chromosomal DNA was extracted and purified using a spin column kit (Clontech, Mountain View, CA, USA)(17).
- **Primer's:** Two PCR primers were designed and used to amplify Nitrate reductases of *Geobacillus* WCH 70 supplied by Integrated DNA Technology(IDT\ Canada). To obtained DNA fragment was 1626bp.

FW: 5'-ATGGCATACGAAAAAGTATGG-3'

RW: 5'-TTATGCTACCGTGCTGAC-3'

- **Amplify Nitrate reductases gene:** Amplification was preformed using automated thermal cycler (Amplifon II; Thermolyne' Dubuque' IA), using 0.5 µl pfu DNA polymerase (Stratagene), 1 µL 20 µM primer pairs of forward/reversward primers were added into PCR mixture, 1.5 µL of *Geobacillus* WCH70 chromosomal DNA was used as a template, 5 µL of 10 mM deoxynucleotide triphosphates (dNTPs) Mix, 5 µL of 10X PCR reaction buffer, and 36 µL of sterile distilled water was added to complete the final volume to 50 µL. PCR cycle was programmed as follows: One cycle of 95 °C for 3 min; 35 cycles of 95 °C for 30 sec, 52 °C for 30 sec, 72 °C for 3 min; and final extension at 72 °C for 5 min. The PCR products were detected by electrophoresis on a 0.8% agarose electrophoresis containing ethidium bromid run at a constant voltage (100 V) for 30 min, and visualized by using a Gel documentation systems (USA). Then the PCR product was purified by StrataPrep® PCR purification kit (Stratagene, Dedar Creek, TX, USA). (17,18).
- **Sequencing of Nitrate reductases gene:** Nitrate reductases gene was amplified from *Geobacillus* WCH70 and detected on agarose gel, then the complete sequenced was performed in the center of applied genomics institute\ Canada. (19) Sequence similarity of *Geobacillus* WCH70 to other Nitrate reductases from other organisms was detected using bioinformatic database in the National center for bioformation (NCBI\BLAST) on website WWW.NCBI.net. Where the accession number in NCBI CP001638.1

Results and Discussion

- **Amplified of Nitrate reductases gene:** Genomic DNA was extracted from G. WCH70 production by using DNeasy® tissue Kit (Qiagen, Valencia,CA, USA). Specific primers were used for amplification. Results indicated in Fig. (1) showed the amplified nitrate reduction gene fragment with a molecular size of about 1626

bp. while in the other studies the nitrate reduction from other bacteria such as *Thiosphaera pantotropha* consisted of cluster genes *napEDABC* encoding the periplasmic nitrate reductase, all five open reading frames have a codon usage and GC bias at the third position similar to that found in *P. denitrificans* genes (9, 20, 21), analysis of the 500 bp 3' to *napC* did not identify any possible open reading frame in this region, thus appears likely that the periplasmic nitrate reductase operon of *T. pantotropha* comprises the five genes *napEDABC*. (22,23)

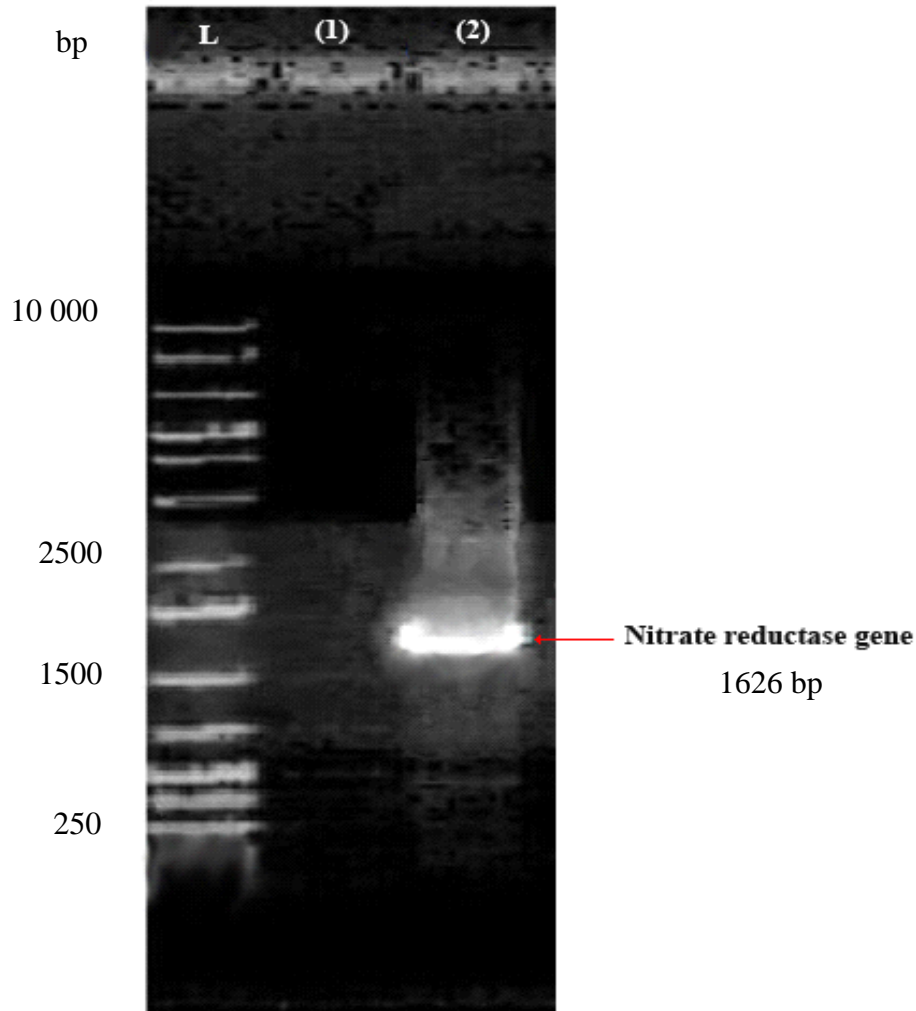


Fig. (1) *Geobacillus* WCH70 Nitrate reductases gene. Electrophoretically analyzed on agarose gel (0.8%) for 1 hour

Lane (1): Landmarker ladder (10000bp).

Lane (2): Nitrate reductases gene fragment.

This fragment was eluted, and sequencing.

- **Sequencing of nitrate reduction gene:** To determine *Geobacillus* sp. WCH70 Nitrate reductase gene sequence identity to sequences of other microorganism nitrate reductase, nitrate reductase gene was firstly sequenced and analysed to determine ORF. Results indicated in Fig. (2) showed the complete *G. WCH 70* nitrate reductase gene of 1626 bp in length, rich in GC 44.7% codes for 541 amino acids, was detected in the center of applied genomics institute/ Canada. Sequencing was achieved according to chain termination method(22), query results of nitrate reductase gene was 100% identical to the nucleotide sequence and deduced amino acids of *Geobacillus* sp. WCH70 nitrate reductase gene covering 100% DNA region of gene as mentioned in Fig. (3). From the results it was found that the degree of similarity between the nucleotide sequence of nitrate reductase gene (query) to the *Geobacillus* WCH70 was 100% identical.

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1 atggcatacgaaaaagtatgggcagataatccgaaattaaacaaaatggaattaaanaag 60
M A Y E K V W A D N P K L N K M E L K K
61 cttaaaaaagatggactaaaaatttttgaagatattccttattatgcaaagcatggattc 120
L K K D G L K I F E D I P Y Y A K H G F
121 tcttcgattccgcaagaagaatgggattatttcaaattgggcagggctttacctgcaacgc 180
S S I P Q E E W D Y F K W A G L Y L Q R
181 ccaaaagaagatggctattttatgatgcgctgtttgtgttccttccggaattttatctaac 240
P K E D G Y F M M R V C V P S G I L S N
241 gagcagctcgaacgtagcagggcagcctgattacggacgcggtttgtttgacatt 300
E Q L E T L A G I A R D Y G R G L F D I
301 acaaccctgcaatctgttcaattccattggctgcaaatcgaacaaattcctgatattttt 360
T T R Q S V Q F H W L R I E Q I P D I F
361 gatcgtttaaaacgggttgattatctaccgctggagcttgcggcgatatcaccggcaac 420
D R L K R V G L S T A G A C G D I T R N
421 atcgtcggcaaccgcttgctggtatcgatccggacgaactggttgacacgagagaaatc 480
I V G N P L A G I D P D E L F D T R E I
481 Gttcgtgaagtatatgagttttccaacataacgaagacttctcgaactgcccgcgcaaa 540
V R E V Y E F F Q H N E D F S N L P R K
541 tataaaatatcgatcagtgccaacgtctacaacacgggtaacgcagaaatccattgcttg 600
Y K I S I S A N V Y N T V N A E I H C L
601 tcgtttacaccggcgaaaaaagtgatcgacggcaaaagaagtgctcggattccatgtgaaa 660
S F T P A K K V I D G K E V L G F H V K
661 Gtcggcgcggtttatcgtctgctccatatcttgcccaaacactggatgtgttcgtgacg 720
V G GG L S S A P Y L A Q T L D V F V T
721 ccagatcgcgtgaaagacgctcgtggtgctgcacaaacgattttccgcgattacggctat 780
P D R V K D V A V A V T T I F R D Y G Y
781 Cgcgaaaaacgccatcgcgcccgtctcaaattccttgcgctgactggggcggttgataaa 840
R E K R H R A R L K F L V A D W G V D K
841 tttaaagaaaaattaattgagctcacggggcctctcccatccaaaggggaagacgtacaa 900
F K E K L I E L T G P L P S K G E D V Q
901 aagggtggaaccctggttacttctacggcgtccatcgccaaaaacaagaaggattaaat 960
K G W N P G Y F Y G V H R Q K Q E G L N
961 ttcacggttttaattgtgccagttggccgcttgagcgcggatgaagtattcgaatcgcg 1020
F I G F N V P V G R L S A D E V F E I A
1021 cgcatagcccgaataacggcaacggcgaagtccgcacatgcaattcacaaacctaatt 1080
R I A R Q Y G N G E V R T C N S Q N L I
1081 attccaaatattccagataaaatattgttgacgctgtattacaggaaaaagtgtttgaacgg 1140
I P N I P D K Y V D A V L Q E K V F E R
1141 atttctatccgtccgaacacattcaccggctattctgtttctgtacaggcaccgagttt 1200
I S I R P N T F T G Y S V S C T G T E F
1201 tgcaatttggcgctcgtcgagacgaaagaacggatgcgggcgctcgtgaatatttagat 1260
C N L A L V E T K E R M R R V A E Y L D
1261 cagcatttagaactggacggtccggtagcattcatatgggttggtgtccaaactcatgc 1320
Q H L E L D V P V R I H M V G C P N S C
1321 ggccaacggcaaatggagatattcggcctgcaaggaatcaaaacaaaaacaaaagacaga 1380
G Q R Q I G D I G L Q G I K T K T K D R
1381 ggcattgattgaagcgtttgaaatttacgtcggcggcagctctacaacggcggcaaac 1440
G M I E A F E I Y V G G T L Y N G G K Y
1441 aatcaaaaattaaaaggaaaagttagatgcggaacgaatttccgaagtgttttgacgcta 1500
N Q K L K G K V D A E R I S E V L L Q L
1501 ttaacctatttcaaagaacaaaattgcccggcgaacattccttagcttacttagatcgc 1560
L T Y F K E T K L P G E T F L A Y L D R
1561 gttggaatagaagcgttgcaaacagaactagacagcatttttagcgaagtcagcagcgta 1620
V G I E A L Q T E L D S I L A Q V S T V
1621 gcataa 1626
A -

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Fig. (2) Nucleotide sequence of open reading frame (ORF) Nitrate reductases gene from *Geobacillus* WCH70.

On the other hand results mentioned in Table (3) showed the G. WCH70 nitrate reductase gene sequence was less identical (77-88%) to other *Geobacillus* sp. (*Geobacillus thermoglucosidasius* strain DSM 2542, complete genome, *Geobacillus thermoglucosidasius* C56-YS93, complete genome, *Geobacillus* sp. Y4.1MC1, complete genome, *Anoxybacillus* sp. B7M1, complete genome).

Table (3) Nucleotide sequence alignment of G. WCH70 nitrate gene with related sequences of different microorganisms. NCBI / Blast (WWW.NCBI.net).

Description	Max identity	Query coverage
<i>Geobacillus</i> sp. WCH70, complete genome	100%	100%
<i>Geobacillus thermoglucosidasius</i> strain DSM 2542, complete genome	88%	100%
<i>Geobacillus thermoglucosidasius</i> C56-YS93, complete genome	88%	100%
<i>Geobacillus</i> sp. Y4.1MC1, complete genome	88%	100%
<i>Anoxybacillus</i> sp. B7M1, complete genome	77%	97%

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