

Effect of lipopolysaccharide on the liver function test**G. S. Abd and B. Hg. Abdul-Hameed****State company for veterinary services\Ministry of Agriculture****Abstract**

Lipopolysaccharide (LPS; endotoxin), a major component of the outer membrane of the Gram-negative bacterial cell wall. Among the changes observed upon exposure to LPS are fever, circulatory shock, disseminated intravascular coagulation, and damage to numerous organs including the liver, acute lung injury and acute renal failure. 40 male Wister albino rats were divided in to 3 groups. Control group was given tap water during the experiment period. At the last day of the experiment, animal were injected intraperitoneally with distilled water. The second group was given ethyl alcohol with drinking water, at the end of the experiment 10 rats were injected intraperitoneally with distilled water and the other 10 rats were injected intraperitoneally with LPS (1µg/gm). Endotoxin group was given tap water during the experiment period, at the last day of the experiment period they were injected intraperitoneally with LPS (1µg/gm). for determining the biochemical tests the blood were collected in test tubes for obtaining the serum samples. Injection of LPS significantly increased the activity of alkaline phosphatase enzyme (SALP) and bilirubin level in serum as well as serum glutamic oxaloacetic transaminase (SAST) and glutamic pyruvic transaminase (SALT) were also increased compared with control rats. LPS significantly increased (SAST), (SALT), the activity of alkaline phosphatase enzyme (SALP) and bilirubin level in serum and it's regarded as a natural results of organs damage and dysfunction which caused by LPS insult.

تأثير متعدد السكريات الدهني على اختبار وظائف الكبد**غسان سامي عبد وبان غسان عبد الحميد****الشركة العامة للسيطرة/ وزارة الزراعة****الخلاصة**

متعدد السكريات الدهني (الذي يمتل الجزء الرئيسي في الغلاف الخارجي لبكتريا سالبة صبغة كرام. معظم التغيرات التي لوحظت هي ارتفاع درجة الحرارة، الصدمة الدموية. تجلط داخل الأوعية الدموية، أذى الأعضاء الداخلية والتي تشمل الكبد والأذى الحاد في الرئة والفشل الكلوي الحاد. 40 ذكر من جردان نوع وستير البيضاء تم تقسيمها بصورة عشوائية إلى ثلاث مجاميع، تم إعطاء مجموعة السيطرة ماء مقطر خلال فترة التجربة. عند اليوم الأخير من التجربة تم حقن الحيوانات بالماء المقطر بالبريتون. المجموعة الثانية تم إعطائها الكحول الاثيلي مع ماء الشرب عند نهاية التجربة عشرة من الجردان تم حقنها بالبريتون بالماء المقطر وعشرة أخرى حقنت بالذيان الداخلي بجرعة مقدارها (1 ملغم/غم) بالبريتون. مجموعة الذيان الداخلي أعطيت ماء مقطر خلال التجربة في اليوم الأخير من التجربة حقنت بالذيان الداخلي بجرعة (1 ملغم/غم) بالبريتون لغرض تحديد التغيرات الكيموحيوية تم جمع الدم بواسطة أنابيب اختبار لتحليل عينات المصل. أظهرت النتائج ان حقن الذيان الداخلي أدى إلى حدوث زيادة معنوية في مستوى الإنزيم القاعدي الفسنتيز (SALP) وزيادة في البليرولبين في مصل الدم مع

زيادة نسبة (SAST) و (SALT) مقارنة مع مجموعة السيطرة. من ذلك نستنتج بوجود زيادة معنوية في كل من إنزيمي (SAST) و (SALT) وزيادة فعالية إنزيم (SALP) والبيروليبين في مصل الدم ويرجع سبب ذلك كنتيجة طبيعية للأذى الحاصل وعدم اختلال وظيفة الكبد بسبب الذيفان الداخلي.

Introduction

Endotoxins, also called lipopolysaccharides (LPS), are a major component of the outer membrane of Gram-negative bacteria. They are composed of a hydrophilic polysaccharide moiety, which is covalently linked to a hydrophobic lipid moiety (Lipid A) (1, 2, 3). LPS from most species is composed of three distinct regions: the O-antigen region, a core oligosaccharide and Lipid A (Lip A). The lipid A is the most conserved part of endotoxin and is responsible for most of the biological activities of endotoxin, i.e. its toxicity (4). The O-antigen is generally composed of a sequence of identical oligosaccharides (with three to eight monosaccharides each), which are strain specific and determinative for the serological identity of the respective bacterium (5). Lipopolysaccharide (LPS; endotoxin) is among the most potent modulators of the innate immune system. It is responsible for a host of toxic effects that occur in patients infected with Gram-negative bacteria, including fever, disseminated intravascular coagulation, and hemodynamic changes, which may lead to multiple organ failure characteristics of the septic shock syndrome (6). Lipopolysaccharide triggers increased production of a variety of proinflammatory mediators, such as tumour necrosis factor- α (TNF- α), interleukin 1 β (IL-1 β), IL-6, and nitric oxide (NO) (7), as well as increased production of chemokines and cell adhesion molecules, leading to the recruitment of several lineages of white blood cells, including neutrophils, to the liver (8). If uncontrolled, these proinflammatory responses contribute to liver injury and acute hepatic failure (9, 10, 11). Disseminated intravascular coagulation (DIC) is characterized by a systemic activation of the blood coagulation system, which results in the generation and deposition of fibrin, leading to microvascular thrombi in various organs and contributing to the development of multiorgan failure (12). Consumption and subsequent exhaustion of coagulation proteins and platelets, due to the ongoing activation of the coagulation system, may induce severe bleeding complications, although microclot formation may occur in the absence of severe clotting factor depletion and bleeding (13). Endotoxin causes multiple organ dysfunctions, including acute lung injury. Acute lung injury is characterized by acute lung inflammation involving the local recruitment and activation of polymorphonuclear neutrophils and the release of proinflammatory mediators, (14, 15) proteases, and reactive oxygen and nitrogen species. Eventually, these processes can cause alveolar-capillary damage with high permeability pulmonary edema and alteration of lung mechanics, resulting in severe gas exchange abnormalities (16). Sepsis is a common cause of acute renal failure (ARF) (17, 18), an entity in which the abrupt decline in glomerular filtration rate is associated with the pathological lesion of acute tubular necrosis (18).

Materials and Methods

- Treatment of animals:

40 male Wister albino rats were housed in cages (5 rats per cage) and randomly divided into 3 groups.

1. Control group: 10 rats were given tap water during the experiment period (90 days). At the end of 90 days the animal were injected intraperitoneally with distilled water.
2. The second group: (20 rats) were given ethyl alcohol with drinking water and the dose of ethanol was increased progressively till its concentration reached 20% at the 5th week,

at the end of the experiment 10 rats were injected intraperitoneally with distilled water and the other 10 rats were injected intraperitoneally with LPS (1µg/gm) (LPS from segma com).

3. Endotoxin group: 10 rats were given tap water during the experiment period, at the last day of the experiment period they were injected intraperitoneally with LPS (1µg/gm) (LPS from segma com).

Twenty– four hours after experimental period the animals were killed and the blood were collected in test tubes for obtaining the serum samples to determine the biochemical tests.

Results

Injection of LPS significantly increased the activity of alkaline phosphatase enzyme (SALP) (Table 1) compared with control rats. (Table 2) shows that there is significant increase in bilirubin level in serum compared with control rats. Furthermore plasma biochemistry glutamic pyruvic transaminase (SALT) and glutamic oxaloacetic transaminase (SAST) were also increased compared with control rats (Table 3,4).

Table (1) Effect of endotoxin to the activity of alkaline phosphatase enzyme (SALP) (IU/L)

	Control	Endotoxin
1	89	138
2	110	121
3	113	149
4	107	82
5	121	131
6	75	89
7	50	92
8	71	82
9	71	92
10	60	107

(P< 0.1)

Table (2) Effect of endotoxin on the level of serum total bilirubin (mg/100 ml)

	Control	Endotoxin
1	0	0.3
2	0	0.2
3	0	0.5
4	0	0.2
5	0	0.5
6	0	0.1
7	0	0.2
8	0	0.3
9	0	0.2
10	0	0.1

(P< 0.1)

Table (3) Effect of endotoxin on serum (SALT) level (IU/L)

	Control	Endotoxin
1	12	18
2	18	18
3	18	29
4	20	22
5	24	18
6	9	24
7	16	22
8	16	22
9	16	36
10	12	24

(P< 0.1)

Table (4) Effect of endotoxin on serum (SAST) level (IU/L)

	Control	Endotoxin
1	64	96
2	60	80
3	60	96
4	47	72
5	80	88
6	31	37
7	31	47
8	39	60
9	35	41
10	17	51

(P< 0.1)

Discussion

Our results show that there is significant increase in alkaline phosphatase enzyme (SALP) activity. Previous studies show that increased alkaline phosphatase level are associated with hepatic damage and it has been demonstrated that alkaline phosphatase attenuates the lipopolysaccharide mediated inflammatory response because alkaline phosphatase plays essential roles in detoxification of pathogenic bacterial (LPS) endotoxin by phosphorylation it's lipid A moiety (19, 20, 21). We show that there is significant increase in bilirubin level in serum compared with control rats. This mean that the liver is incapable of removing bilirubin from the blood (22). Plasma bilirubin increased due to hepatocyte dysfunction as a result of hepatocellular disease (eg. Infective, auto immune, toxic, septic, hypoxia) (23, 24) demonstrated that hyper bilirubinemia can occur after (G+ve) or (G -ve) bacteremia and endotoxin- mediated cytokine release is the likely cause of septic jaundice .a variety of bacterial infection affect the liver either as a result of direct hepatocellular or biliary invasion or through the production of toxins (LPS) or endotoxin is a potent inducer of cytokines. endotoxin- mediated cytokine release may be the bases for jaundice of many disorders, including sepsis. Early studies showed areduction in bile flow and biliary excretion after administration of endotoxin to isolate perfused rat liver. Studies have shown that canalicular bile acid and organic anion transport are markedly impaired in endotoxemia therefore, endotoxemia severely impaires the transport of organic anions at both the sinusoidal and canalicular membrane. Because of impaired hepatic organic anion transport both bile acid-dependent and bile acid-independent component of bile flow decreased with administration of endotoxin. LPS significantly increased blood (SALT) and (SAST) (markers of organs injury) (25). (SALT) and (SAST) is acytoplasmic

hepatocellular enzyme whose increase in blood is highly indicative for hepatocellular damage or injury from different types of disease (26). Endotoxin shock can induce the production of nitric oxide (27) and several inflammatory mediators such as TNF- α , IL-6 and IL-1 β leading to multiple organ dysfunction and death (27, 28).

References

1. Ogikubo, Y.; Norimatsu, M.; Noda, K.; Takahashi, J.; Inotsume, M.; Tsuchiya, M. & Tamura, Y. (2004). Evaluation of the bacterial endotoxin test for quantification of endotoxin contamination of porcine vaccines. *Biologicals*, 32:88-93.
2. Hirayama, C. & Sakata, M. (2002). Chromatographic removal of endotoxin from protein solutions by polymer particles. *J. Chrom. B.*, 781:419-432.
3. Raetz, C. R.; Ulevitch, R. J.; Wright, S. D.; Sibley, C. H.; Ding, A. & Nathan, C. F. (1991). Gram-negative endotoxin: an extraordinary lipid with profound effects on eukaryotic signal transduction. *The FASEB J.*, 5(12):2652-2660.
4. Magalhaes, P.; Lopes, A.; Mazzola, P.; Rangel-Yagui, C.; Penna, T. & Pessoa, A. (2007). Methods of Endotoxin Removal from Biological Preparations: a Review. *J. Pharm. Pharmaceut. Sci.*, 10(3):388-404.
5. Petsch, D. & Anspach, F. B. (2000). Endotoxin removal from protein solutions. *J. of Biotechnol.*, 76:97-119.
6. Gao, B.; Wang, Y. & Tsan, M. (2006). The heat sensitivity of cytokine-inducing effect of lipopolysaccharide. *J. of Leukocyte Biol.*, 80:359-366.
7. Liu, D.; Zeng, B. X.; Zhang, S. H.; Wang, Y. L.; Zeng, L. & Geng, Z. L. (2005). Rosiglitazone, a peroxisome proliferator-activatedreceptor-gamma agonist, reduces acute lung injury in endotoxemic rats. *Crit. Care Med.*, 33: 2309-2316.
8. Han, D. W. (2002). Intestinal endotoxemia as a pathogenetic mechanism in liver failure. *World J. Gastroenterol.*, 8: 961-965.[Web of Science][Medline].
9. Bilzer, M.; Roggel, F. & Gerbes, A. L. (2006). Role of Kupffer cells in host defense and liver disease. *Liver Int.*, 26: 1175-1186.[Cross Ref][Web of Science][Medline].
10. Qin, X. & Gao, B. (2006). The complement system in liver diseases. *Cell Mol. Immunol.*, 3: 333-340.[Medline].
11. Jirillo, E.; Caccavo, D.; Magrone, T.; Piccigallo, E.; Amati, L.; Lembo, A.; Kalis, C. & Gumenscheimer, M. (2002). The role of the liver in the response to LPS: experimental and clinical findings. *J. Endotoxin Res.*, 8: 319-327. [Cross Ref] [Medline].
12. Vincent, J. L. & De Backer, D. (2005). Does disseminated intravascular coagulation lead to multiple organ failure. *Crit. Care Clin.*, 21(3):469-77. [Medline].
13. Levi, M. & Ten Cate, H. (1999). Disseminated intravascular coagulation. *N. Engl. J. Med.*, 341(8):586-92. [Medline].
14. Wright, R. M.; Ginger, L. A.; Kosila, N.; Elkins, N. D.; Essary, B. & McManaman, J. L. (2004). Mononuclear phagocyte xanthine oxidoreductase contributes to cytokine-induced acute lung injury. *Am. J. Respir. Cell Mol. Biol.*, 30:479-490.
15. Shinbori, T.; Walczak, H. & Krammer, P. H. (2004). Activated T killer cells induce apoptosis in lung epithelial cells and the release of pro-inflammatory cytokine TNF-alpha. *Eur. J. Immunol.*, 34: 1762-1770.
16. You, S.; San-peng, X.; Yan, W.; Yuan-xu, J.; Zhou-yang, W.; Shi-ying, Y. & Shang-long, Y. (2009). Melatonin reduces acute lung injury in endotoxemic rats. *Chin Med. J.*, 122(12):1388-1393.

17. Wu, X.; Guo, R.; Wang, Y. & Cunningham, P. (2007). The role of ICAM-1 in endotoxin-induced acute renal failure. *AJP-Renal Physiol.*, 293(4):F1262-F1271.
18. Cunningham, P.; Holers, V.; Alexander, J.; Guthridge, J.; Carroll, M. & Quigg, R. (2000). Complement is activated in kidney by endotoxin but does not cause the ensuing acute renal failure. *Kidney International*, 58: 1580-1587.
19. Beumer, C.; Wulferink, M.; Raaben, W.; Fiechter, D.; Ruud Brands, R. & Seinen, W. (2003). Calf Intestinal Alkaline Phosphatase, a Novel Therapeutic Drug for Lipopolysaccharide (LPS)-Mediated Diseases, Attenuates LPS Toxicity in Mice and Piglets. *JPET*. 307(2): 737-744.
20. Bates, J.; Akerlund, J.; Mittge, E. & Guillemin, K. (2007). Intestinal alkaline phosphatase detoxifies lipopolysaccharide and prevents inflammation in zebrafish in response to the gut microbiota. *Cell Host and Microbe*.2(6):371-382.
21. Lackeyram, D.; Yang, C.; Archbold, T.; Swanson, K. & Fan, M. (2010). Early Weaning Reduces Small Intestinal Alkaline Phosphatase Expression in Pigs. *J. of Nut.*, 140(3): 461-468.
22. Lab Test Online. (2001- 2011) by American Association for Clinical Chemistry.<http://labtestonline.org.au/>.
23. Jones, G. (2010). Bilirubin. SydPath The Pathology Service of St Vincent's Hospital Sydney, Australia (gjones@stvincents.com.au).
24. Schiff, E. R.; Sorrell, M. F. & Maddrey, W. C. (2007). Schiff's diseases of the liver. Lippincott Williams & Wilkins. Wolters Kluwer business. PP.1379-1380.
25. Wu, W.; Hu, T.; Lin, N.; Subeq, Y.; Lee, R. & Hsu, B. (2010). Low-dose erythropoietin aggravates endotoxin-induced organ damage in conscious rats. *Cytokine*. 49(2):155-162.
26. Nellithady, G. S.; Anila, K.; Kumar, K. K. & Kaveri, H. (2010). Lack of Association of Chronic Liver Disease in Patients with Oral Lichen Lanus. *J. Carcinogene Mutagene*.1(3): 2157-2518.
27. Wen-Jinn, L.; Chin, T.; Jeng-Yuan, W.; Shiu-Jen, C.; Jih-Hsin, W. & Chin-Chen, W. (2003). Inhibition by Terbutaline of Nitric Oxide and Superoxide Anion Levels of Endotoxin-Induced Organs Injury in the Anesthetized Rat. *Shock*. 19(3): 281-288.
28. Guerra-Ruiz, A.; Casafont, F.; Cobo, M.; Terán, A.; de-la-Peña, J.; Estebanez, A. & Pons-Romero, F. (2010). Increased Bactericidal/Permeability Increasing Protein in Patients with Cirrhosis. *Liver International*, 30(1):94-101.