

Iraqi Journal of Veterinary Sciences



www.vetmedmosul.com

Molecular effects of nimesulide and aspirin on caspase-3, PPAR- α , and COX-2 gene expression in mice

T.A. Yahya[®] and Y.J. Mousa[®]

Department of Physiology, Biochemistry, and Pharmacology, College of Veterinary Medicine, University of Mosul, Mosul, Iraq

Article information Abstract Article history: The characterization of NSAIDs in reducing pain, inflammation, and fever by inhibiting Received 28 April, 2024 cyclooxygenase (COX) leads to the inhibition of prostaglandin synthesis, which is the Accepted 10 July, 2024 primary mechanism; still, there are other mechanisms of NSAIDs to produce these effects. Published online 23 September, 2024 In this study, we report a comparison between nimesulide (NIM) (selective COX-2) and Keywords: aspirin (ASP) (non-selective COX-2) inhibitors in mice model at the molecular level on the Analgesic caspase-3, peroxisome proliferation-activated receptors-alpha (PPAR-a) and gene Antipyretic expression of COX-2. Individual ED₁₀₀ doses of NIM and ASP (15.8 and 424.5 mg/kg, i.m.) Anti-inflammatory Apoptosis were injected in mice for 5 consecutive days with and without acetic acid. This study reports Cyclooxygenase-2 new pharmacological mechanisms for NIM and ASP to contribute as anti-inflammatory drugs by other important intracellular mechanisms. NIM outperforms ASP for caspase-3 Correspondence: inhibition in the kidney, liver, and plasma (anti-apoptotic and anti-inflammatory effects). T.A. Yahva Also, NIM was better than ASP in molecular effects at the PPAR-a reducing inflammation taimaaadlan1980@gmail.com and producing its antinociceptive effects. NIM and ASP in therapeutic doses reduced the COX-2 gene expression in the kidney (down-regulation), which causes the potentiation effect as an anti-inflammatory and analgesic drug. This research provided that NIM outperforms ASP in efficacy for the new mechanisms of NIM and ASP for inhibition of caspase-3 and binding with PPAR- α which worked as an anti-inflammatory action and reduced pain plus the suppression of COX-2 gene expression (down-regulation), which all contributing to reducing inflammation, pain and fever.

DOI: <u>10.33899/ijvs.2024.149230.3638</u>, ©Authors, 2024, College of Veterinary Medicine, University of Mosul. This is an open access article under the CC BY 4.0 license (<u>http://creativecommons.org/licenses/by/4.0/</u>).

Introduction

Nimesulide (NIM) belongs to the sulfonanilide group (*N*-(4-Nitro-2-phenoxyphenyl)-methanesulfonamide) of nonsteroidal anti-inflammatory drugs with selective cyclooxygenase enzyme (COX-2) inhibition thus preventing the production of prostaglandins that are responsible for producing pain, inflammation, and fever were produced through the COX-2 enzyme induction (1,2). NIM can prevent the inflammatory effects with fewer side effects, for example, gastrointestinal bleeding, ulcers, and renal prostaglandin involvement (3). NSAID inhibition of caspases is COX-independent. This is considered a new antiinflammatory mechanism (4). Caspases are well-known in cancer, inflammation, arthritis, and neurodegenerative disorders (5). Caspases are a family of genes essential for maintaining homeostasis through regulating cell death and inflammation (5). NIM reduces chondrocyte apoptotic cell death by inhibiting caspase-3. Thus, nimesulide may represent a new preventive option for osteoarthritis by blocking apoptotic events (6). Another study found that NIM plays a role in apoptosis by inhibiting prostaglandins, which modulate cell proliferation (7). Another molecular mechanism by which nimesulide exerts its antiinflammatory and analgesic effects is by inhibiting peroxisome proliferator-activated receptor alpha (PPAR- α)

and COX-2 mRNA expression, therapeutic concentrations of NIM have treated synovial osteoarthritis in human fibroblasts (8). NIM reduces cytokine-induced COX-2 expression at therapeutic doses (9) and inhibits expression levels of COX-2, both at the mRNA and protein levels (3), so this drug was shown to have anticancer effects in neoplastic pancreatic cells by inhibiting proliferation and inducing apoptosis (10). The efficacy of NIM on the COX-2 gene expression was reported in a previous study, which recorded that NIM inhibited cytokine-induced COX-2 expression and protein at both sub-therapeutic and therapeutic doses (9). The inhibition of COX-2 expression may play a role in tumor development (11). Acetylsalicylic acid, commonly known as aspirin (ASP), has been extensively used as an analgesic and anti-inflammatory medication for about ten decades. It acts by inhibiting the cyclooxygenase enzyme, which suppresses prostaglandin production and leads to relieving inflammation, pain, and fever (12). Recently, it has been recognized that the nuclear receptor peroxisome proliferator-activated receptor-alpha (PPAR- α) acts as a unique receptor of ASP; there are three subtypes of PPARs (PPAR- α , γ , β / δ) (13). PPAR- α exerts anti-inflammatory effects and inhibits the formation of macrophages by regulating gene expression; this receptor limits inflammation (14) and proved in other research that PPARs exert anti-inflammatory effects in rat models with carrageenan-induced paw inflammation (15). ASP also affects caspase-3 by decreasing rat neuron levels (16). Furthermore, it was found that ASP decreased apoptosis by reducing cleaved caspase-3 levels (17), in regard to ASP, at therapeutic concentrations, blocked COX-2 mRNA. These results propose that ASP produces its anti-inflammatory action by inhibiting the COX-2 induction, thereby suppressing the production of proinflammatory prostaglandins in mice at doses of 10-30 mg/kg orally and reducing human colon cancer by suppressing COX-2 expression (18) in other studies proved that ASP coadministered with cisplatin reduced COX-2 expression by inactivating NF-kB signaling in humans (19).

Our study confirms that NIM and ASP have molecular mechanisms to exert anti-inflammatory and analgesic effects through the actions of drugs on the caspase-3, PPAR- α , and COX-2 gene expression and proved that NIM (selective COX-2 inhibition) is more effective than ASP (non-selective COX) in treating inflammatory conditions. This study aimed to detect the new molecular mechanisms of NIM and ASP in reducing inflammation, thus promoting its effect as an analgesic and antipyretic drug.

Materials and methods

Ethical approve

The study was standardized (the animal use and experimental design) by the care committee affiliated with

the College of Veterinary Medicine, Mosul University, with approval code no. UM.VET.2022.076.

Animals

In this study, 78 Swiss albino mice, weighing 24-34 g, males and females, were used throughout the study. Animals were kept in the animal facility at the College of Veterinary Medicine, Mosul University, under standard conditions, including a 14/10 h dark/light cycle with $22\pm2^{\circ}$ C facility temperature. Food and water were provided according to standard protocols.

Drugs

Nimesulide (10%, Instant Pharmaceuticals, India) and aspirin (pure powder, Sanofi, France) were diluted with normal saline and administered via the intramuscular (i.m.) route at an injection dose of 5 ml/kg.

Blood and tissue collection

Blood samples were collected from the choroid venous sinus of mice using capillary tubes containing EDTA (anticoagulant) (20-23). For the caspase-3 test, the samples were incubated at room temperature for 10-20 minutes while the blood tube was placed at 2-8°C for 15 minutes within 30 min of collection (in the PPAR- α test). The tubes were centrifuged for 20 minutes at 3000 rpm. The supernatant was collected as plasma samples. Tissue samples were cut, weighed, and frozen at -20°C. PBS (PH=7.4) was added before homogenization, performed at 4°C. The supernatant was collected after centrifugation of the samples for 20 minutes at 3000 rpm. The aliquot was then used for the ELISA assay of caspase-3 and PPAR- α .

Measuring the analgesic ED₅₀ of NIM and ASP in mice

By using the up-and-down method (24), the ED_{50} of NIM and ASP was determined by using the thermal method (hot plate) at 56°C (25-28). Determination of the initial doses depended on the preliminary and previous studies (29-32). The decrease and increase in the dose were at a constant value for both drugs (33-39).

Effect of NIM and ASP on apoptosis in mice

The effects of NIM and ASP on apoptosis were compared by measuring the activity of Caspase-3 in plasma, liver, and kidney. The mice were divided into six groups, each containing 5 animals; the control group was injected with normal saline, while the second and third groups were injected with the ED100 of NIM and ASP i.m. (15.8 and 424.5 mg/kg, respectively). The fourth group was injected with 1% acetic acid i.p., and the fifth and sixth groups were injected with NIM+AA and ASP+AA, respectively. All groups were treated for 5 days (one dose daily). Then, 30 minutes after the injection on the fifth day, blood was collected from the eye, and the mice were euthanized. The liver and kidney were then harvested. The organs were stored at -20°C for examination in the laboratory using a specific ELISA kit for mouse caspase-3 (Elabscience, USA) (40).

Effect of NIM and ASP on PPAR-α in mice

The comparison between NIM and ASP on the peroxisome proliferator-activated receptor-alpha (PPAR- α) involved 6 groups, each containing 5 mice. The groups were similar to those in the previous experiment. After 5 days of daily administration of the treatments, plasma, liver, and kidney samples were collected as in the previous experiment and examined in the laboratory using a specific ELISA kit for mouse PPAR- α (Elabscience, USA) (40).

The inhibitory effects of NIM and ASP on the COX-2 gene expression in the kidney of mice

In this experiment, we examine the inhibitory effects of NIM and ASP on the COX-2 gene expression using the 6 groups, similar to the previous trial except that it assessed the COX-2 gene expression of the kidney tissue using a specialized kit for COX-2 gene expression in mice (Gena Bioscience, Germany) (40).

mRNA extraction based on kit instructions for determination of COX-2 gene expression

The mRNA extraction for assessment of COX-2 gene expression was conducted according to Archer (40).

Measurement of COX-2 mRNA contents in kidney tissue

The first step in determining gene expression through reverse transcription polymerase chain reaction (RT-PCR) was the quantification of the extracted RNA, which was measured using QubitTM equipment (Qubit Fluorometer, Invitrogen, USA). A high-sensitivity kit was used for the procedure, and the working solution was in a clean plastic tube. The final volume in each tube was 200 µL. The sample volume was 1-20 µL added to the 180-199 µL working solution, then mixed by vertexing 2 - 3 seconds and incubated at room temperature for 2 minutes. All samples were measured using a Qubit[®] Fluorometer. The same quantitative procedure for RNA quantification was used to quantify complementary DNA (cDNA).

Expression of COX-2 gene in kidney tissue

The RT-PCR kit (Bioron GmbH, Germany) was used for quantification of COX-2 gene according to Archer (40) and Rao et al., (41) by using RT-PCR apparatus (MiniAmp PlusTM Thermocycler, USA). The RT-PCR β -actin Forward / primer sequences 5' Reverse were 5' TTGTGATGGACTCCGGAGAC 3', TGATGTCACGCACGATTTCC 3', respectively and F-5' CCCCTCTCTACGCATTCTGT 3', R-5' TGGCAGAACGACTCGGTTAT 3' for COX-2 gene (Macrogen company, Korea). The PCR threshold standard curve was based on an exponential model of the initial phase

of a PCR run where template replication efficiency is constant from cycle to cycle. The electrophoresis (Cleaver Scientific, UK) was achieved on 2% agarose gel and the RT-PCR bands were 202 bp COX-2 cDNA and 186 bp β -action cDNA.

Statistical analysis

Statistical Package for the Social Sciences (SPSS) version 20 was used in statistical analysis. One-way analysis was the statistical test applied for parametric data, which was accomplished by comparing means. Further analysis was done using an unpaired T-test to compare the mean of the two resembling groups (42). The significance was at P<0.05.

Results

Measuring the analgesic ED₅₀ of NIM and ASP in mice

The dose of NIM injected i.m. for mice causing the analgesic response in 50% of the experimental animals was 7.9 mg/kg, and for ASP was 212.23 mg/kg.

Effect of NIM and ASP on apoptosis in mice

NIM and ASP significantly inhibited caspase-3 in the kidney, liver, and plasma in mice compared with control groups of normal saline (negative group) and acetic acid (positive group). These results reveal the effects of NIM and ASP on apoptosis because the inhibition of caspase 3 reduced apoptosis (cell proliferation), and NIM was more effective than ASP in inhibiting caspase-3 in the ED_{100} dose (Table 1).

Table 1: Concentration of caspase-3 in mice

Groups	Kidney	Liver	Plasma
Saline	2.83±0.15	3.70±0.46	3.93±0.80
NIM + NS	2.65 ± 0.17	2.34±0.27*	3.56±0.19
ASP + NS	2.76±0.32	3.43 ± 0.20^{a}	3.80 ± 0.25
Acetic acid	$3.43 \pm 0.18^{*}$	4.04±0.23	$4.37 \pm 0.25^{*}$
NIM + AA	$2.45 \pm 0.13^{+}$	$2.37 \pm 0.36^{+}$	3.41±0.36+
ASP + AA	$2.58 \pm 0.20^{+}$	$2.48 \pm 0.17^{+}$	3.78±0.18

Numbers were as mean \pm Std.E. 5 mice/group. P<0.05. *mean differs significantly from the normal saline group. + means differs significantly from the acetic acid group. a mean differs significantly from the NIM group in the same treatment paradigm.

Effect of NIM and ASP on PPAR-α in mice

Effects of NIM and ASP on the PPAR- α –mediated induction of COX-2 expression were inhibited by therapeutic concentrations ED ₁₀₀ of NIM and ASP, which were injected i.m. and measured in kidney, liver, and plasma by ELISA kit. NIM and ASP acted as ligand dependent receptor activators, inhibiting the PPAR- α (dependent- transactivation of the target genes belonging to COX-2), as in table 2.

Groups	Kidney	Liver	Plasma
Saline	376.6±27.1	365.2 ± 20.08	18.67±4.7
NIM + NS	353.0±26.7*	361.0±23.9*	9.9±1.5*
ASP + NS	346.1±3.8*	360.8±8.5*	9.7±1.3*
Acetic acid	349.8 ± 25.2	363.1±24.7	16.0 ± 2.6
NIM + AA	312.5±4.9+	352.2±11.9+	5.8±0.3+
ASP + AA	$315.8 \pm 23.8^+$	$356.4\pm20.4^+$	$5.1\pm0.9^{+}$

Table 2: Concentration of PPAR-α in mice

Numbers were as mean±Std.E. 5 mice/group. P<0.05. *mean differs significantly from the normal saline group. + means differs significantly from the acetic acid group.

Effects of NIM and ASP on the COX-2 gene expression

NIM and ASP affected the COX-2 gene expression by decreasing the gene expression of COX-2 in comparison with positive and negative control groups after treating the mice for 5 days (one dose daily), as in table 3 and figures 1-3.

Table 3: concentration of COX-2 gene in mice

Groups	Kidney	
Saline	417±30.90	
NIM + NS	412±3.61	
ASP + NS	410±14.73	
Acetic acid	468±48.12	
NIM + AA	390±4.04	
ASP + AA	400±22.50	

Numbers were as mean±Std.E. 3 mice/group.

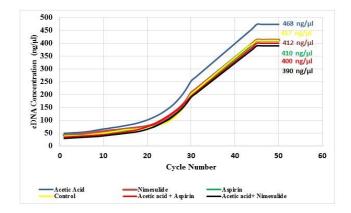


Figure 1: Comparison of the effectiveness between NIM and ASP on COX-2 gene expressions in the kidney tissue of mice using quantitative RT-PCR.

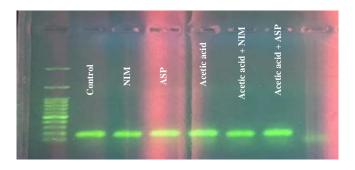


Figure 2: Positive β -*actin* located in lanes 1-2 (186 bp), positive COX-2 gene (202 bp) located in lanes 3-6, and negative control in lane 7. Lane M: 100 bp DNA ladder.

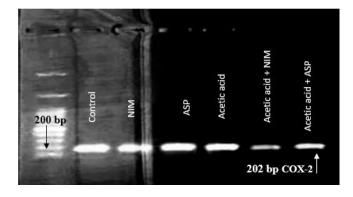


Figure 3: Kidney tissue COX-2 cDNA, RT-PCR product electrophoresis.

Discussion

The study aimed to estimate the drug comparison between NIM (selective COX-2 inhibitor) and ASP (nonselective COX inhibitor) at the level of effects on the caspase-3, PPAR-a, and COX-2 gene expression; in previous studies, there were conflicting results about effects of NIM and ASP on caspase-3 and PPAR-α. Therefore, there was confusion about the effects of these drugs and the molecular mechanisms, so this report explored the antiinflammatory mechanism of NIM and ASP COXindependent to show other behaviors of NSAID as antiinflammatory drugs, Caspases are members of cysteineaspartic protease, it plays a vital role in apoptosis and inflammation (40). When activation initiator caspases will be proteolytically activated and then activate executioner caspases (caspase-3, 6, and 7), which leads to immunological cell death ended by apoptosis (42). Caspase has a role in cancer, inflammation, arthritis, cardiovascular disease, and neurodegenerative disorders (43). NSAIDs reduction of caspases is action COX-independent, representing a novel anti-inflammatory mechanism in humans (44). A previous study proved that NIM decreased cleavage of caspase 3 in 5 days after being treated with the drug in humans (45), which was similar to the result found in this research. So, the

inhibition of caspase-3 leads to the inhibition of apoptosis, which is useful in treating cancer. ASP was recorded in the previous study to reduce etoposide-induced caspase-3 activation in hepatocellular carcinoma, and ASP has an antiapoptotic effect by blocking caspase 3 in humans (46). Also, other research reported that ASP showed significantly decreased expression of TNF- α , caspase-3, and apoptotic index (47), and these results are similar to what emerged from this research, where ASP decreases caspase-3, which is responsible as executioner caspase to generate apoptosis.

Peroxisome proliferator-activated receptors (PPARs) were known over ten years ago and were categorized as nuclear receptors. 3 PPAR subtypes have been discovered as PPAR- α , β/δ , γ , and the diverse PPAR subtypes have been revealed to play vital roles in important disorders like obesity, atherosclerosis, diabetes, fertility, and cancer. Most studies focus on the role of PPARs in inflammatory manners. Many studies have shown that agonists of PPAR- α and PPAR-y produce anti-inflammatory effects both in vivo and in vitro. Using the carrageenan-induced paw edema model for inflammation, a recent study presented that these agonists affect the initiation phase but not the late phase of the inflammatory process (15). PPAR- α has a role in inflammation by modulating inflammation. This is evidenced by the proposal of Leukotriene B4 (LTB4), a powerful chemotactic inflammatory eicosanoid (48). The binding to PPAR- α activates the transcription of genes of the ω - and β -oxidation pathways that can trigger the catabolism of LTB4 itself (49). Activation of PPAR- α by the NSAIDs donates to these medications' anti-inflammatory, antipyretic, and analgesic properties by activating the oxidative pathways involved in catabolizing the eicosanoids (50). Reduction of the proinflammatory molecules synthesis like IL-6 and prostaglandins seems to participate in PPAR- α -mediated anti-inflammatory effect through reduced activity of NF- κ B (51). A previous study found that NIM inhibited PPAR- α and γ isoforms agonist stimulation of the COX-2 expression and synthesis in humans (8).

This result agrees with our study on the effects of NIM on the PPAR-α by inhibiting it. The effects of ASP on PPAR- α in humans and mice registered in the previous study mentioned that a high dose of ASP reduced the expression of PPAR-α, suggesting a novel pharmacologic effect of ASP COX-independent (52). This also converges with our study, and these effects on the PPAR- α were given another mechanism as an anti-inflammatory-influenced, influenced un-depending on COX-2 inhibitor. With regards to the effects of NIM and ASP on gene expression of COX-2, in a previous study, NIM significantly decreased COX-2 gene expression in different doses 100,200, 400µmol/L led to significantly decreased pancreatic cancer group than those in the control group in humans (11), Also in other research, NIM caused down-regulation of the survivin and COX-2 expressions (at mRNA and protein levels) in FaDu cells that show a significant role in NIM-induced growth inhibition of hypopharyngeal carcinoma in human (53). Also, NIM was reported to significantly reduce COX-2, survivin, and PCNA expression levels at both the protein and mRNA levels in nude mice to treat laryngeal squamous cell carcinoma (3). Other research has proved that ASP suppressed COX-2 expression in humans and mice and has been useful in reducing human colon cancer (18). The recent study mentioned that molecular mechanisms of ASP seem to contribute to tumor suppression by inhibiting COX-2 gene expression to suppress the production of elevated neoplastic prostaglandins and dysregulation of the NF-kB signaling pathway, which produced inhibiting gene expression of COX-2 (19). Another study observed that ASP caused downregulation in the expression of COX-2, suggesting that COX-2 plays an important role in colorectal cancer (54). This report found that NIM and ASP suppression of COX-2gene expression in different levels, ASP had more effected than NIM on COX-2 gene expression in the negative group, still in the positive group (inducible inflammation) by acetic acid, NIM outperformed ASP in inhibition of the COX-2 gene expression because of the action of NIM (selective COX-2 inhibitor) appeared in inflammation or inducible inflammation when was injected NIM with AA. It is worth noting that for the first time, this research mentions the annotation of the COX-2 gene, whereas no annotation has been found in the gene bank of COX-2 in mice yet, so no exon or intron region has been established.

Conclusions

This research provided that NIM outperforms ASP in efficacy for the new mechanisms of NIM and ASP for inhibition of caspase-3 and binding with PPAR- α , which worked as an anti-inflammatory action and reduced pain. Plus, the suppression of COX-2 gene expression (down-regulation) all contributed to reducing inflammation, pain, and fever.

Acknowledgments

We thank the College of Veterinary Medicine/University of Mosul for providing the crucial equipment for conducting this research.

Conflict of interest

The authors declare there is no conflict of interest.

References

- Gunaydin C, Bilge SS. Effects of nonsteroidal anti-inflammatory drugs at the molecular level. Eurasian J Med. 2018;50(2):116-121. DOI: 10.5152/eurasianjmed.2018.0010
- Caiazzo E, Ialenti A, Cicala C. The relatively selective cyclooxygenase-2 inhibitor nimesulide: What's going on?. Eur J Pharmacol. 2019;848:105-111. DOI: <u>10.1016/j.ejphar.2019.01.044</u>

- Liang Z, Liu J, Li L, Wang H, Zhao C, Jiang L, Qin G. Effect of nimesulide on the growth of human laryngeal squamous cell carcinoma. Am J Otolaryngol. 2014;35(2):120-129. DOI: 10.1016/j.amjoto.2013.10.009
- Smith CE, Soti S, Jones TA, Nakagawa A, Xue D, Yin H. Non-steroidal Anti-inflammatory drugs are caspase inhibitors. Cell Chem Biol. 2017;24(3):281-292. DOI: <u>10.1016/j.chembiol.2017.02.003</u>
- McIlwain DR, Berger T, Mak TW. Caspase functions in cell death and disease. Cold Spring Harb Perspect Biol. 2013;5(4):a008656. DOI: 10.1101/cshperspect.a008656
- Mukherjee P, Rachita C, Aisen PS, Pasinetti GM. Non-steroidal antiinflammatory drugs protect against chondrocyte apoptotic death. Clin Exp Rheumatol. 2001;19:7-11. [available at]
- Shaik MS, Chatterjee A, Singh M. Effect of a selective cyclooxygenase-2 inhibitor, nimesulide, on the growth of lung tumors and their expression of cyclooxygenase-2 and peroxisome proliferatoractivated receptor-gamma. Clin Cancer Res. 2004;15;10(4):1521-9. DOI: <u>10.1158/1078-0432.ccr-0902-03</u>
- Kalajdzic T, Faour WH, He QW, Fahmi H, Martel-Pelletier J, Pelletier JP, Di Battista JA. Nimesulide, a preferential cyclooxygenase 2 inhibitor, suppresses peroxisome proliferator-activated receptor induction of cyclooxygenase 2 gene expression in human synovial fibroblasts: Evidence for receptor antagonism. Arthritis Rheum. 2002;46(2):494-506. DOI: 10.1002/art.10055
- Fahmi H, He Y, Zhang M, Martel-Pelletier J, Pelletier JP, Di Battista JA. Nimesulide reduces interleukin-1beta-induced cyclooxygenase-2 gene expression in human synovial fibroblasts. Osteoarthritis Cartilage. 2001;9(4):332-40. DOI: <u>10.1053/joca.2000.0393</u>
- Ferreira RG, Narvaez LM, Espíndola KM, Rosario AS, Lima WN, Monteiro MC. Can nimesulide nanoparticles be a therapeutic strategy for the inhibition of the KRAS/PTEN signaling pathway in pancreatic cancer?. Front Oncol. 2021;11:594917. DOI: 10.3389/fonc.2021.594917
- Chu M, Wang T, Sun A, Chen Y. Nimesulide inhibits proliferation and induces apoptosis of pancreatic cancer cells by enhancing expression of PTEN. Exp Ther Med. 2018;16(1):370-376. DOI: <u>10.3892/etm.2018.6191</u>
- Patel D, Roy A, Kundu M, Jana M, Luan CH, Gonzalez FJ, Pahan K. Aspirin binds to PPARα to stimulate hippocampal plasticity and protect memory. Proc Nat Acad Sci USA. 2018;115(31):7408-7417. DOI: 10.1073/pnas.1802021115
- Patel D, Roy A, Pahan K. PPARα serves as a new receptor of aspirin for neuroprotection. J Neurosci Res. 2020;98(4):626-631. DOI: <u>10.1002/jnr.24561</u>
- Zandbergen F, Plutzky J. PPAR alpha in atherosclerosis and inflammation. Biochem Biophys Acta. 2007;1771(8):972-982. DOI: 10.1016/j.bbalip.2007.04.021
- Delerive P, Fruchart JC, Staels B. Peroxisome proliferator-activated receptors in inflammation control. J Endocrinol. 2001;169(3):453-9. DOI: <u>10.1677/joe.0.1690453</u>
- Ozdemir E, Avcı O, Inan ZDS, Taskiran AS, Gunes H, Gursoy S. Aspirin attenuates morphine antinociceptive tolerance in rats with diabetic neuropathy by inhibiting apoptosis in the dorsal root ganglia. Metab Brain Dis. 2023;38(6):2145-2158. DOI: <u>10.1007/s11011-023-01226-2</u>
- Liu PP, Liu HH, Sun SH, Shi XX, Yang WC, Su GH, Zhao J. Aspirin alleviates cardiac fibrosis in mice by inhibiting autophagy. Acta Pharmacol Sin. 2017;38(4):488-497. DOI: <u>10.1038/aps.2016.143</u>
- Xu XM, Sansores-Garcia L, Chen XM, Matijevic-Aleksic N, Du M, Wu KK. Suppression of inducible cyclooxygenase 2 gene transcription by aspirin and sodium salicylate. Proc Nat Acad Sci USA. 1999;96(9):5292-5297. DOI: <u>10.1073/pnas.96.9.5292</u>
- Jiang W, Yan Y, Chen M, Luo G, Hao J, Pan J, Hu S, Guo P, Li W, Wang R, Zuo Y, Sun Y, Sui S, Yu W, Pan Z, Zou K, Zheng Z, Deng W, Wu X, Guo W. Aspirin enhances the sensitivity of colon cancer cells to cisplatin by abrogating the binding of NF-κB to the COX-2 promoter. Aging (Albany, NY). 2020;12(1):611-627. DOI: <u>10.18632/aging.102644</u>

- Abdulrazaq H, Ameen Q. Genetic relationship between local guinea fowl, quail and chicken using RAPD–PCR technique. Mesop J Agric. 2023;51(4):39-49. DOI: 10.33899/mja.2023.142638.1265
- Ami S, Guri R. EXISTENCE, MOLECULAR IDENTIFICATION AND GENETIC VARIATION OF NEW ISOLATES OF SEED GALL NEMATODE Anguina tritici PARASITIZING ON WHEAT AND BARLEY IN IRAQ. Mesop J Agric. 2023;51(1):32-47. DOI: 10.33899/magrj.2023.135099.1197
- 22. Ahmed S, Abdul-Rahman S. EFFECTS OF CASTRATION AND SEX HORMONES ON ANTIOXIDANT STATUS AND SOME BIOCHEMICAL PARAMETERS OF MALE RABBITS EXPOSED TO OXIDATIVE STRESS. Mesop J Agric, 2023;51(1):92-114. DOI: 10.33899/magrj.2023.138538.1220
- Qader G, Tayeb I. EFFECT OF MEDICINAL PLANTS AND VITAMIN E ON PRODUCTIVE PERFORMANCE, SOME PHYSIOLOGICAL, IMMUNOLOGICAL PARAMETERS AND ANTIOXIDANT STATUS OF BROILER UNDER COLD STRESS. Mesop J Agric, 2024;52(1):60-78. DOI: 10.33899/mja.2024.143936.1282
- 24. Dixon WJ. Efficient analysis of experimental observations. Annu Rev Pharmacol Toxicol. 1980;20:441-62. DOI: 10.1146/annurev.pa.20.040180.002301
- Yahya TA, Mousa YJ. Comparison of the analgesic, antipyretic, and anti-inflammatory efficiency between nimesulide and aspirin in mice. Egypt J Vet Sci. 2024;55(6):1697-1703. DOI: 10.21608/EJVS.2024.253221.1698
- Mousa YJ, Mahmood MB, Mohammed ZT. Simultaneous determination of pethidine pharmacokinetics in rats: The impact of tramadol coadministration. Iraqi J Vet Sci. 2023;37(1):259-265. DOI: 10.33899/ijvs.2022.133824.2307
- Mousa YJ. Effect of chlorpheniramine on acute dichlorvos poisoning in chicks. Iraqi J Vet Sci. 2009;23(2):35-43. DOI: 10.33899/ijvs.2009.5738
- Al-Zubaidy MH, Mousa YJ, Hasan, MM, Mohammad FK. Acute toxicity of veterinary and agricultural formulations of organophosphates dichlorvos and diazinon in chicks. Arh Hig Rada Toksikol. 2011;62:317-323. DOI: <u>10.2478/10004-1254-62-2011-2139</u>
- Mohammad FK, Mousa YJ, Hasan MM. Acute toxicity and neurobehavioral effects of diphenhydramine in chicks. J Poult Sci. 2012;49(1):51-56. DOI: <u>10.2141/jpsa.011050</u>
- Mousa YJ, Mahmood MB, Mohammad MS. Administration of ketamine with the central and peripheral analgesics for induction of balanced anesthesia in the chicks. IOP Conf Ser Earth Environ Sci. 2019;388:012021. DOI: <u>10.1088/1755-1315/388/1/012021</u>
- Mousa YJ. Effect of nefopam in normal chickens and its relationship to hydrogen peroxide-induced oxidative stress. Iraqi J Vet Sci. 2021;35(I):7-12. DOI: <u>10.33899/ijvs.2021.127013.1433</u>
- Mousa YJ, Mahmood MB. Effect of meloxicam coadministration on the anaesthetic potency of thiopental sodium in a chick model. Vet Stanica. 2022;53(2):155-163. DOI: <u>10.46419/vs.53.2.5</u>
- Mousa Y. Etomidate anesthesia in chicks: Effect of xylazine. J Hell Vet Med Soc. 2020;71(4):2463-2470. DOI: <u>10.12681/jhvms.25921</u>
- Jassim OY, Mousa YJ. Nefopam and ketorolac: Isobolographic analysis of analgesic effect and pharmacokinetic profile in chicks. Iraqi J Vet Sci. 2022;36(1):145-150. DOI: <u>10.33899/ijvs.2021.129540.1660</u>
- Mousa YJ, Khalil KA, Mahmood MB. Interaction of meloxicam and phenylbutazone on the level of cyclooxygenase-2 in mice. Iraqi J Vet Sci. 2022;36(4):1011-1016. DOI: <u>10.33899/ijvs.2022.132859.2140</u>
- Abdulah RL, Mousa YJ. Effects of hydrogen peroxide-induced oxidative stress on the plasma concentration and pharmacokinetics of ketorolac in chicks. Iraqi J Vet Sci. 2023;37(1):83-88. DOI: 10.33899/ijvs.2022.133592.2260
- Abdulhamid SK, Mousa YJ. The pharmacokinetics of phenylbutazone and its interaction with dexamethasone in chicks. Iraqi J Vet Sci. 2023;37(1):137-142. DOI: <u>10.33899/ijvs.2022.133338.2209</u>
- Albadrany YM, Naser AS, Shaban KA. Detection of potential effects of orphenadrine upon anesthesia with propofol and/or thiopental in mice. Iraqi J Vet Sci. 2024;38(1):239-243. DOI: 10.33899/ijvs.2023.141780.3135

- Naser AS, Albadrany YM, Abdullah MA. Evaluation of the advantages of orphenadrine in anaesthesia caused by ketamine in mice. Iraqi J Vet Sci. 2024;38(1):233-238. DOI: <u>10.33899/ijvs.2023.141516.3119</u>
- Archer BG. Note on the PCR threshold standard curve. BMC Res Notes. 2017;10(1):731. DOI: <u>10.1186/s13104-017-3036-4</u>
- 41. Rao X, Huang X, Zhou Z, Lin X. An improvement of the 2[^] (-delta delta CT) method for quantitative real-time polymerase chain reaction data analysis. Biostat Bioinforma Biomath. 2013;3(3):71-85. [available at]
- Petrie A, Watson P. Statistics for veterinary and animal science. USA: John Wiley and Sons; 2013. 391 p. [available at]
- Lamkanfi M, Dixit VM. Inflammasomes and their roles in health and disease. Annu Rev Cell Dev Biol. 2012;28:137-161. DOI: <u>10.1146/annurev-cellbio-101011-155745</u>
- Lamkanfi M. Emerging inflammasome effector mechanisms. Nat Rev Immunol. 2011;11(3):213-220. DOI: <u>10.1038/nri2936</u>
- Paul AG, Sharma-Walia N, Chandran B. Targeting KSHV/HHV-8 latency with COX-2 selective inhibitor nimesulide: A potential chemotherapeutic modality for primary effusion lymphoma. PLoS One. 2011;6(9):e24379. DOI: <u>10.1371/journal.pone.0024379</u>
- 46. Feng X, Lu B, Xu Y, Li Q, Zhou W, Yang Z, Yang Z, Zhao W, Shen Z, Hu R. Aspirin reduces the apoptotic effect of etoposide via Akt activation and up-regulation of p21(cip). Int J Mol Med. 2011;28(4):637-643. DOI: <u>10.3892/ijmm.2011.713</u>
- 47. Akhmad YP, Bremmy L, Prima NF, Anita DA, Sofie RK, Debbie SR, Ani Melani M, Erlina W. Effects of low dose aspirin on caspase 3, TNFα and Apoptotic index levels in preclampsia maternal serum-induced placental trophoblast cell line in vitro. Int J PharmTech Res. 2016;9(11):47-53. [available at]
- Chinetti G, Fruchart JC, Staels B. Peroxisome proliferator-activated receptors (PPARs): Nuclear receptors at the crossroads between lipid metabolism and inflammation. Inflamm Res. 2000;49(10):497-505. DOI: <u>10.1007/s000110050622</u>
- Poynter ME, Daynes RA. Peroxisome proliferator-activated receptor alpha activation modulates cellular redox status, represses nuclear factor-kappaB signaling, and reduces inflammatory cytokine production in aging. J Biol Chem. 1998;273(49):32833-32841. DOI: 10.1074/jbc.273.49.32833
- Ivan T, Andreas W, Wilfried S, Philipp E, Salvatore C, Bernhard F. Aspirin regulates expression and function of scavenger receptor-BI in macrophages: studies in primary human macrophages and in mice. FASEB J. 2006;20(9):1328-1335. DOI: <u>10.1096/fj.05-5368com</u>
- Tager AM, Bromley SK, Medoff BD, Islam SA, Bercury SD, Friedrich EB, Carafone AD, Gerszten RE, Luster AD. Leukotriene B4 receptor BLT1 mediates early effector T cell recruitment. Nat Immunol. 2003;4(10):982-90. DOI: <u>10.1038/ni970</u>
- Devchand PR, Keller H, Peters JM, Vazquez M, Gonzalez FJ, Wahli W. The PPARalpha-leukotriene B4 pathway to inflammation control. Nature. 1996;384(6604):39-43. DOI: <u>10.1038/384039a0</u>
- Jia-Jun T, Su-Mei L, Liang Y, Ju-Ke M, Ya-Kui M, Hai-Bo W, Wei X. Nimesulide inhibited the growth of hypopharyngeal carcinoma cells via suppressing Survivin expression. Head Neck Oncol. 2012;4:7. DOI: 10.1186/1758-3284-4-7
- 54. Ramzy MM, Abdelwahab SA. The Effect of aspirin on survivin and COX-2 expression and their correlation in aberrant crypt foci. Bull Egypt Soc Physiol Sci. 2010;30(1):207-220. DOI:<u>10.21608/BESPS.2010.36174</u>

التأثيرات الجزيئية للنيميسولايد والاسبرين على أنزيم الكاسبيز-٣، مستقبلات البيروكسيسوم المنشط- الفا والتعبير الجيني لأنزيم الأكسدة الحلقية-٢ في الفئران

تيماء عدلان يحيى و يعرب جعفر موسى

فرع الفسلجة والكيمياء الحياتية والأدوية، كلية الطب البيطري، جامعة الموصل، الموصل، العراق

الخلاصة

تتميز مضادات الالتهاب غير الستيرويدية بتقليل الألم والالتهاب وخافض للحرارة عن طريق تثبيط إنزيم الأكسدة الحلقية الذي يؤدي إلى تثبيط تخليق البر وستوكلاندين المسؤول عن الألم الالتهابي، وكانت الآلية الأساسية لعلاج حالات الالتهاب، ولكن كانت هناك آليات أخرى لعلاج الألم. في هذه الدراسة نورد مقارنة بين دواء النيميسولايد (المثبط الانتقائي) والأسبرين (المثبط غير الانتقائي) لأنزيم الأكسدة الحلَّقية-٢ في نموذج الفئر إن عند مستوى التأثير إت الجزيئية على أنزيم الكاسبيز -٣، مستقبلات البير وكسيسوم المنشط-الفا فضلا عن التعبير الجيني لأنزيم. الأكسدة الحلقية-٢. تم حقن الفئر إن بجر عة فر دية من كل من النيميسو لايد والأسبرين ١٥٫٨ و ٤٢٤,٥ ملغم/كغم، في العضل لمدة ٥ أيام متتالية مع أو بدون حمض الخليك. سجل لدينا هدفا دوائيا جديدا للنيميسو لايد والأسبرين للمساهمة كأدوية مضادة للالتهابات من خلال آليات مهمة أخرى داخل الخلايا، إذ تفوق النيميسولايد على الأسبرين في تأثيره المثبط لأنزيم الكاسبيز -٣ في الكلي والكبد والبلاز ما والذي تم تسجيله في هذه الدر اسة ويؤدي إلى تقليل الالتهاب، كما كان النيميسو لايد افضل من الأسبرين في التأثير التثبيطي على مستقبلات البير وكسيسوم المنشط-الفا مما يؤدى ألى تقليل الالتهاب وعمل كل من النيميسو لايد والأسبرين وبالجرع العلاجية على التقليل من التعبير الجيني لأنزيم الأكسدة الحلقية-٢ في الكلي وتسببت في تقليل الالتهاب. قدم هذا البحث آلية جديدة لكل من ألنيميسولايد والاسبرين عن طريق تثبيط الكاسبيز-٣ ومستقبلات البير وكسيسوم المنشط-الفا والذي يعمل كمضاد للالتهابات ويقلل الألم بالإضافة إلى التقليل من التعبير الجيني المستحث لأنزيم الأكسدة الحلقية-٢ (التقليل من عدد المستقبلات)، والتي تساهم جميعها في تقليل الالتهاب والألم والحمي.