Determination of blood cholinesterase level by electrometric method and inhibition by diazionon, dichlorvous and cypermethrin in vivo in homer pigeon (racing pigeon)

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Abstract

An environmental pollution with commonly used insecticides effected the homer pigeon this study used dichlirvous, diazinon and cypermethrin in several subacute doses The enzyme activity was measured by modified electometric method as $\Delta pH/30$ min. The Confidence interval in the plasma were 0.197 respectively and those of the erythrocytes were 0.934 respectively. The organophosphorus insecticides diazinon at dose (5-10) mg\ kg b.w. significantly inhibited plasma (28~53%) and erythrocyte 53% and dichlorvous at dose (5-10) mg\kg b.w. significantly inhibited plasma (33~38%) and erythrocyte 52% cholinesterase *in vitro* in a concentration-dependent tanner. The insecticides cypermethrin at dose (20-40) mg\ kg b.w. significantly inhibited plasma lactic dehydrogenase in (4~9.5%).

تحديد فعالية انزيم الكولين استراز بالدم بالطريقة الكهرومترية بواسطة التثبيط بالديازينون والدايكلورفس والسايبر مثرين في داخل الجسم بطائر الزاجل

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الخلاصة

ان التلوث البيئي المصاحب لاستعمال المبيدات الحشرية الشائعة الاستعمال لها تأثير ضار لطائر الزاجل الشتملت هذه الدراسة على عدة مبيدات بالديازينون الدايكلورفس والسابير مثرين بجرعات تحت الحادة متعدد وتعقياس مستوى انزيم الكولين استراز بواسطة الطريقة الكهرومترية المحورة من خلال تغير الباء هاء في 37 درجة مئوية حيث كان معامل الاختلاف في البلازما 0.179 وفي كريات الدم الحمراء 0.934، ان المبيد الحشري الديازينون ثبط الانزيم معنويا بنسبة (33–38%) في البلازما وبنسبة 52% في كريات الدم الحمراء بجرعة (5-10) ملغم/ كغم من وزن الجسم، أما المبيد الدايكلورفس ثبط الانزيم معنويا بنسبة (28–39%) في البلازما وبنسبة 53% في كريات الدم الحمراء بجرعة (5-10) ملغم/ كغم من وزن الجسم، حيث أدى إعطاء المبيد الساييرمثرن إلى تثبيط إنزيم اللاكتيك ديهييدروجنيز معنويا بنسبة (5.9-4%) في البلازما وبنسبة 53% في كريات الدم الحمراء بجرعة (5-10) ملغم/ كغم من وزن الجسم.

Introduction

Inhibition of plasma or brain cholinesterase activity is a biomarker endpoint of exposure of wild birds to organophosphate and cypermethrin (1, 2, 3, 4, 5). Organophosphate insecticides are extensively used in public health, veterinary practice and agriculture (6, 7). These insecticides induce of enzyme cholinesterase with subsequent accumulation of acetylcholine at the nerve terminals and neuromuscular junctions (3, 8, 4). Birds are naturally deficient in cholinesterase activity in erythrocytes. Therefore the

diagnosis and monitoring of exposure to anticholinesterase insecticides rely on the measurement of plasma and erythrocytes cholinesterase activities in wild birds (1, 9, 10, 11, 12). The used an electrometric method for measurement of plasma and erythrocytes cholinesterase activity in several animal species. Application of an electrometric method for measuring plasma and tissue cholinesterase activities in four indigenous birds, The method of *in vivo* cholinesterase inhibition shown aspect intoxication by organophosphrous insecticide (13, 14, 15, 16).

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Materials and Methods

- **Birds:** The birds Homer pigeon used in the present study were apparently healthy adults of both sexes. The birds were obtained from regions in which there were no activities of insecticide application for at least two months. The birds of each species were separately kept in captivity cages (80 × 60 × 70 cm) at about 25 °C with water and poultry feed for 14 days before experiments. Blood samples were collected using EDTA's (AFCO, Jordan) test tubes. Plasma was separated from blood by centrifugation at 3.000 rpm (Hettich, Germany) for 15 min.
- An electrometric method for measurement of cholinesterase activity: A pH 8.1 barbital-phosphate buffer solution (1.237 g sodium barbital, 0.163 g potassium dihydrogen phosphate and 35.07 g sodium chloride/L of distilled water) (15, 16). We determined cholinesterase activity in the plasma and RBCs samples by an electrometric method (17). The reaction mixture of the enzyme in a 10-ml vial contained 3 ml of the pH 8.1 barbital-phosphate buffer, 0.2 ml plasma or whole brain homogenate and 3 ml of distilled water. Initial pH of the mixture (pH 1) was measured by pH meter (Hanna, China), and then 0.12 ml of the aqueous solution of the substrate 7.5% acetylthiocholine iodide was added to the mixture as substrate which was incubated at 37 °C for 30 min. At the end of the incubation period, the pH of the reaction mixture (pH 2) was measured. The enzyme activity expressed as ΔpH/30 min was calculated as follows: Cholinesterase activity:

 $(\Delta pH/30 \text{ min.}) = (pH 1-pH 2) - \Delta pH \text{ of blank } (15, 16).$

- Lactic dehydrogenase activity by cypermethrin: This method is based on colormetric method by using lactic dehydrogenase kit (Biolab, France) measured in computerized U. V. spectrophotometer (Labmed, USA) the kit prepare to zero conc. of cypermethrin as control and the residual cypermethrin activity in the mixture was measure as before.
- **Statistics:** The data were presented as means + SE and they were subjected to analysis of variance followed by the least significant difference test (18). The accepted level of statistically significant difference level was at p<0.05.

Results

- **Preliminary reference ChE activity:** A table 1 shows the normal ChE values, 95% confidence interval and related statistic for plasma and erythrocyte ChE activity of 12 homer pigeon. Preliminary reference values of the main cholinesterase activity (Δ pH/ 30 min) and confidence in the plasma were 0.197, respectively, and those of the erythrocyte were 0.943, respectively (Table 1). Erythrocyte ChE activity was significantly higher than that of the plasma.

Table (1) Preliminary reference cholinesterase activity (Δ PH/ 30 min) in the plasma and erythrocyte of homer pigeon

	3	1 9
Sample	Plasma	Erythrocytes
Mean	0.298	0.492
Standard error	0.14	0.024
Standard deviation	0.683	0.983
Confidence	0.197	0.943

- In vivo ChE inhibition:

1. The insecticide dichlorvous significantly and in a dose-dependent inhibited plasma (28~39%) and erythrocyte 53% ChE activity *in vitro* (Table 2).

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Table (2) *In vivo* inhibition of homer pigeon plasma and erythrocyte cholinesterase (ChE) by dichloryous (mean \pm SE)

Toxic dose	Plasma ChE		Erythrocyte ChE	
mg\kg b.w.	ΔPH / 30 min	% inhibition	ΔPH / 30 min	% inhibition
0 mg\kg b.w.	0.21 ± 0.018	0	0.39 ± 0.091	0
2.5mg\kg b.w.	0.05 ± 0.1	16	0.32 ± 0.022	19
5 mg\kg b.w.	$0.03 \pm 0.08*$	28	0.37 ± 0.05	35
10 mg\kg b.w.	0.02 ± 0.41 *	39	0.21 ± 0.6 *	53

*significantly different from the respective control (0 mg/kg b.w.) p<0.05

2. The insecticide diazinon significantly and in a dose -dependent inhibited plasma (33~38%) and erythrocyte 52% ChE activity *in vitro* (Table 3).

Table (3) *In vivo* inhibition of homer pigeon plasma and erythrocyte cholinesterase (ChE) by diazinon (mean \pm SE)

Toxic dose	Plasma ChE		Erythrocyte ChE	
mg\kg b.w.	$\Delta PH / 30 min$	% inhibition	ΔPH / 30 min	% inhibition
0 mg\kg b.w.	0.19 ± 0.033	0	0.25 ± 0.05	0
2.5mg\kg b.w.	0.12 ± 0.05	19	0.29 ± 0.31	21
5 mg\kg b.w.	$0.09 \pm 0.09*$	33	0.2 ± 0.18	33
10 mg\kg b.w.	$0.05 \pm 0.022*$	38	0.19 ± 0.096 *	52

*significantly different from the respective control (0 mg/kg b.w.) p<0.05.

3. The insecticide cypermethrin significantly and in a dose -dependent inhibited plasma (4~9.5%) lactic dehydrogenase activity *in vitro* (Table 4).

Table (4) *In vivo* inhibition of homer pigeon of plasma lactic dehydrogenase by cypermethrin

Toxic dose	Plasma ChE		
mg∖kg b.w.	Absorption	% inhibition	
0 mg\kg b.w.	7.65 ± 0.27	0	
20 mg\kg b.w.	$3.94 \pm 0.124*$	4	
40 mg\kg b.w.	$2.91 \pm 0.021*$	9.5	

^{*}significantly different from the respective control (0 mg/kg b.w.) p<0.05.

Discussion

The electrometric method is the common diagnosing organophosphate and carbamate poisoning in birds, It is therefore important to measure homer pigeon cholinesterase activities periodically to assess any environmental contamination by insecticides (9, 15, 20). The present electrometric technique for measurement of cholinesterase activity was used to assess poisoning induced by organophosphate and carbamate insecticides in chickens (15, 17, 21). The variations in the plasma and erythrocytes cholinesterase activities primarily in homer pigeon (17, 20, 22). The technique of *in vitro* cholinesterase inhibition by organophosphate insecticides can be used to assess the potential toxicity of anticholinesterase chemicals (17, 23). Organophosphate and carbamate insecticides in the avian species as well as in mammals primarily inhibit nervous system cholinesterases with subsequent development of nicotinic, muscarinic and central nervous toxicities (1, 4, 9). The inhibiton by cypermethrin *in vitro* effective to several enzymes present in plasma especially lactic dehydrogenase, it is necessary for aerobic metabolic path ways for conversion of lactic acid to pyrovate. (20) in addition it's effect on CYP450 in the liver. Their activities could be routinely measured using the described electrometric method to assess exposure of wild birds to agricultural insecticides polluting the environment.

References

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- Wilson, B. W.; McCurdy, S. A.; Henderson, J. D.; McCarthy, S. A. & Billiti, J. E. 1998. Cholinesterases and agriculture. Humans, laboratory animals, wildlife. In: Doctor BP, ed. Structure and function of cholinesterases and related proteins. Plenum Press, New York, USA, PP. 539-546.
- York, USA, PP. 539-546.

 2. Cocker, J.; Mason, H. J.; Garfitt, S. J. & Jones, K. 2002. Biological monitoring of exposure to organophosphate pesticides. Toxicol. Lett., 134: 97-103.
- 3. Kwong, T. Č. 2002. Organophosphate pesticides: biochemistry and clinical toxicology. Therap. Drug Mon, 24: 144-149.
- 4. Wilson, B. W. 2005. Cholinesterase inhibition. In: Wexler P, ed. Encyclopedia of toxicology. 2nd ed. Vol. 1. Elsevier, New York, NY, USA, PP. 588-599.
- 5. Fairbrother, A. & Bennett, J. K. 1988. The usefulness of cholinesterase measurements. J. Wildlife Dis., 24: 587-590.
- 6. Coggon, D. 2002. Work with pesticides and organophosphate sheep dips. Occupational Medicine (London) 52: 467-470.
- 7. Jaga, K. & Dharmani, C. 2003. Sources of exposure to and public health implications of organophosphate pesticides. Rev. Panam. Sal. Pb., 14: 171-185.
- 8. Parsons, K. C.; Matz, A. C.; Hooper, M. J. & Pokras, M. A. 2000. Monitoring wading bird exposure to agricultural chemicals using serum cholinesterase activity. Environ. Toxicol. Chem., 19: 1317-1323.
- 9. Cairns, M. A.; Maguire, C. C. & Williams, B. A. 1991. Brain cholinesterase activity of bobwhite acutely exposed to chlorpyrifos. Environ. Toxicol.Chem., 10:657-664.
- 10.Blakely, B. R. & Yole, M. J. 2002. Species differences in normal brain cholinesterase activities of animals and birds. Vet. Hum. Toxicol., 44: 129-132.
- 11.Iko, W. M.; Archuleta, A. S. & Knop, F. L. 2003. Plasma cholinesterase levels of mountain plovers (CHARADRIUS MONTANUS) wintering in central California, USA. Environ. Toxicol. Chem., 22: 119-125.
- 12. Claudie, R.; Grolleau, G.; Chamoulaud, S. & Riviere, J. 2005. Plasma B-esterase activities in European raptors. J. Wildlife Dis., 41: 184-208.
- 13.Long, H.; Kirrane, B.; Nelson, L. S. & Hoffman, R. S. 2003. Crabaryl inhibition of plasma cholinesterase activity. J. Toxicol. Clin. Toxicol., 41: 737-740.
- 14.Khan, A. A.; Coppock, R. W.; Schuler, M. M. & Lillie, L. E. 1990. Effects of dichlorvos on blood cholinesterase activities of cattle.Am.J.Vet.Res.,51: 79-82.
- 15.Mohammad, F. K.; Faris, GA-M. & Al-Kassim, N. A. 1997. A modified electrometric method for measurement of erythrocyte acetylcholinesterase activity in sheep. Vet. Hum. Toxicol., 39: 337-339.
- 16.Mohammad, F. K.; Al-Baggou, B. K.; Alias, A. S. & Faris, GA-M. 2006. Application of an electrometric method for measurement of in vitro inhibition of blood cholinesterases from sheep, goats and cattle by dichlorvos and carbaryl. Vet. Med. (Czek), 51: 45-50.
- 17. Mohammad, F. K.; Alias, A. S. & Ahmed, O. A. H. 2007. Electrometric measurement of plasma, erythrocyte and whole blood cholinesterase activities in healthy human volunteers. J. Med. Toxicol., 3: 25-30.
- Med. Toxicol., 3: 25-30.

 18.Petrie, A. & Watson, P. 1999. Statistics for veterinary and animal science. Blackwell Science Ltd, Oxford, UK.
- 19. Hooper, M. J. 1988. Avian cholinesterases: Their characterization and use in evaluating organophosphate insecticide exposure. Dissert. Abst. Int., 6: 2068.
- 20.Fossi, M. C.; Lari, L. & Casini, S. 1996. Interspecies variation of 'B' esterases in birds: the influence of size and feeding habits. Arch. Environ. Contam. Toxicol., 31: 525-532.
 21.Mohammad, F. K. & Al-Baggou, B. 2005. Electrometric cholinesterase determination in the
- 21. Mohammad, F. K. & Al-Baggou, B. 2005. Electrometric cholinesterase determination in the chicken treated with dichlorvos and carbaryl. Online J. Vet. Res., 9: 1-5.
- 22.Hill, E. F. & Murray, H. C. 1987. Seasonal variation in diagnostic enzymes and biochemical constituents of captive northern bobwhites and passerines. Comp. Bioch. Physiol., 87B: 933-940.
- 23. Iyaniwura, T. T. 1990. *In vitro* toxicology of organophosphorus pesticide combinations. *In vitro* Toxicol., 3: 373-377.