## BACTERIAL QUALITY OF BEEF CARCASSES AND SANITARY CONDITION OF BUTCHER'S SHOPS IN BASRAH CITY

Ala AL-Deen H. Jawad \*, Alaa T. Abdul Alwahid\*\*

Department of Physiology, College of Veterinary Medicine, University of Basrah, Basrah, Iraq Department of Microbiology, College of Veterinary Medicine, University of Basrah, Basrah, Iraq

(Received 22 April 2006, Accepted 13 January 2007)

**Keywords:** Bacteria, Beef, Butchers.

#### **ABSTRACT**

The microbial quality of beef carcasses, sanitary conditions in butchers' shops and possibility of the presence of human pathogens associated with food poisoning outbreaks such as salmonella and staphylococci was investigated in 160 samples of beef carcasses, cutting blocks, knives, workers' hands and air (32 samples for each) during January and ended in June. All samples examined bacteriologically for aerobic plate count, total coliform count, *Staphylococcus aureus* count and an attempt was made for isolation and identification Salmonella spp.

This study have shown that there was a gradual increment in the count of all microorganisms starting in January and ending with June, the minimum mean of aerobic plate count of beef carcasses was  $27.32 \times 10^4$  cfu/cm<sup>2</sup> in January where as the maximum mean was  $79.94 \times 10^4$  cfu/cm<sup>2</sup> in June, the minimum mean of total coliform count was  $0.67 \times 10^3$  cfu/cm<sup>2</sup> in January and the maximum mean was  $1.16 \times 10^3$  cfu/cm<sup>2</sup> in June. The minimum mean of *Staphylococcus aureus* count was  $7.36 \times 10^2$  cfu/cm<sup>2</sup> in January whereas the maximum mean was  $27.11 \times 10^2$  cfu/cm<sup>2</sup> in June.

The same results were observed in the cutting blocks, knives, and workers' hands concerning the minimum and maximum mean count of the studied bacteria.

Salmonella could not be isolated from any examined samples and the percentage of Coagulase positive *Staph. aureus* which were isolated from beef carcasses, cutting blocks, knives and workers' hands samples was 100%.

## INTRODUCTION

Meat is considered as essential foods beings tasty, easily digested and an excellent source of amino acid as well as vitamins and minerals (1), it is exposed to biological and chemical contamination and it is regarded as a good medium for the growth of microorganisms as it provides them with nutrient, available water and optimal PH requirement. Hence, it is considered as a source of different sorts of diseases. Further more meat originates from animals which may be infected with many zoonotic diseases that can be transmitted from the animal to human being via meat consumption (2).

In fact the predominant source of carcass contamination is the animal, particularly the hide and gastrointestinal tract; additional contamination may be acquired from the processing environment, which can be further exacerbated through poor hygiene practices adopted by plant operatives (3, 4, 5). This may lead to spoilage of meat and act as a public health hazard to consumer. Aerobic plate count at the end of slaughtering operations give an important indicator of hygienic quality of the meat, sanitary condition of slaughtering and handling processes (6, 7, 8). It is found to be a suitable indication to assess the number of the organisms and microbial activity (9).

On the other hand coliforms count was required due to *Escherichia coli* which is a member of the coliform group. It is common in the faeces of man and animal. Other coliforms such as Aerobacter and Klebsiella are found to be wide spread in soil, water and plants (10).

Escherichia coli, Aerobacter and Klebsiella are the indicators for the sanitary quality of foods and the presence of one or more of them in great number could easily give rise to public health hazard (11). More over Miskimin et al. (12) found that E. coli count is a suitable indication for the microbiological quality of foods, but to assure safety of a food specific pathogen testing was necessary.

Staphylococci are wide spread in nature and recoverable from many inanimate sources. One of their main hosts is man, and their ready transference from the human body to a variety of foodstuffs accounts for their role as an important cause of food poisoning. By no means all varieties of staphylococcus are concerned. Only those that are coagulase-positive are capable of producing enterotoxins. Food poisoning staphylococci may be isolated from the hands and noses of normal people. Contaminations of towels, tables, knives, dishes, are almost unavoidable. Also staphylococci can be isolated from air, and dust of a contaminated area. Flies in such areas may be infected, and are probably turned into an important source of food contamination by staphylococci (13).

Salmonella can gain access to meat at any stage during butchering by unwashed hands, cutting boards, counters, knives, and other utensils cross contamination of carcasses and meat products could continue, during subsequent handling, processing preparation and distribution (14, 15).

Therefore the present study was carried out to determine the microbial quality of beef carcasses and sanitary condition of butcher's shops in Basrah city through study the effect of month on the microbial load.

## MATERIALS AND METHODS

A total of 160 samples of beef carcasses, cutting blocks, knives, workers' hands and air (32 samples of each) were collected from butchers' shops of Basrah city during three climate periods. The period was started in January and ended in June and all samples examined bacteriologically for aerobic plate count, total coliform count, *Staphylococcus aureus* count and an attempt was made for isolation and identification of Salmonella spp.

Sampling of Beef Carcasses: Sampling of beef carcasses was carried out by using rinse swab method (16). For each sample, a sterile tube containing 10 ml of 0.1 % peptone water and another tube containing two cotton swabs perfectly sterilized were employed. At the time of samples collection, a sterile swab was removed from its tube and moistened with peptone water by dipping it into tube containing 10 ml of 0.1 % peptone water. The sterile metallic template was pressed against the surface to be sampled. The tip of moistened swab was rubbed over the area to be sampled and reswabbed by the other dry swab, then both swabs (moist and dry) were broken off into the tube containing peptone water by using the neck of the tube for leverage, taking aseptic precautions, about 2cm above the swab tip. The tube containing the broken off swabs tip was shaken well for about two minute. The carcass area used for sampling was 25 cm2 from thigh.

**Sampling of Equipment:** The equipment samples in the butchers' shops were knives and cutting blocks. The surface of knives and cutting blocks were swabbed by using rinse swab technique similar to those employed in beef carcasses except the sampling area which was 5 cm<sup>2</sup>.

**Sampling of Workers' Hand:** Each worker's hand was sampled by rolling the swab over the fingers using swab technique similar to that employed in sampling of equipment.

**Sampling of Air:** Samples of air were done by exposure plate method (17) which involved exposing 2 standard size petri-plate (85mm in diameter) containing about 20 ml of Nutrient agar for two minute . The petri-plates were placed about one meter upper the ground and one meter away from the carcasses.

**Transportation of Samples:** The samples, swabs in peptone water tubes and petri-plates duly covered, were transferred by a vehicle immediately to the laboratory by kept them in a cold insulated box, these samples were then subjected to bacteriological examination immediately on

reaching the laboratory .At the time of processing of one sample, the other samples were kept in the refrigerator at 4°C.

**Bacteriological analysis:** Serial dilution was used to prepare duplicate plates for the determination of aerobic plate counts (APCs), coliforms, and *Staph. aureus*.

APCs were determined by using Nutrient agar (Himeda, India) and plates were incubated at 37C° for 48 h. Then all colonies on plates were counted (18).

Colifrom counts were determined by using MacConkey agar (Himeda, India), typical colonies were identified as round, red to pink 0.5-2 in diameter, surrounded with a red to pink halo (19).

For the *Staph. aureus* counts 'Mannitol salt agar( Himeda, India)was used and the plates incubated at 37°C for 24 to 48 hr (APHA,1978). All typical colonies on Mannitol salt agar was counted ,selected colonies from the agar surfaces were tested for coagulase activities using rabbit plasma(20, 21, 22).

For the enrichment of salmonella, five ml from the original sample was transferred to a 45 ml of tetrathionate broth (oxoid, U.K) and incubated at 37°C for 24 hr. One ml from enrichment culture was transferred to a brilliant green agar (oxoid, U.K) was used for selective plating .Presumptive salmonella colonies selected from each of selective plates were subjected to the biochemical test including: Triple Sugar Iron agar for fermentation and H2S production, Urease broth for urea production and Sulphide Indole Motility medium for indole production were inoculated (23). All cultures were incubated at 37°C for 24 hours.

**Statistical Analysis:** The results were analyzed by One-way ANOVA test, using statistical package for the social sciences (SPSS, version 9.0). All data were expressed as Mean±Std.Error. Differences between data were compared by least significant deference (24).

## **RESULTS AND DISCUSSION**

The aerobic plate count of beef carcasses varied from  $3 \times 10^4$  to  $1.72 \times 10^6$ , with mean values of  $5.84 \times 10^5 \pm 1.21 \times 10^5$  cfu /cm² (Table 1). These results were higher than the standard regulation for fresh beef  $(1 \times 10^4$  to  $1 \times 10^5$  cfu/cm²) under typical stipulating conditions (25). On the other hand, the results of present study was less than what has been recorded by Hoshyare (26) who reported that the mean of aerobic plate count in Baghdad was  $2.78 \times 10^6$  cfu/cm². All samples of beef carcasses had aerobic plate count which is more than  $1 \times 10^3$  and only 9.38 % of them had the count  $>1 \times 10^4$  to  $<1 \times 10^5$ . These results were lower than the results of Abdul-Wadood (27) who found that 80.95% of beef carcasses had aerobic plate count  $1 \times 10^4$  cfu/gm²

The difference obtained in the present study may be related to defect in the sanitation, transportation, temperature, handling and ways of presentation of the meat in the butchers' shops.

The data recorded in Table(2) showed that the minimum means of aerobic plate count /cm² of beef carcasses was 27.32x10⁴ in January while the maximum means was 79.94x10⁴ in June These results were in agreement with those reported by other studies (27;28) who reported that the minimum means of all examined bacteria of beef carcasses was in January while the maximum means was observed in June, but these results differed from Hoshyare (26) who found that minimum means of aerobic plate count of beef carcasses was in January and the maximum means was in April and the count of all examined bacteria showed gradual increase during March ,April ,May and June as compared to the situation in January and February.

The aerobic plate count of cutting blocks, knives, workers' hands and air samples had varied from  $0.12 \times 10^4$  to  $17.80 \times 10^4$ ,  $0.03 \times 10^4$  to  $1.40 \times 10^4$ ,  $0.02 \times 10^4$  to  $0.38 \times 10^4$  and  $0.08 \times 10^4$  to  $0.11 \times 10^4$ . With mean values  $2.36 \times 10^4 \pm 0.71 \times 10^4$ ,  $0.46 \times 10^4 \pm 0.07 \times 10^4$ ,  $0.12 \times 10^4 \pm 0.01 \times 10^4$  cfu/cm² and  $9.79 \pm 0.14$ /cm² per minute of exposure ,respectively. The same trend of results was observed in the cutting blocks, knives, and workers' hands concerning the minimum and maximum mean counts of the studied bacteria (Table 2). This gradual increment of bacterial count in the present study may be due to gradual elevation of the temperature in these months.

These results seem to be high in compared to the standard (<1x10³ organism/cm²) as suggested by Patterson (29). Up to our knowledge there were few reports concerning the aerobic plate count in cutting blocks, knives and workers' hands in butchers' shops. However, Jawad (30) and Abdul-Wadood (31) found that the aerobic plate count of knives was 1.69x10⁵ cfu/cm² and 13.94x10⁴ cfu/cm² respectively. The difference of results may be related to poor sanitation practice in the butchers' shops. In the retail market, Narasimha and Ramesh (32) reported that additional contamination is usually taken places with reference to knives, saws, cleavers, slices, chopping blocks, scales, containers, flies, hands and garments of the workers, market operators, and air. Total numbers of microorganisms in a beef may increase due to the pollution of air with microorganisms. The numbers of microorganisms in the air at any given time depend upon a number of factors including movement of people, sunshine, humidity, location, aerosol, and by ventilation (17; 33).

The total coliform count/cm<sup>2</sup> of beef carcasses varied from  $0.26 \times 10^3$  to  $1.76 \times 10^3$  with mean values of  $0.99 \times 10^3 \pm 0.07 \times 10^3$  cfu/cm<sup>2</sup> (Table 3). These present results were lower than that reported by Hoshyare (26) who stated that the mean of total coliform count in butchers' shops in Baghdad was  $8.1 \times 10^3$  cfu/cm<sup>2</sup>. All samples of beef carcasses had total coliform count which is more than  $1 \times 10^2$  and only 53.13% of beef carcasses samples had the counting  $>1 \times 10^3$  to  $<1 \times 10^4$ . On the other hand, the minimum means of total coliform count /cm<sup>2</sup> of beef carcasses was  $0.67 \times 10^3$  in January and the maximum means was  $1.16 \times 10^3$  in June (Table 2)

These results were higher than the result of Goepfert and kim (34) who reported that 17.5% of beef carcasses exceed  $1x10^3$  cfu/gm. The difference obtained at the present study may be related to the difference in the procedure of coliform counting (Most Probable Number), hygienic measurement, climate, personal hygiene and education. In general the presence of coliforms in the food has been considered by some authors as a criterion for the existence of unsanitary conditions (12; 25; 35; 36). However, the presence of coliforms in the carcasses in the present study may indicate contamination from either faecal or non faecal sources. The high coliform counts of carcasses, when considered along with the total bacterial counts would suggest the presence of unsanitary conditions in the processing plants. The total coliform count/cm<sup>2</sup> of cutting blocks, knives and workers' hands samples had varied from  $0.24 \times 10^3$  to  $1.32 \times 10^3$ ,  $0.22 \times 10^3$  to  $1.60 \times 10^3$  and  $0.20 \times 10^3$  to  $1.10 \times 10^3$ , respectively. With mean values  $0.87 \times 10^3 \pm 0.06 \times 10^3$ ,  $0.82 \times 10^3 \pm 0.06 \times 10^3$  and  $0.73 \times 10^3 \pm 0.05 \times 10^3$  cfu/cm<sup>2</sup> (Table 4).

Staphylococcus aureus count/cm² of beef carcasses varied from  $0.6 \times 10^2$  to  $84 \times 10^2$  with mean values of  $16.18 \times 10^2 \pm 3.22 \times 10^2$  cfu /cm² (Table 5). The present results were lower than the result of Hoshyare (26) who reported that the mean of *Staph. aureus* count in butchers' shops in Baghdad was  $7.19 \times 10^3$  cfu/cm². Four samples of beef carcasses in the present study had *Staph.aureus* count less than  $10^2$  and only 37.5% of them had counting  $>1 \times 10^2$  to  $<1 \times 10^3$  (Table 6). These results were higher than the results of Wyatt and Guy (37) who found that 5% of beef carcasses exceed  $1 \times 10^2$  *Staph. aureus* count.

The total of *Staph. aureus* count/cm<sup>2</sup> of cutting blocks, knives and workers' hands samples were varied from  $18 \times 10^2$  to  $13.80 \times 10^4$ ,  $0.80 \times 10^2$  to  $20 \times 10^2$ , and  $0.60 \times 10^2$  to  $1.20 \times 10^2$  with the mean values of  $15.98 \times 10^2 \pm 41.82 \times 10^2$ ,  $7.76 \times 10^2 \pm 1.21 \times 10^2$  and  $0.89 \times 10^2 \pm 0.03 \times 10^2$  cfu/cm<sup>2</sup>, respectively, (Table 5).

These results were lower than the results of Abdul-Wadood (31) who revealed that *Staph* .aureus count of knives was  $13.94 \times 10^4$  cfu/cm<sup>2</sup>. The difference in results may be related to the certain bad habits of handlers and absence of the hygienic education concerning good standards of healthy and hygienic care which must be provided by medical officers, public health inspection (31; 38). Mostly raw meats are contaminated with staphylococci at the time of slaughtering or during handling after slaughtering (39).

#### Bas.J,.Vet.Res.Vol.6,No.2,2007.

The beef carcasses, cutting blocks, knives and workers' hands samples showed positive results in coagulase test. These results were in agreement with the results of Abdul-Wadood(27) who reported that all isolated (100%) *Staph. aureus* were coagulase positive, but this results differ from the results of Hoshyare(26) and Al-kasei (40) who found that 97.5% and 93.3% of *Staph. aureus* isolates were coagulase positive.

.Also the minimum means of *Staph. aureus* count  $/\text{cm}^2$  of beef carcasses was  $7.36 \times 10^2$  in January while the maximum means was  $27.11 \times 10^2$  in June (Table 6).

The beef carcasses, cutting blocks, knives and workers' hand samples showed negative results in Salmonella identification testing. The results were in agreement with the results of Lotfi *et al.*, **(41)** and Abd El-Aziz, **(42)** who got negative results in relation to salmonella identification.

The beef carcasses, cutting blocks, knives and workers' hand samples showed negative results in Salmonella identification testing. The results were in agreement with the results of Lotfi *et al.*, **(41)** and Abd El-Aziz, **(42)** who got negative results in relation to salmonella identification.

Table (1): The aerobic plate count in examined samples

Source of somples			
Source of samples	Mean	Mean Range	
Carcasses	58.41	$3.00-172 \times 10^4$	±12.16
Cutting blocks	2.36	$0.12\text{-}17.80\text{x}10^4$	± 0.71
Knives	0.46	$0.03-1.40x10^4$	± 0.07
Workers' hands	0.12	$0.02 - 0.38 \times 10^4$	± 0.01
Air	9.79	$0.08 - 0.11 \times 10^4$	± 0.14

Results are expressed as mean colony forming units x 10<sup>4</sup> per cm<sup>2</sup>.

Table (2): The aerobic plate count of examined samples during six months under the study.

the study.								
Sources	Aerobic plate count							
of	January	Februar	March	April	May	June		
samples		y						
carcass es	27.32 <sup>M</sup> 3-114 <sup>R</sup> ±21.86 <sup>S</sup> .E	35.80 3.2-152 ±29.07	63.20 6-146 ±33.41	64.28 3.40-152 ±35.41	71.36 8-166 ±38.23	79.94 3.60-172 ±27.67		
cutting blocks	0.52 0.12- 0.90 ±0.15	0.75 0.25- 1.10 ±0.16	2.10 0.03- 5.60 ±0.97	2.22 0.14- 5.80 ±1.00	4.37 0.52-15.80 ±2.89			
knives	0.52 0.03- 0.94 ±0.19	0.55 0.03-1 ±0.17	0.65 0.03- 1.40 ±0.25	0.654 0.03-1.20 ±0.20	0.68 0.03-1.20 ±0.21	0.74 0.03-1.14 ±0.18		
workers ' hands	0.05 0.02- 0.09 ±0.01	0.12 0.03- 0.32 ±0.05	0.125 0.07- 0.30 ±0.04	0.13 0.0332 ±0.049	0.138 0.0338 ±0.06	0.16 0.09-0.34 ±0.04		
air	9.32 0.08- 0.10 ±0.47	9.35 0.08- 0.10 ±0.44	9.81 0.08- 0.10 ±0.33	9.88 0.08-0.10 ±0.42	10.04 0.09-0.11 ±0.32	10.16 0.09-0.11 ±0.24		

M= means counts, R= range between, S.E= standard error Results are expressed as mean colony forming units  $\times 10^4$  per cm<sup>2</sup>.

Table (3): The total coliform count in examined samples

Course of comples			
Source of samples	Mean	Range	Std.Error
Carcasses	0.99	$0.26 - 1.76 \times 10^3$	±0.07
Cutting blocks	0.87	$0.24-1.32 \times 10^3$	± 0.06
Knives	0.82	$0.22 - 1.60 \times 10^3$	± 0.06
Worker's hands	0.73	$0.20 - 1.10 \times 10^3$	± 0.05

Results are expressed as mean colony forming units x10<sup>3</sup> per cm<sup>2</sup>.

Table (4): The total coliform count of examined samples during six months

Sources	Total coliform count					
of samples	January	Februar y	March	April	May	June
carcass es	$0.67^{M}$ $0.26$ - $1.32^{R}$ $\pm 0.1$ S.E	0.87 0.28- 1.32 ±0.20	1.00 0.28- 1.54 ±0.24	1.10 0.66-1.32 ±0.12	1.11 0.56-1.54 ±0.1	1.16 0.3-1.765 ±0.20
cutting blocks	0.63 0.24- 1.20 ±0.15	0.79 0.26- 1.20 ±0.18	0.38 0.26- 1.32 ±0.21	0.94 0.54-1.32 ±0.15	1 0.60-1.20 ±0.11	1.67 0.28-5.40 ±2.71
knives	0.60 0.22- 1.10 ±0.19	0.72 0.24- 1.10 ±0.17	0.79 0.24- 1.32 ±0.20	0.86 0.52-1.20 ±0.13	0.91 0.26-1.20 ±0.17	0.96 0.52-1.60 ±0.13
workers 'hands	0.57 0.20-1 ±0.12	0.12 0.22-1 ±0.15	0.125 0.22- 1.10 ±0.16	0.80 0.50-1.10 ±0.11	0.808 0.241.10 ±0.15	0.82 0.50-1.10 ±0.08

M= means counts, R= range between, S.E= standard error Results are expressed as mean colony forming units  $x ext{ } 10^3 ext{ per cm}^2$ .

Table (5): Stapylococcus aureus count in examined samples

Course of complex		Staph.aureus count		
Source of samples	Mean	Range	Std.Error	
Carcasses	16.18	$0.6-84 \times 10^2$	±3.22	
Cutting blocks	15.98	$18-1380 \times 10^2$	± 41.82	
Knives	7.76	$0.80-20x10^2$	± 1.21	
Workers' hands	0.89	$0.60-1.20 \times 10^2$	± 0.03	

Results are expressed as mean colony forming units x10<sup>2</sup> per cm<sup>2</sup>

Table (6): The *Staph. aureus* count of examined samples during six Months under the study

Withing the study								
Sources of	Staph.aureus count							
samples	January	Februar	March	April	May	June		
samples		y						
	7.36M	13.20	1.3.37	15.32	16.32	27.11		
carcasses	0.60-16R	0.60-30	0.64-32	1-50	2-36	2.20-84		
	±2.75S.E	±5.09	±5.96	±8.90	$\pm 7.28$	±10.48		
cutting blocks	79.20 18.40- 180 ±32.39	122.80 36-240 ±36.48	125.20 20-220 ±41.93	140 28-260 ±38.79	151.60 38-260 ±40.58	171.14 30-480 ±58.09		
knives	2.96 1.18-6 ±0.87	3.96 0.8015 ±2.76	8.56 2-20 ±3.81	9.32 2.60-18 ±2.98	10.28 2.60-19 ±3.48	10.43 3-18 ±2.60		
workers' hands	0.75 0.66- 0.88 ±0.05	0.78 0.60-1 ±0.07	0.28 0.60-1 ±0.08	0.94 0.80-1.10 ±0.05	0.97 0.80- 1.10 ±0.05	1.03 0.80-1.20 ±0.06		

M= means counts, R= range between, S.E= standard error Results are expressed as mean colony forming units  $x \cdot 10^2$  per cm<sup>2</sup>

# النوعية البكتيرية لذبائح الأبقار والحالة الصحية لمحلات القصابين في البصرة عليه البكتيرية الدبن حسن جواد و آلاء طار ق عبد الواحد

فرع الفسلجة ، كلية الطب البيطري، جامعة البصرة ، البصرة ، العراق.

فرع الاحياء المجهرية ، كلية الطب البيطري، جامعة البصرة ، البصرة، العراق.

#### الخلاصة

تم استقصاء النوعية الميكروبية لذبائح الأبقار و الحالة الصحية لمحلات القصابين واحتماليه وجود الجراثيم المرضية المرتبطة بتغشي حاله التسمم الغذائي في الإنسان مثل السالمونلا والمكورات العنقودية في مائه وستين عينه من ذبائح الأبقار ، ألواح التقطيع ، السكاكين ، أيدي العاملين والهواء (32 عينه من كل نموذج ) جمعت هذه العينات من محلات القصابين الواقعة في أربعه أسواق محليه في مدينه البصرة (الجمهورية ، البصرة ، العشار و خمسه ميل) ابتداء من شهر كانون الثاني وانتهاء بحزيران 2004. جميع العينات فحصت بواسطة العد الكلي للجراثيم الهوائية ، العد الكلي لبكتريا القولون ، عد المكورات العنقودية الذهبية ومحاولة عزل وتشخيص السالمونلا عدا عينات الهواء التي فحصت بواسطة العد الكلي للجراثيم الهوائية فقط أظهرت نتائج هذه الدر اسه بأن هناك زيادة تدريجية في عدد جميع الميكروبات ابتداء من كانون الثاني وانتهاء بحزيران وان الحد الأدنى لمعدل العد الكلي للجراثيم الهوائية لذبائح الأبقار كان  $27,32 \times 10^4$  و. ت.م/ سم في كانون الثاني بينما الحد الأعلى المعدل كان 1,000 و. ت.م/ سم في كانون الثاني بينما الحد الأعلى للمعدل كان 1,000 و. ت.م/ سم في كانون الثاني بينما الحد الأعلى للمعدل كان 1,000 و. ت.م/ سم في كانون الثاني بينما الحد الأعلى للمعدل كان 1,000 و. ت.م/ سم في حزيران فيما يتعلق بالحد الأدنى و الحد الكتيريا المكورات العنقودية الذهبية كان 1,000 و. ت.م/ سم في كانون الثاني بينما الحد الأدنى و الحد الأعلى لمعدل عد البكتير با المدر و سة.

لم نتمكن في هذه الدراسة من عزل جرثومة السالمونيلا من جميع العينات، وأن نسبة الكورات العنقودية التي اظهرت نتيجة ايجابية في فحص انزيم التخثر والمعزولة من ذبائح الابقار والواح التقطيع والسكاكين وايدي العاملين 100%.

#### REFERENCES

- 1- Briggs, G.M. (1985). Muscle food and human health. Food Technol.2: 54-58.
- **2-Corry, J.E. and Hinton, M.H. (1997).** Zoonosis in the meat industry. Acta. Vet. Hung. 45: 457-479.
- **3- Al-Tai, M.A.J.(1986).** Fish and Meat Technology Dar Al-kutob for press. Basrah Univ. (in Arabic)
- **4-Biss, M.E. and Hathaway, S.C.** (1996). Effect of pre-slaughter washing of lambs on the microbiological and visible contamination of the carcasses. Vet. Rec.138: 82-6.
- **5-Galland, J.C. (1998).** Risks and prevention of contamination of beef carcasses during the slaughter process in the United States of America .Rev .Sci .Off. Int.Epiz.16: 395-404
- **6-Nottingham, P.M.N.; Pennoy, I. and Herrisori, J. C. L. (1974).** Microbiology of beef processing/beef dressing hygiene. Newzealand. Agr. Res. 17:79-83.
- **7-Ingram, M. and Roberts 'T. A. (1976).** The microbiology of the red meat carcass and the slaughter house. Royal Soc. Health J. 96:720-276.
- **8-Roberts, T. (1980).** Contamination of meat. The effects of slaughter practices on the bacteriology of the red meat carcass. Royal Soc. Health J .100:3-9
- **9-Johanson, L.; Underdal, B.; Grostand, K.; Whelehan, O.P. and Roberts, T.A. (1983).** A survey of the hygienic quality of beef and pork carcasses in Norway. Acta. Veterinaria Scandinavica. 24:1-3.
- **10-Breed, R.S.; Murray, E.G.D. and Smith, N.R.** (1957). Bergey's manual of determinative bacteriology.7th ed. The Williams and Wilkins Company, Baltimore.
- **11-Freeman, W.H.** (1960). The science of meat and meat products. Amer. Inser. Found. Reinhold pub. Corporation, New York.
- 12-Miskimin, D.K.; Berkowitz, K.A.; Solberg, M.; Rhia, W.E.; Franke, W.C.; Buchman, R.L. and O'Lear, V. (1976). Relation ships between indicator organisms and specific pathogens in potentially hazards food. J. Food Sci.41: 1001-1006.
- **13-Christie, A. B.** (1980). Infectious Diseases Epidemiology and clinical Practice.3rd ed., churchill Livingstone, Edinburgh, P.24-36, 60-70.
- **14-Kampelmacher, E.H.; Guinee, P.A.M.; Hofstra, K. and Van Keulen, A. (1961).** Studies on Salmonella in slaughterhouses. Zentra- 1b1. Veterinaermed.Reihe.8: 1025-1032.
- **15-Adesiyun, A.A. and Oni, O.O.** (1989). Prevalence and anti- biograms of Salmonella in slaughtered cattle slaughter areas and effluents in Zaria abattoir, Nigeria. J.Food Prot.52:232-235.
- **16-Hall, M.A. and Maurer, A. J. (1980).** The microbiological aspects of a duck processing plant. Poultry Sci. 59: 1795-99
- **17-Gregory, P.H.** (1961). The microbiology of the atmosphere. Interscience Publishers (Division of John Wiley and Sons, Inc.) New York
- **18-American public Health Association.** (**1978**). Standard Methods for the Examination of Dairy products. 14th ed., Washington , U.S.A.
- **19-Mahmood, K.H.; Mohamed, S.H. and Al-Kashaly, S.S. (1992).** The microbial quality of raw ground beef from local markets in Basrah, Iraq. Basrah J. Agric. Sci.5: 7-14.
- **20-.Gillespie**, **E.H.** (**1943**). The routine use of the coagulase test for Staphylococci. Monthly Bulletin Emergency Public Health Laboratory Service, 2-19.
- **21-Williams, R.E.O. and Harpper, G.J. (1946).** Determination of coagulase and alphahaemolysin production by Staphylococci. Brit.J.Exp.Pathol.27: 72.
- **22-Sproer, W. H. and Tatini, S. R. (1975).** Interpretation of the tube coagulase test for identification of Staphylococcus aureus. Appl. Microbiol. 29:502-505.
- **23-International Organisation for Standarisation 6579.(1998).** Microbiology of food and animal feeding stuff. Horizontal method for detection of salmonella. ISO, Geneva.

- **24-Snedecor. G.W. and Cochran, W.G. (1971).** Statistical Methods. 6th ed., Iawa State University Press. Ames, Iawa, USA
- **25-Frazier, W. C.(1967).** Food Microbiology. 2<sup>nd</sup> ed. New York, Macgraw Hill, Inc.
- **26-Hoshyare, D. F.** (1978). Microbial quality of meat in Baghdad. M. Sc. Thesis, collage of Agriculture. Baghdad. Univ. (in Arabic)
- **27-Abdul-Wadood, E. (1999).** Effect of microwave oven on some groups of microorganisms of beef in Baghdad.M.Sc. Thesis, College of Vet. Med./ Baghdad Univ.
- **28-Berrada**, S.A.(1972). Bacteriological study of minced meat in Casablanca. Thesis No.43 FSTA Abstr.1973.5:885-983.
- **30-Jawad, A.A.** (1982). Hygienic evaluation of commercially processed poultry in Iraq .M. Sc. Thesis, college of Veterinary Medicine. Baghdad Univ.
- **31- Abdul-Wadood, E. (2002).** Study on some hygienic aspect of Basrah Abattoirs. Bas .J. Vet. Res., Vol. 1, No.2. (in Arabic)
- **32-Narasimha, R. and Ramesh, B. S. (1988).** Microbial profiles of minced meat. J. Meat Sci. 23:279-291..
- **33-Heldman, D.R.; Hedrick, T.I. and Hall, C.W. (1964).** Air—borne microorganism population in food packaging area .J. Milk Food Technol.27: 245-251.
- **34-Goepfert, J. M. and Kim, H. U. (1976).** Behavior of selected food- borne pathogens in raw ground beef. J. Milk Food Technol. 449-450.
- **35-Thatcher, F.S. and Clark, D.S.** (1959). Microorganisms in foods. Their significance and method of enumeration. Toranto: University of Toranto press.
- **36-**American Public Health Association..(1966). Recommended methods for the examination of food. 2<sup>nd</sup>. New York.
- **37-Wyatt, C.J. and Guy, V. (1980).** Relationships of microbial quality of retail meat samples and sanitary conditions. J. Food Prot. 43:385-380.
- 38-Hobbs, B.C. and Gilbert, R.J. (1968). Food hygien.4th ed., London, Edward Arnold.
- **39-Minor, T. E. and Marth, E. H. (1976).** Staphylococci and Their Significance in Food. Elseveier Scientific Publishing Company, Amsterdam. Holland.
- **40-Al-Kasei, S.H.M.** (1985). Study on microbial content in meat product in Baghdad. M. Sc. Thesis, college of Agriculture. Baghdad Univ.(in Arabic)
- 41-Lotfi, A.; Nasr, S.; Yaussef, K.; Abdel Rahman, H.; Hefnawy, Y.; El-Tinawy, A. and Gobran, R. (1986). Enterobacteriaceae in meat products in Upper Egypt. Assiut Vet. Med. J. 17: 77.
- **42-Abd El-Aziz, A.S.** (1988). Salmonella in locally produced meat products. M.Sc. Thesis, College of Vet.Med.Cario Univ. (in Arabic)