

## First- and Second-Order Derivative Spectrophotometry for Individual and Simultaneous Determination of amoxicillin and cephalixin

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### Abstract

A new spectrophotometric method for individual and simultaneous determination of amoxicillin and cephalixin depending on the first and second derivative mode techniques. The first and second derivative spectra of these compounds permitted individual and simultaneous determination of amoxicillin and cephalixin in concentration interval of (10 – 60  $\mu\text{g}.\text{ml}^{-1}$ ) by measuring the amplitude of peak-to-base line, zero cross at certain wavelengths and the area under peak at selected spectrum intervals. The methods showed reasonable precision and accuracy and have been applied to determine amoxicillin and cephalixin in four different pharmaceutical preparations.

### الخلاصة

تم تطوير طريقة طيفية جديدة لتقدير الاموكسيسلين و السفلكسين بشكل مفرد و بشكل آني بالاعتماد على تقنية المشتقة الأولى والمشتقة الثانية لأطياف هذه المركبات. لقد وجد ان المشتقة الأولى والمشتقة الثانية لأطياف هذه المركبات تمكن من التقدير الانني لالاموكسيسلين و السفلكسين بمدى تراكيز يتراوح بين ( ١٠ – ٦٠ مكغم.مل<sup>-١</sup>) و ذلك بقياس ارتفاع القمة-خط القاعدة و قياس ارتفاع المنحني في نقطة التقاطع الصفري عند اطوال موجية محددة لكل مركب، وكذلك من خلال قياس المساحة تحت الحزمة لمجالات طيفية محددة. لقد كانت النتائج التي تم الحصول عليها من تحليل المركبات قيد الدراسة متوافقة ودقيقة بشكل مقبول، كما وأمكن تطبيقها لتقدير الاموكسيسلين و السفلكسين بشكل ناجح في اربعة مستحضرات صيدلانية.

### Introduction

Although many methods have been reported for sensitive and selective determination of antibiotics including solid phase extraction<sup>(1,2)</sup>, thin layer chromatography<sup>(3)</sup>, liquid chromatography-mass spectrometry<sup>(4,5)</sup>, light scattering-resonance Raleigh<sup>(6,7)</sup>, radioimmunoassay<sup>(8)</sup> and spectrophotometry<sup>(9)</sup>, little attention have been paid to develop new method for simultaneous spectrophotometric determination of antibiotics.

The importance of the derivative spectroscopy for interpretation of UV-VIS spectra and for quantitative analysis is well known and documented<sup>(10)</sup>. It has a great utility to resolve the overlapping spectra especially in those methods which lack selectivity making them suitable for application<sup>(11)</sup>. Derivative spectrophotometric methods have been utilized for simultaneous determination of metal ion mixtures<sup>(12,13)</sup> and could extend its uses to other analytical fields.

The purpose of this work is to determine amoxicillin and cephalixin using first and second derivative spectrophotometry and to demonstrate that these methods can be very useful tools for determining amoxicillin and cephalixin in mixture, without tedious and time consuming separation procedures.

## Experimental

### Apparatus

A Shimadzu UV1601 double beam UV-VIS spectrophotometer was loaded with Shimadzu UVProb Version 1.10 software and interfaced to Pentium-4 computer and Canon-810 laser printer to record the spectra and perform subsequent calculations of their derivatives.

The spectrophotometric measurements were made at wavelength range 200-350 nm using 1-cm quartz matched cells. The derivative spectra were recorded with a fast scan speed, sampling interval=1.0 and slit width=2.0 nm.

### Reagents

**Amoxicillin Standard Solution** ( $100 \mu\text{g}\cdot\text{ml}^{-1}$ ): 0.01 g of amoxicillin (obtained from the state company for drug industries and medical appliance (S.D.I.), Samara-Iraq) is dissolved in 1 ml 0.1 M sodium hydroxide solution then the mixture is diluted to 100 ml in a volumetric flask with doubled distilled water.

Working solutions were freshly prepared by subsequent dilutions.

**Cephalexin Standard Solution** ( $100 \mu\text{g}\cdot\text{ml}^{-1}$ ): 0.01 g of cephalixin (obtained from the state company for drug industries and medical appliance (S.D.I.), Samara-Iraq) is dissolved in 1 ml 0.1 M sodium hydroxide solution then the mixture is diluted to 100 ml in a volumetric flask with doubled distilled water.

Working solutions were freshly prepared by subsequent dilutions.

**Analysis of capsule:** The content of 10 capsules were mixed well and a certain portion of the fine powder was

accurately weight to give an equivalent to 0.01g of amoxicillin or cephalixin, and dissolved in 1 ml 0.1 M sodium hydroxide solution. The resulted solution was diluted to 100 ml with double distilled water in a volumetric flask. The solution was filtered by using Whatmann filter paper No. 40 to avoid any suspended or un dissolved material before use.

### Procedure

Individual determination of amoxicillin and cephalixin:

In 10 ml calibrated flask, transfer aliquots of amoxicillin or cephalixin solutions expected to contain ( $100 - 600 \mu\text{g}$ ) and dilute to the mark with doubled distilled water. The absorption spectra were recorded and show absorption maxima at 226 nm and 274 nm for amoxicillin, and 212 nm for cephalixin.

Determinations were made by measuring the first and second derivative values and area under peaks of their spectra at certain given wavelengths and wavelengths regions. The concentration of amoxicillin and cephalixin could be determined respectively.

Simultaneous determination of amoxicillin and cephalixin:

( i ) The content of a series of 10 ml calibrated flasks, containing  $200 \mu\text{g}$  of cephalixin and different amounts ( $100 - 600 \mu\text{g}$ ) of amoxicillin, were diluted with doubled distilled water. The absorption spectra were recorded against blank( prepared by the same manner as test solution but without cephalixin or amoxicillin) ; then by measuring the derivative values of their first and second spectra, the concentration of amoxicillin could be determined.

( ii ) The content of a series of 10 ml calibrated flasks, containing  $200 \mu\text{g}$  of amoxicillin and different amounts ( $100 - 600 \mu\text{g}$ ) of cephalixin, were diluted with doubled distilled water. The absorption spectra were recorded against blank( prepared by the same

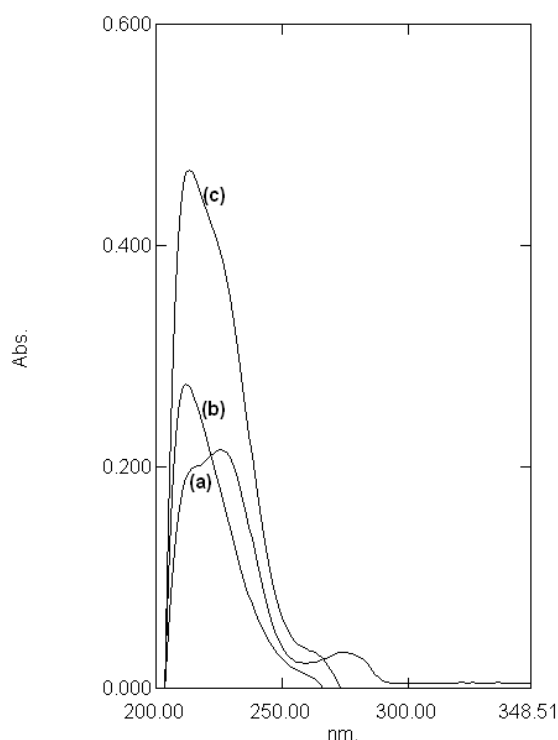
manner as test solution but without amoxicillin or cephalixin); then by measuring the derivative values of their first and second spectra at selected wavelengths and reagents, the concentration of cephalixin could be determined.

## Results and Discussion

### Absorption spectra

The absorption spectra of amoxicillin and cephalixin and for their mixture were recorded. Fig. 1 (a)

shows the absorption spectrum of amoxicillin solution ( $20 \mu\text{g. ml}^{-1}$ ) with two maxima at wavelength 226 nm and 274 nm, curve (b) show the absorption spectrum of cephalixin with maximum wavelength of absorption at 212 nm. The total spectrum of mixture of ( $200 \mu\text{g}$  of each per 10 ml) is shown in curve c with  $\lambda_{\text{max}}$  (214 nm) between the absorption maxima of the two components.



**Figure 1: Absorption spectra of (a)  $20 \mu\text{g. ml}^{-1}$  of amoxicillin, (b)  $20 \mu\text{g. ml}^{-1}$  of cephalixin, (c) amoxicillin plus cephalixin.**

### First and second derivative modes

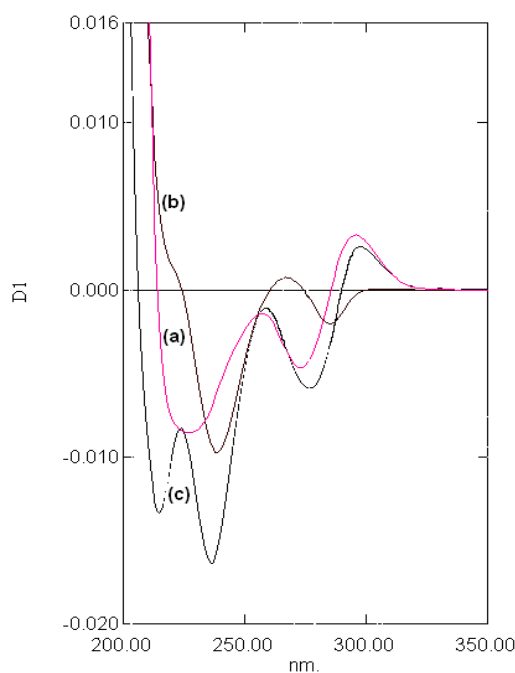
The first order and second order derivative spectra of amoxicillin and cephalixin and for their mixture are shown in Fig. 2 and Fig. 3 respectively.

It is obvious that there is a large overlap of the spectra of amoxicillin and cephalixin therefore, their determination, using the zero order absorption measurements, when present in the same solution is very difficult when using traditional two wavelengths of maximal absorption or the tangential base-line approximation<sup>(14)</sup> techniques. On the

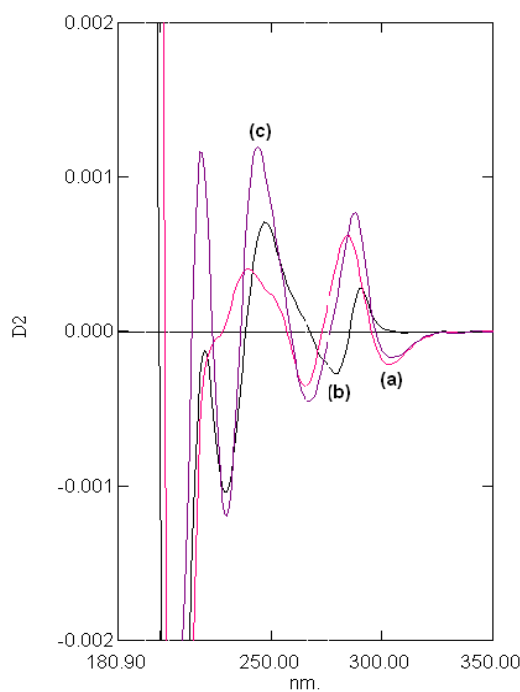
other hand, derivative spectrophotometric technique is of a particular utility in determining the concentration of single component in such mixtures, with a large spectral overlapping. For this reason, derivative spectrophotometric methods have been applied. Both first and second order modes were tested, the results obtained show that these techniques could successfully applied when the measurements are carried out under optimum selection of slit width, response time, and scan speed for the monochromator. These were done by

measuring the magnitude of derivative at several slit widths and scan speed with different response times. A slit

width of 2 nm, a response time of 4 seconds and fast scan speed were found to be optimum.



**Figure 2: First derivative spectra of : (a) 20  $\mu\text{g.ml}^{-1}$  of amoxicillin, (b) 20  $\mu\text{g.ml}^{-1}$  of cephalixin, (c) amoxicillin plus cephalixin**



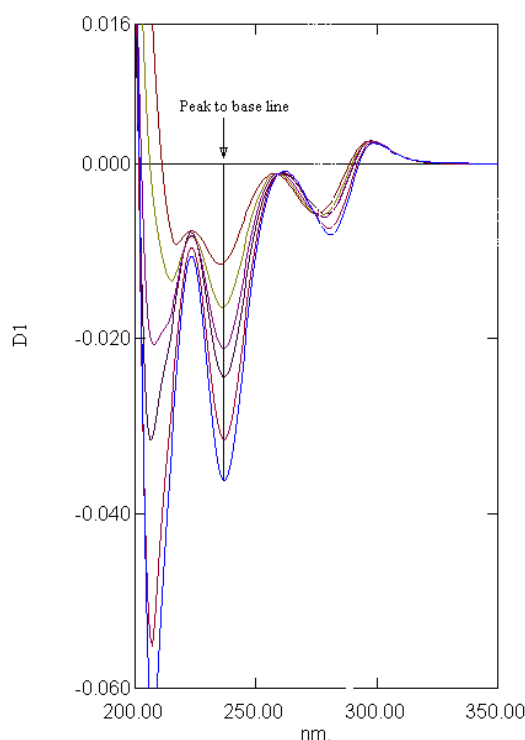
**Figure 3: Second derivative spectra of : (a) 20  $\mu\text{g.ml}^{-1}$  of amoxicillin, (b) 20  $\mu\text{g.ml}^{-1}$  of cephalixin, (c) amoxicillin plus cephalixin**

In the present work, graphically (peak-to-base line), zero-crossing technique in addition to peak area were used to deal with derivatives spectra to carry out the measurements. In fact that all these techniques in the first and second derivative modes show good proportionality to amoxicillin and cephalixin concentrations in their mixtures.

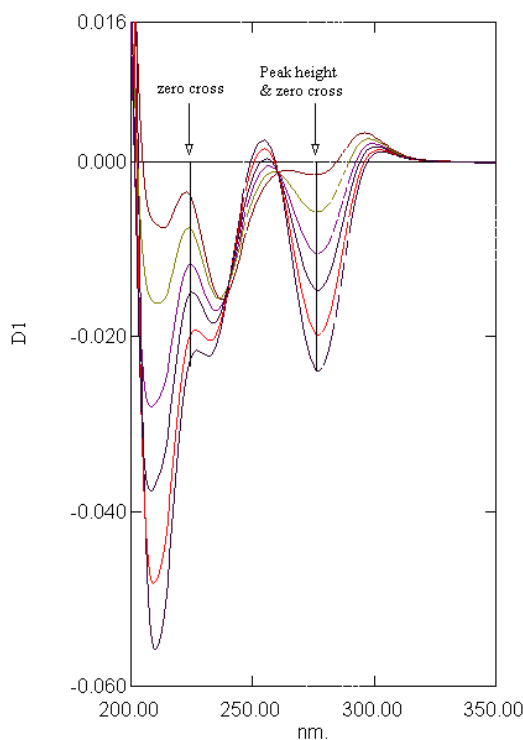
To select the derivative order, the first, second, third and fourth derivative spectra of amoxicillin and cephalixin were studied. The study reveals that first and second order spectra were simple and gave results of highest accuracy and detection limits.

Fig. 4 and Fig. 5 show sets of first order spectra of mixtures containing different amounts of each of amoxicillin and cephalixin in the presence of ( $20 \mu\text{g}.\text{ml}^{-1}$ ) of the other compound respectively.

The results in Fig. 4 indicate that when the concentration of cephalixin is kept constant and the concentration of amoxicillin varied, the peak areas at the intervals (202 – 224 nm) and (224–262 nm) were proportional to the concentration of amoxicillin. Moreover, the peak-to-base line (i.e. amplitude measured in millimeter) at (236nm) was found in proportion to the amoxicillin concentration. The same features were found when inspecting Fig. 5 for the determination of cephalixin, i.e. peak areas in the wavelengths intervals of (225–254nm), (254–294 nm) and the peak amplitude measured at peak-to-baseline (275.5nm) and at zero cross of amoxicillin (223.9 nm) were in proportion to the concentration of cephalixin (Table1).



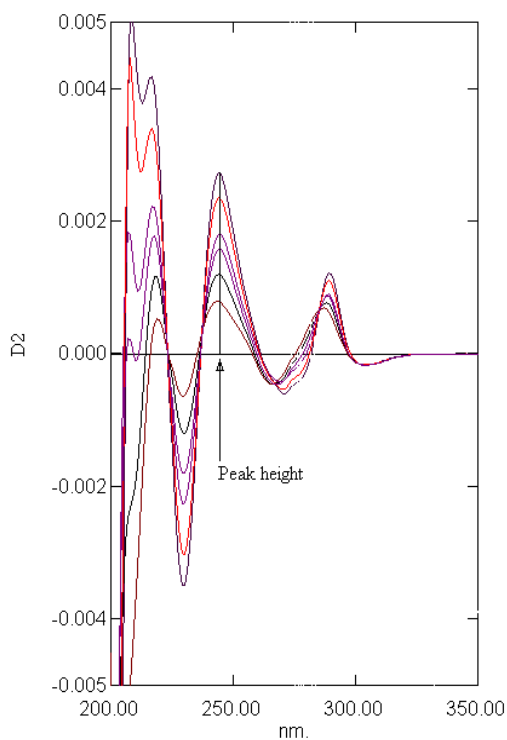
**Figure 4: First derivative spectra of mixtures containing (10 – 60  $\mu\text{g}.\text{ml}^{-1}$ ) amoxicillin and 20  $\mu\text{g}.\text{ml}^{-1}$  of cephalixin.**



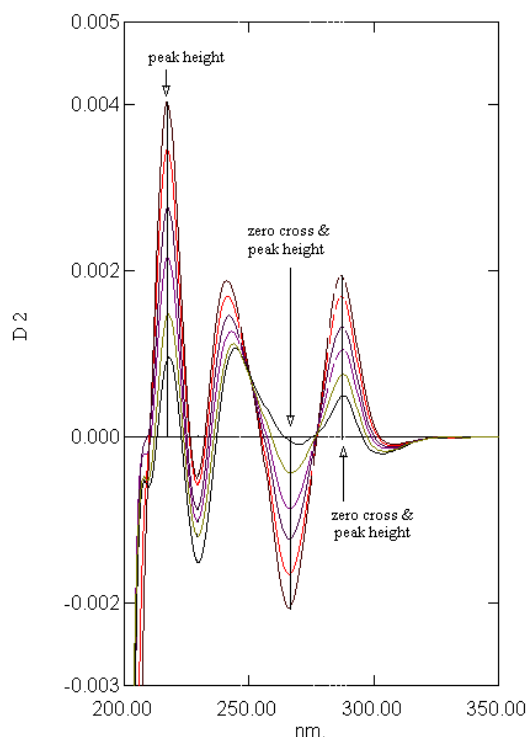
**Figure 5: First derivative spectra of mixtures containing (10 – 60  $\mu\text{g.ml}^{-1}$ ) cephalixin and 20  $\mu\text{g.ml}^{-1}$  of amoxicillin.**

Fig. 6 and Fig. 7 show further sets of second derivative of the same above mixtures. Applying the same mentioned techniques in measuring peak amplitudes (in millimeter) at

peak-to-base line and at zero crossing point of the other compound, and peak areas at selected wavelengths intervals enable the measurement of amoxicillin and cephalixin respectively (Table 1).



**Figure 6: Second derivative spectra of mixtures containing (10 – 60  $\mu\text{g.ml}^{-1}$ ) amoxicillin and 20  $\mu\text{g.ml}^{-1}$  of cephalixin.**



**Figure 7: Second derivative spectra of mixtures containing (10 – 60  $\mu\text{g.ml}^{-1}$ ) cephalixin and 20  $\mu\text{g.ml}^{-1}$  of amoxicillin.**

#### ***Calibration graph and statistical analysis***

The analytical characteristics and most statistical data for each of the proposed methods are given in Table 1. Under optimum conditions, linear calibration graphs were obtained in the range of (10 – 60  $\mu\text{g.ml}^{-1}$ ) with correlation coefficient values ranging between (0.9995 – 0.9842) and detection limits values in the range of (4.2-0.21  $\mu\text{g.ml}^{-1}$ ) for different techniques of measurements.

#### ***Accuracy and precision***

Under the optimum conditions, the accuracy and precision of the proposed method (two different techniques for each of first and second order derivative modes) were checked. Table 2 shows the values of relative error percent and relative standard deviation percent for two different level of

concentration of amoxicillin and cephalixin.

#### ***Application of the methods***

Two of the proposed methods (namely first derivative – peak-to-base at 236 nm and second derivative - peak-to-base at 244.6 nm) were successfully applied for direct determination of amoxicillin in two different drugs. The results obtained are presented in Table 3, and are in quite agreement with the spiked values. On the other hand, amoxicillin has also been successfully determined in two different pharmaceutical preparations by two of the proposed methods. The results are shown in table 3.

**Table 1: Statistical analysis of the determination of amoxicillin and cephalixin.**

Compound	Order of derivative	Mode of calculation	$\lambda$ (nm)	Regression equation	r	D. L. $\mu\text{g.ml}^{-1}$
Amoxicillin	First	Peak area	202-224	$Y=0.1105-0.01181 X$	-0.8921	6.20
	First	Peak area	224-262	$Y=-0.0135-0.00774 X$	-0.9987	0.56
	Second	Peak area	224-237	$Y=0.0001+0.00047 X$	0.9957	0.80
	Second	Peak area	237-259	$Y=0.0055+0.00057 X$	0.9991	2.00
	First	Peak to base line	236	$Y=0.0645+0.000485 X$	0.9987	0.21
	Second	Peak to base line	244.6	$Y=0.0004+(3.87e-5) X$	0.9993	0.27
Cephalexin	First	Peak area	225-254	$Y=0.2088-0.00245 X$	-0.9846	-
	First	Peak area	254-294	$Y=0.1426-0.01133 X$	-0.9991	2.00
	Second	Peak area	259-277	$Y=-0.0046+0.00052 X$	0.9984	0.51
	Second	Peak area	277-320	$Y=0.0001+0.00042 X$	0.9965	-
	First	Zero cross & peak- to-base line	275.7	$Y=-0.00365+0.00045X$	0.9992	3.40
	First	Zero Cross	223.9	$Y=0.00037+0.00037X$	0.9994	2.10
	Second	Zero cross & peak- to-base line	287.5	$Y=0.00017+(2.98e-5)X$	0.9976	0.83
	Second	Zero cross & peak- to-base line	266	$Y=-0.0003+(3.29e-5)X$	0.9986	4.13
	Second	Peak- to-base line	229	$Y= 0.0003+(6.27e-5)X$	0.9995	0.53

**Table 2: Precision and Accuracy of the methods.**

Compound	Method of analysis	Taken ( $\mu\text{g.ml}^{-1}$ )	Fond * ( $\mu\text{g.ml}^{-1}$ )	Relative error %	Relative standard deviation %
Amoxicillin	First order (peak-to-base line) at 236nm	10	10.213	+2.130	2.130
		60	60.460	+0.766	0.299
	Second order (peak-to-base line) at 244.6 nm	10	10.197	+1.970	0.568
		60	60.343	+0.572	0.165
Cephalexin	First order (peak area) in the range 254-294 nm	10	10.410	+4.100	0.748
		60	60.200	+0.333	0.135
	Second order (peak-to-base line) at 229 nm	10	10.243	+2.43	0.878
		60	60.388	+0.646	0.277

\* Average of four determinations.



**Table 3: Results for analysis of amoxicillin and cephalixin in four pharmaceutical formulation samples.**

Pharmaceutical preparation	Method of analysis	Labeled amount mg/tablet	Found amount mg/tablet		
			Mean value*	RSD%	E%
Amoxycillin S. D. I. - Iraq	First order (peak-to-base line) at 236nm	250	244.53	0.74	-2.18
		500	493.66	0.42	-1.27
	Second order (peak-to-base line) at 244.6 nm	250	245.90	0.64	-1.64
		500	494.10	0.40	-1.18
Pulmoxy MICRO LABS Ltd India	First order (peak-to-base line) at 236nm	250	242.67	0.98	-2.93
		500	491.00	0.44	-1.80
	Second order (peak-to-base line) at 244.6 nm	250	242.79	0.59	-2.88
		500	492.81	0.50	-1.44
Cephalexin S. D. I. -Iraq	First order (peak area) in the range 254-294 nm	250	250.77	2.10	+0.31
		500	498.60	0.92	-0.28
	Second order (peak-to-base line) at 229 nm	250	248.80	0.71	-0.48
		500	497.31	0.52	-0.54
Cefalexin APKES Ajanta-Pharma Ltd	First order (peak area) in the range 254-294 nm	250	244.10	1.98	-2.15
		500	489.61	0.93	-2.08
	Second order (peak-to-base line) at 229 nm	250	245.01	0.65	-1.99
		500	489.99	0.41	-2.00

\* Average of three determinations.

## References

- Vimal K. B., Ken A. T. and John T.; *Journal of Chromatography A*, 2006, **1131**(1-2), 1.
- Caro E., Marcé R. M., Cormack P. A. G., Sherrington D. C., Borrull F. ; *Analytica Chimica Acta*, 2005, **552**(1-2), 81.
- Gulkowska A., He Y., So M. K., Yeung L. W. Y., Leung H. W., Giesy J. P., Paul K.S. Lam, Michael M. and Bruce J. R.; *Marine Pollution Bulletin*, 2007, **54**(8), 1287.
- Choi K. J., Kim S. G., Kim C., Kim C. W., Kim S. H.; *Chemosphere*, 2007, **66**(6), 977.
- Lopez M. I., Pettis J. S., Smith I. B., Chu P. S.; Agric J.; *Food Chem*, 2008, **56** (5), 1553.
- Liu, S. P.; Hu, X. L.; Li, N. B.; *Analytical Letters*, 2003, **36** (13), 2805.
- Liu, S. P.; Hu, X. L.; Luo, H. Q.; *Analytical Sciences*, 2003, **19** (6), 927.
- Qin, Y.; Bao, L. J.; Zhu, L. M.; *Fenxi Ceshi Xuebao*, 2003, **22** (5), 60.

9. Jiang, H.; Liu, S. P.\*; Hu, X. L.; Liu, Z. F.; Zhan, H. L.; Kong, L.; Qin, Z. H.; *Fenxi Huaxue*, 2003, **31**( 10 ), 1207.
10. Askal H. F., Refaat I. H., Darwish I. A. , Marzouq M. A.; *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 2008, **69**(4), 1287.
11. El-Sayed, A. Y. and Khalil, M. M. H.; *Talanta*, 1996, **43**, 583.
12. El-Sayed, A. Y., Abdel Rahem, M. A. and Omran, A.A.; *Analytical Sciences*, 1998, **14**, 577.
13. Koleva B. B., Kolev T. M., Spiteller M.; *Journal of Pharmaceutical and Biomedical Analysis*, 2008, **48**(1), 201 .
14. O' Haver, T. C. and Green, G. L.; *Anal. Chem.*, 1976, **48**, 312.