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# Traditional and molecular diagnosis of *Haemonchus contortus* in sheep in Babylon province, Iraq

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#### **Abstract**

Haemonchus contortus one of gastrointestinal helminthes characterized by miner intraspecific variation and a major variation among species which exploited to determine species simultaneously depend on PCR techniques, by amplification of DNA from parasites so goal of study was traditional and molecular detection of this parasite in sheep. Experiment was conducted using adult worms collected from abomasum of sheep obtained from abattoir in Babylon province. All worms divided into two groups, first group for morphological study and second group stored in 70% ethanol for DNA extraction, ITS-2 spacer and 28S gene were amplified using PCR technique sequenced and analysis with a phylogenic tree. According to the available data this study recorded Haemonchus contortus in sheep depend on ITS-2 spacer and 28S gene sequences for the first time in Iraq with accession no. LC552170 and LC552171 using molecular data. The phylogeny analysis depended on ITS-2 spacer and 28S gene partial sequences were closely related and high identity 94% with Germany H. similis sequence MN708992.1 and 93% identity with New-Zealand and Austria H. contortus sequence KC998713.1 and KJ724288.1 respectively, with a low genetic variation among all comparison sequenced isolates.

DOI: 10.33899/ijvs.2021.130533.1842, @Authors, 2022, College of Veterinary Medicine, University of Mosul. This is an open access article under the CC BY 4.0 license (<a href="http://creativecommons.org/licenses/by/4.0/">http://creativecommons.org/licenses/by/4.0/</a>).

#### Introduction

Gastrointestinal helminthosis infections in ruminants represent the major problems and the major factors responsible for lowered economics of sheep production (1). Haemonchus ssp. infection clinical signs include hypoproteinemia, epithelium hyperplasia within nodules affects the full mucosal necropsy thickness, weakness, collapse and death because of unspecific signs (2,3). In addition, it reported that Haemonchus was a blood-feeding parasite responsible for important morbidities and mortalities in sheep (4). The major signs of this parasite anemia, pale mucous membranes, submandibular edema, and sudden death (5). acute haemonchosis form occurs with varying rates affected animals appeared weaker depending on intake of infective larvae and blood loss rate, while chronic form characterized by weight loss, poor weight

gain, general diagnosis and anemia detected by assessment of conjunctiva membrane color (4). Drug resistance due to excessive and uncontrolled anthelmintic use lead to problems with genetic resistance at high gene flow level to this parasite and enhance resistance (6). Due to the few published studies about genetic diversity that depend on genome-wide data, high polymorphism rate in *Haemonchus* reference sequences make discernable differentiation between H. contortus populations in different host species challenge for genome assembly (7). The species identity and comparison of H. contortus isolated from sheep and goat indicated that it was more closely related to each other depended on internal transcribed spacer ITS-1, ITS-2 and the 5.8S rRNA regions (8). Many studies depend on ITS sequence indicated that it this region was good genetic marker for distinguishing sympatric *Haemonchus* ssp. (7,8).

# Materials and methods

Total 67 sheep samples were checked in study, abomasum was obtained from slaughtered sheep at Babylon province local abattoir, abomasum were placed into a bucket containing physiological saline and transported to Laboratory of Parasitology, Baghdad. Research had been done between June 2019 to May 2020. A total of 86 worms were collected from abomasum continent washed with PBS at pH 7.4 prepared by dissolved 9.86 g of phosphate buffer powder in 1000 ml of distill water according to the manufacture instruction and adjusted PH to 7.2 for use. All worms were divided into two groups, first group loaded into a lactophenol for diagnosis species level of body, cervical papillae, esophagus and vulval distance and morphology to detected morphological characters depend on light microscope (40x) diagnosis (9).

#### **Genomic DNA isolation**

The second group of the worms was stored in 70% ethanol; the genomic materials were isolated according to (Qiagen, Germany Kit) directive. DNA estimation for all samples used a Nanodrop (ThermoScientific, USA) reading absorbent at (260 /280 nm); samples were stored at-20°C (10).

# **PCR** amplification

Depended on Shen *et al.* (11) about ~300 bp fragment encompassing the nuclear ribosomal DNA ITS-2 spacer and 28S gene was PCR amplified by added 2 µl of individual worm genomic DNA as a template, 1 µl of each forward ITS-2F-28S-5'-ACGTCTGGTTCAGGGTTGTT-3' and reverse ITS-2R-28S-5'-TTAGTTTCTTTTCCTCCGCT-3' with 10 (pcoml) to master mix (Maxime PCR PreMix) then complete reaction volume to 20 µl by nuclease-free water. All above were used without DNA template as a negative control; all the tubes were transported to Exispin vortex centrifugation for 1 minute at 2000 rpm, and positioned in PCR Thermo-cycler.

# PCR thermocycler conditions and electrophoresis

A PCR thermo-cycler condition was done by utilizing a conventional PCR thermo-cycler (T100 thermal cycler. BioRad USA) (Table 1). To detected PCR products and to verify represented single bands, Agarose gel 1% concentration was prepared according to (12) as 1 gm agarose dissolved in 1X TBE, heated in microwave 1 min, then cool down at 60°C.

#### DNA sequence and phylogenic tree analysis

Positive products of PCR were sent by ice bag by DHL to Macrogen Company in Korea for completed DNA sequencing by AB DNA sequencing method. Sequence data were deposited in Gene-Bank and get accession number to identify genetic variation between Iraqi sequences isolates and other sequences that submitted in NCBI. Sequencing of

DNA study was directed using utilizing Mega 6.0. The development distances were computed utilizing; Phylogenetic analyses were carried out employing (Phylogny.fr / advanced method).

Table 1: PCR thermo-cycler system conventional

PCR steps	Temp.	Time	Cycles
Initial Denaturation	95°C	5 minutes	1
Denaturation	95°C	30 second	
Annealing	52°C	30 second	35
Extension	72°C	1 minute	
Final extension	72°C	5 minutes	1
Hold	4°C	-	-

#### Results

# Morphological identification

The results showed that 50 adult worms, 36 adult female and 14 male worms were *Haemonchus spp.*, all obtained female samples were detected by their body length, which demonstrated 29.75 mm, cervical papillae recorded 4.55 mm from head (Figure 1), valval flap and uterus morphology was also detected (Figure 2). In male nematodes, the results recorded a morphological attribute that include; body length which demonstrated 18.16 mm as a male average length, 4.9 mm of left and right spicules, barbed tip, and gubernaculum 8.9 mm in length (Figure 3).

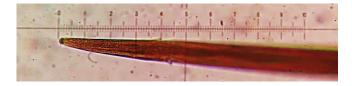


Figure 1: cervical papillae of *Haemonchus spp.* 40X-light microscope.

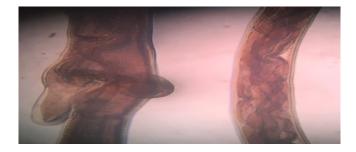


Figure 2: Vulval flap and uterus of *Haemonchus spp.* 40X-light microscope.

# **Genomic study results**

Total genomic DNA were extracted from individual worm's samples (second group of worms) with high concentration and purity measurement 350 ng/ $\mu$ l and 1.74 respectively.

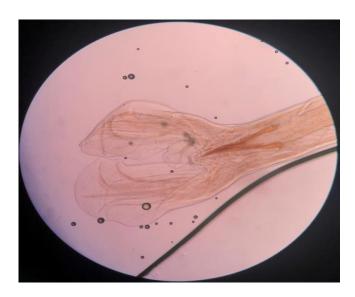


Figure 3: male spicules, barbed tip, and gubernaculum of *Haemonchus spp.* 40X-light microscope.

# PCR amplification and sequencing

Specific oligonucleotide primer pairs for ITS-2 spacer and 28S gene we're using with purified worm's DNA as a template in PCR technique were exhibited successfully amplified showing a bands of DNA fragment that indicated the presence of positive results for *Haemonchus*. Representative bands are compared to 100bp ladder in electrophoretic with 1% agarose concentration (Figure 4). Positive PCR product were sequenced and deposed in gene bank NCBI with accession numbers LC552170 and LC552171 at DDBJ and ENA databases for the first time in Iraq.

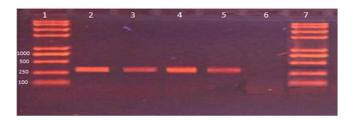


Figure 4: Gel electrophoresis for PCR product of 289bp using (1%) agarose for 90 minutes at 80 volts. Lance (1) molecular marker, Lances (2-5) positive samples for *Haemonchus*, and Lance (6) negative control.

# Phylogenic tree analysis

The phylogeny depended on ITS-2 spacer, and 28S gene sequences of neighbor-joining analysis in length sequence was approximately 289 bp editing and alignment using (Phylogny.fr, advanced method). The results indicated that all Iraqi *Haemonchus spp.* isolates in this study were *Haemonchus contortus* and recorded a low genetic variation (0.01) among all comparison isolates (Figure 5).

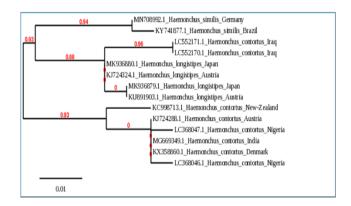


Figure 5: Phylogenetic tree of *Haemonchus contortus*.

#### **Discussion**

Many researchers detected specific primers for different genes for *Haemonchus* PCR diagnosis (13). The current study investigated primers that amplify ITS-2 spacer and 28S gene for *Haemonchus* with amplicon size 289 bp as amplifiable targets for PCR that did not match with parasites that infect sheep, which indicated of PCR technique high sensitivity for parasites detection (14,15). Sequence analyses of ITS2 spacer and 28S gene partial sequence were obtained from all worm's specimens that isolated from sheep abomasum and positive amplified by PCR were used to determine taxonomic status of *Haemonchus spp*. for the first time in Iraq, since other studies indicated for many years DNA sequencing was available and good technology to differentiated closely related parasites species (16).

Nucleotide ITS-2 spacer sequencing, BLASTEn alignment, and graphic analysis presented highly similar sequences (megablast) found in NCBI database after sequences blasted at BLASTn program and the study revealed that sequence alignment of ITS-2 spacer and 28S gene lies in Haemonchus group with different max score 534 and equal to the total score, E-value were equal to 8e-148 and identity 100% as clarified in NCBI database, and that agree with Jassem et al. (17) who indicted that gene sequencing was an excellent target to differentiation isolates by molecular detection. Phylogeny of Iraqi Haemonchus contortus isolates were closely related and high identity with Germany and Brazil H. similis sequence and showed 93-94% identity respectively, 88% identity with those of Japan, Austria H. longistipes sequence. Additionally, there were 93% identities with H. contortus sequences of New-zealand, Austria, Nigeria, India, Denmark and Nigeria. The previous studies have shown that the sequencing followed by phylogenic tree analysis was used to detect parasite species (18,19).

Some previous studies have indicated that diagnosis of parasites species based on their morphological characters alone may give imprecise results for accurate identification, due to the interaction between the many morphological parameters between *Haemonchus spp.* isolates such as parasite length, spicules length, spicules barb length, and many other parameters, as well as the difficulties that the researchers could be facing them in microscopic examination (20).This interaction will lead measurements overlapping between species discriminate function in species identification, so other techniques like immunological, histopathological, or molecular methods were required (19-22).

# Conclusion

ITS-2 spacer and 28S gene was an excellent molecular marker for detected *Haemonchus spp.*, sequencing followed by phylogeny confirmed the phylogenetic relationship of all species recorded in NCBI database determines genetic affinity percentage among them.

# Acknowledgments

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#### **Conflict of interest**

The authors declare that there are no conflicts of interest regarding the publication of this manuscript.

# References

- Thomas N, Teshale S, Kumsa B. Abomasal nematodes of sheep and goats slaughtered in Awassa (Ethiopia): Species composition, prevalence and vulvar morphology. Helminthol. 2007;44(2):70-75. DOI: 10.2478/s11687-007-0006-8
- Simpson HV, Lawton D E, Simcock DC, Reynolds G W, Pomroy WE. Effects of adult and larval *Haemonchus contortus* on abomasal secretion. Inter J Parasitol. 1997;27:825-831. DOI: 10.1016/S0020-7519(97)00037-4
- Scott I, Stear M J, Irvine J, Dick A, Wallace D S, M ckellar Q A. Changes in the zymogenic cell populations of the abomasa of sheep infected with *Haemonchus contortus*. Parasitol. 1998;116:569-577. DOI: 10.1017/S0031182098002704
- Besier IRB, Kahn LP, Sargison ND, Van JA. Diagnosis, treatment and management of *Haemonchus contortus* in small ruminants. Advan Parasitol. 2016;93:181-238. DOI: <u>10.1016/bs.apar.2016.02.024</u>
- Gasser RB, Bott NJ, Chilton NB, Hunt P, Beveridge I. Toward practical, DNA based diagnostic methods for parasitic nematodes of livestock-bionomic and biotechnological implications. Biotechnol Adv. 2008;26(4):325-34. DOI: 10.1016/j.biotechadv.2008.03.003
- Yin F, Gasser RB, Li F, Bao M, Huang W, Zou F. Genetic variability within and among *Haemonchus contortus* isolates from goats and sheep in China. Parasit Vectors. 2013;6(1):279-84. DOI: 10.1186/1756-3305-6-279
- Laing IR, Martinenelli A, Tracey A, Holroyd N, Gilleard JS, Cotton JA. Haemonchus contortus: Genome structure, organization and comparative genomics. Advan Parasitol. 2016;93:569-98. DOI: 10.1016/bs.apar.2016.02.016
- Umer C, Elizabeth MR, Muhammad A, Raman M, Kamran A, John SG. Genetic evidence for hybridisation between *Haemonchus*

- contortus and Haemonchus placei in natural field populations and its implications for interspecies transmission of anthelmintic resistance. Inter J Parasitol. 2015;45(2-3):149-159. DOI: 10.1016/j.ijpara.2014.09.002
- Mirabeau MN, Gedeon PM, Paul ANM, Michael R, Meral E, Ayola AA. Diagnostic techniques of soil-transmitted helminths: Impact on control measures. Trop Med Infect Dis. 2020;5:93. DOI: 10.3390/tropicalmed5020093
- Isihak F A. Diagnosis of reovirus infection in broiler breeders flocks by using PCR technique in Erbil province. Iraqi J Vet Sci. 2020;34(1):77-81. DOI: <u>10.33899/ijvs.2019.125469.1007</u>
- Shen DS, Ji-fei W, Dan-yu Z, Zhi-wei P, Tian-yun Y, Zhao-ding W, Dwight D B, Zhi-jun H, Zhen-sheng L. Genetic diversity of Haemonchus contortus isolated from sympatric wild blue sheep (Pseudois nayaur) and sheep in Helan Mountains, China. Int Parasites Vectors. 2017;10:437. DOI: 10.1186/s13071-017-2377-0
- Ismael SS, Omer LT. Molecular identification of new circulating *Hyalomma asiaticum* asiaticum from sheep and goats in Duhok governorate, Iraq. Iraqi J Vet Sci. 2021;35(1):79-83. DOI: 10.33899/ijvs.2020.126330.1298
- Zarlenga DS, Hoberg EP, Tuo W. The Identification of *Haemonchus* species and diagnosis of Haemonchosis. Adva Parasitol. 2016;93:145-180. DOI: 10.1016/bs.apar.2016.02.023
- AlFatlawi MA, Ismail YK, Ali MJ, Karawan AC, Al-Ibadi IN. Molecular differentiation of *Thysaniezia* (Helictometra) *giardi* and *Moniezia* species based on 18s rRNA gene in small ruminants. Iraqi J Vet Sci. 2021;35(1):105-108. DOI: 10.33899/ijvs.2020.126407.1313
- Hypy A, Salah AS, Mona MS, Mohamed A, Mahmoud E, Hanan HA. High prevalence of *Coxiella burnetii* infection in humans and livestock in Assiut, Egypt: A serological and molecular survey. Vet World. 2020;13:2578-86. DOI: <u>10.14202/vetworld.2020.2578-2586</u>
- Faraj AA, Hade BF, AlAmery AM. Conventional and molecular study of *Babesia spp*. natural infection in dragging horses at some areas of Baghdad city, Iraq. Iraqi J Agricul Sci. 2019;50(3):909-915. DOI: 10.36103/ijas.v50i3.707
- Jassim A, Alfatlawi A M, Jarad N I, Klaif S F. Clinical and molecular identification of ruling *Theileria annulata* strains in cattle calves in Al-Diwaniyah province, Iraq. Iraqi J Vet Sci. 2021;35(1):115-119. DOI: 10.33899/ijvs.2020.126429.1319
- Vera M, Herjuno AN, Zahrah PA, Yura DR, Surbakti BR, Rini W. Genetic characterization and phylogenetic study of Indonesian indigenous catfish based on mitochondrial cytochrome B gene Dorothea. Vet World. 2020;(13):96-103. DOI: 10.14202/vetworld.2020.96-103.
- Hade BF. Molecular sequencing and phylogenic analysis to virulence nmuc-1 gene in visceral larvae migrance. Iraqi J Agricul Sci. 2020;51(3):894-902. DOI: 10.36103/ijas.v51i3.1044.
- Santos MC, Amarante MR, Silva MR, Amarante AF. Differentiation of *Haemonchus placei* from *Haemonchus contortus* by PCR and by morphometrics of adult parasites and third stage larvae. Rev Bras Parasitol Vet. 2014;23:495-500. DOI: <u>10.1590/S1984-29612014085</u>.
- Alhayali N S, Hasan M H, Al-Mallah K H. Natural heavy infection with immature sarcocysts of *Sarcocytis* spp. in sheep in Mosul city: A case report. Iraqi J Vet Sci. 2020;34(2):373-376. DOI: 10.33899/ijvs.2019.125994.1210
- 22. Hade BF, Al-Amery AM, Ibrahim Z, Saadedin S. Histopathological study of different VLM Stages *Toxocara canis* infection in liver of rabbits. Iraqi J Biotecnol. 2018;17(2):47-56. [available at]

# التشخيص التقليدي والجزيئي للدودة السلكية الملتوية في الأغنام في محافظة بابل، العراق

بلقيس فاضل هادي، سهى طارق البياتي و حيدر محمد على الربيعي

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#### الخلاصة

الدودة السلكية الملتوية أحد أنواع الديدان التي تتميز بمستوى منخفض من التباين داخل النوع وتباين مرتفع بين الأنواع لذلك تم الاعتماد على التقنيات الجزيئية لتشخيص الطفيلي اعتمادا على تصخيم المادة الوراثية. الهدف من الدراسة هو الكشف عن الطفيلي في الأغنام باستخدام الطرق التقليدية والجزيئية في التشخيص. أجريت الدراسة باستخدام ديدان بالغة تم جمعها من منفحة الأغنام والتي تم الحصول

عليها من مجزرة في محافظة بابل إذ تم نقلها إلى مختبر الطفيليات، كلية الطب البيطري، جامعة بغداد لغرض الدراسة. تم تقسيم جميع الديدان إلى مجموعتين، المجموعة الأولى للدراسة الشكلية التقليدية للطفيلي والمجموعة الثانية المحفوظة في ٧٠ ٪ من الإيثانول الستخراج المادة الوراثية للطفيلي، إذ تم تضخيم منطقة 2-ITS والجين 28S باستخدام تقنية تفاعل البلمرة المتسلسل وتسجيل متتابعات القواعد النيتروجينية و التحليل الجزيئي للنتائج باستخدام شجرة التطور الوراثي. نتائج الدراسة تؤكد تسجيل الدودة السلكية الملتوية في الأغنام اعتماداً على منطقة 2-ITS وجين 28S لأول مرة في العراق بالأرقام LC552170 و LC552171 إذ استخدام المتتابعات الوراثية للقواعد النيتروجينية ل منطقة 2-ITS و جين 28S للعزلة العراقية التي ارتبطت ارتباطًا وثيقًا وبنسبة عالية ٩٤ ٪ مع طفيلي H. similis MN708992.1 للعزلة الألمانية بنسبة ٩٣ ٪ مع نتابعات الدودة السلكية الملتوية النمساوي ونيوزيلندا KC998713.1 و KJ724288.1 على التوالي، مع تسجيل اختلاف وراثي منخفض بين جميع عز لات المقارنة.