

Article review

Some Classes of epipelic in Iraqi waters

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Abstract.

This review aimed to survey some classes of epipelic composition in Iraqi waters. These data were taken from investigations of epipelic in Iraq, covering both freshwater and saline waters in many cities. 266 species of epipelic algae belonging to 133 genera were identified. Bacillariophyta (diatoms) was the predominant taxon (161 species) followed by Chlorophyta (53 species), Cyanophyta (41 species), Euglenophyta (8 species) and Dinophyta (3 species).. This study proposes to present a checklist of epipelic in Iraq, built on the compilation of preceding surveys.

Keywords: review, epipelic, classes, Iraqi water.

Introduction

The importance of benthic algae in Iraqi aquatic ecosystems was subjected to investigations recently . Many authors have previously recognized epipelic algae as bioindicators in different aquatic ecosystems of Iraq. Benthic algae are an important part of algae community and have been widely used as a tool to assess water quality[1]

The studies of epipelic algal ecology was pioneered in freshwater habitats by Round [2, 3, 4, 5, 6]. Lately, Algae can be found in all aquatic systems, and are the most diverse assemblage of organisms that can be sampled easily and identified readily to species or varieties [7] Epipelic can be the heralds of infection or decease. Some of them create biotoxins, making these kinds accountable for blooms or “red tides,” or harmful algal. These toxins can cause irritation in the respiratory system in creatures and other risks [8].

In Iraq, abundant efforts have been used on distribution and identification of epipelic. So, this review offered data of the composition of non-diatom algal taxa in Iraq.

Sources and Methods

Literature review. Data were taken from thirty-one reference (22 research papers, four Ph. D. thesis, and five M. Sc. thesis, dealing with the epipelic. Data from the references were gathered to make the current list of epipelic. The references are (9 - 44): Ali, *et al.* (2018), Hassan, *et al* (2017), Salman ,*et al*(2013), Al Fatlawi (2011), Hassan, *et al.*, (2014), Hassan and Shaawait (2013), Hassan and Al- Budulameer(2012), Abbas and Hassan(2018), Hadiand

Al-Saboonchi.(1989), Kssaim and Al- lami (2002), Kassim.andAl-Saadi,(1994), Kassim (2007) Al-Timimi,. (2012), Al-Ghanmi (2011), Al-Nashi (2012), Salman, *et al.* (2013), Hindi. (2016), Al-Saboonchi (1995), Kassim. (2007), Al-Tae, (2010), .Kadhim *et al.*, (2013) Al-Saadi and Hadi (1987), Hassan, And Shaawiat,(2015) AL-Lami, Al-Shawi (1999), Maulood. and Toma (2004), AL-Lami,*et al.*, (1999), Al-Shaban (1996), Kadhim, *al.*, (2013).

Alkam., and Al-Nashi, (2013) AL-Saadi *et al* .,. (1979), Hassan, *et al.* (2014), Hassan, *et al.* (2010), Hassan, (1995), Hassan, *et al.* (2007), Al-Zubaidi(2012),

Kassim and Mukai(2006), Khthim and Al-Amari and (2013), Hassan *et al.*, (2017), Al-Handal(1992)

Results and discussion :

This checklist of non-diatom epipelagic taxa has 133 taxa (266 recognized to species level) which vary among four division (table1). The variety of epipelagic present in Iraqi waters referred to their diverse limnological characteristics. The factors effect on qualitative of plankton. Uncontaminated ecosystems are recognized via high values of diversity [2, 4, 5]. While, the existence and domination of some algae as *Nitzschia*, *Navicula*, *Cymbella*, *Gomphonema*, *Surirella*, *Cocconeis*, *Aulacoseira*, *Oscillatoria*, *Lyngbya*, *Spirulina*, and *Scenedesmus* , referred to organic contamination in the water [1, 3, 6].

As a outcome, this review included five divisions, among these divisions, Chlorophyta was dominant, representing 19.29. % of the detected taxa (27taxa and 53 species), followed via Cyanophyta (14 taxa and 41 species). Euglenophyta (2 taxa and 8 species), Dinophyta (22 taxa and 21 species), Almost all surveys verified Chlorophyta was dominated from non diatom divisions (20, 64). Other divisions as, Cyanophyta, Dinophyta, and Euglenophyta had fewer domination, this may be because they have narrow number of species, likewise they want many nutrients [2, 4, 5]. This review listed 266286 species only, because rather insufficient surveys on epipelagic from Iraqi waters.

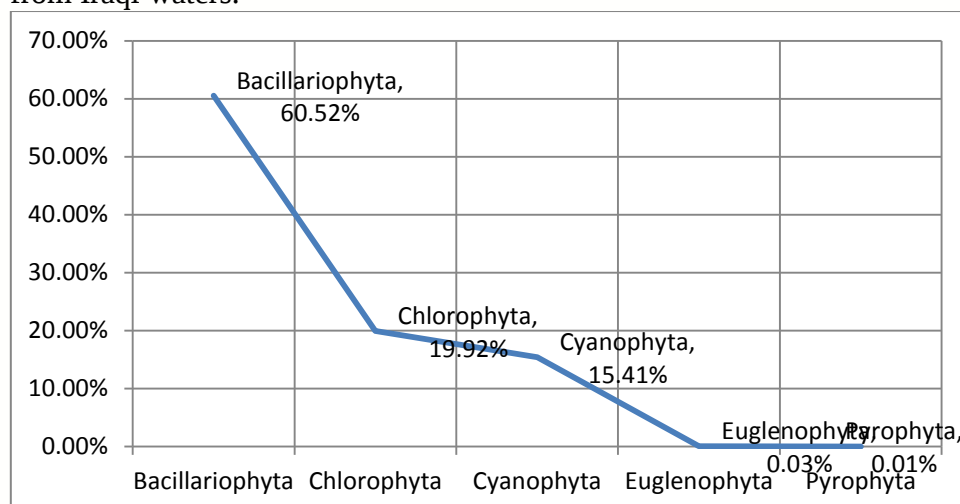


Figure1.The percentage of Divisions in this review

Table 1.List of epipelagic algae that found in Iraq

Division	Taxa
Bacillariophyta centrales	
	<i>Aulacoseira ambigua</i> (Grunow) Simonsen <i>A. granulata</i> (Ehr.) Simonsen <i>Coscinodiscus asteromphalus</i> Ehrenberg <i>C. granii</i> L.F.Gough <i>C. denarius</i> A. Schmidt <i>Cyclotella comta</i> Kützing <i>C. meneghiniana</i> Kützing <i>C. ocellata</i> Pantocsek <i>Melosira. ambigua</i> (Grunow) Otto Müller <i>M. granulate</i> (Ehr.) Ralfs <i>M. varians</i> Agardh <i>Stenopterobia delicalissinia</i> (F.W.Lewis) Brébisson ex Van Heurck <i>Stephanodiscus agassizensis</i> Håkansson & Kling <i>Thalassiosira baltica</i> (Grunow) Ostenfeld
Pennales	

Achnanthes affinis Grunow
A. minutissima var. *affinis* (Grunow) Lange-Bertalo
A. longipes
A. microcephala (Kütz.) Grunow
Achnanthidium exiguum
Grunow
Amphora bioculata Cleve
A. ovalis (Kütz.) Kützing
A. ovalis var. *pediculus* (Kütz.) Van Heurck
A. pediculus (Kütz.) Grunow ex A.Schmidt
A. veneta Kützing
Anomoeoneis sphaerophora Pfitze
Bacillaria paradoxa J.F.Gmelin
B. paxillifera (O.F.Müller) T.Marsson
Caloneis amphisbaena (Bory.) Cleve
C. amphisbaena var. *subsalina* (Donkin) Cleve
C. bacillum (Grunow) Cleve
C. permagna (Bailey) Cleve
C. ventricosa (Ehr.) F.Meister
Cocconeis pediculus Ehrenberg
C. placentula Ehrenberg
C. placentula var. *euglypta* (Ehr.) Cleve
C. amphisbaena var. *subsalina* (Donkin) Cleve
C. placentula var. *lineata* (Ehr.) Van Heurck
Cymatopleura elliptica (Bréb) W.Smith
C. aspera (Ehr.) Cleve
C. Agardh
C. cistula (Hemp.) Grunow
C. lanceolata (C.Agardh) Kirchner
C. lange-bertalotii Krammer
C. ovalis (Kütz.) Brébisson & Godey
C. prostrate (Berk.) Cleve
C. solea (Breb.) W.Smith
C. solea var. *apiculata* (W.Smith) Ralfs
C. tumida (Breb.) Van.Heurck
C. turgid (Greg.) Cieve
C. turgida (Greg.) Cleve
C. turgidula var. *kappii* Chohnoky
C. ventricosa (C.Agardh)

Diatoma vulgare Bory
Diploneis elliptica(Kütz.) Clev
Encyonema reichardtii(Krammer) D.G.Mann
E. silesiacum(Bleisch) D.G.Mann
Fragilaria .capucina var. *gracilis* (Oestrup) Huste
F. capucina var. *rumpens*(Kütz.) Lange-Bertalotex
Bukhtiyarova
F.capucina Desmazieres
F. construens (Ehr.)Grunow
F. construens f. *binodis* (Ehr.) Husted
F. elliptica Schumann
F. intermedia Grunow
F. ulna (Nitzsch) Lange-Bertalot
F.ulna var. *acus* (Kütz.) Lange-Bertalot
F. ulna var. *biceps*(Kütz.) Compère
F. virescens Ralfs
Gomphoneis olivecum(Horne)P.Dawson
ex. Ross et Simith
Gomphonema. affine Kützing
G. clavatum Ehrenberg
*G. insignaffine*E.Reichardt
G. lagenula Kützing
G. mesta (S.I.Passy-Tolar &R.L.Lowe) E.Reichardt
G. minuta P.Fusey
G. minutum (C.Agardh)C.Agardh
G. olivaceum (Lyng.)Kützing
G. paravalum (Kütz.)Grunow
G. rhombicum Fricke
G. truncatum Ehrenberg
Gyrosigma acuminatum(Kütz.) Rabenhorst
G. attenuatum (Kütz.)Rabenhorst
G. fasciola (Ehr.)J.W.Griffith & Henfrey
Mastogloia elliptica(C.Agardh) Cleve
M. smithii Thw. Ex. W. Smith
Mayamaea atomus(Kütz.) Lange Bertalot
Navicula affinis Ehrenberg
N. ambigua Ehrenberg
N. bacillum Ehrenberg
N. capitatoradiata H.Germain
N. cryptocephala Kützing
N. rhynchocephala Kützing
N. seminulum Grunow
N. veneta Kützing
N. viridis (Nitzsch) Ehrenberg

	<i>S. capitata</i> Ehrenberg <i>S. fasciculate</i> (C.Agardh) Kützing <i>S. rumpens</i> Kützing <i>S. tenera</i> W.Smith <i>S. ulna</i> (Nitzs.) Ehrenberg <i>S. ulna</i> var. <i>biceps</i> (Kütz.) Schönfeldt <i>Tabellaria fenestrata</i> (Lyng.) Kützing
Chlorophyta	
	<i>Actinastrum hantzschii</i> Lagerheim <i>Ankistrodesmus falcatus</i> (Cord.) Ralfs <i>Asterococcus limneticus</i> G.M. Smith <i>A. superbus</i> (Cienk.) Scherffel <i>Carteria klebsii</i> (Dang.) Dill <i>Characium limneticum</i> Lemmermann <i>Chlamydomonas angulosa</i> Dill <i>C. globosa</i> Snow <i>Chlamydomonas</i> sp <i>Chlorella bejerinck</i> <i>Chlorella</i> sp <i>Closteriopsis longissima</i> Lemmermann <i>C. strigosum</i> Berb. <i>Coelastrum reticulatum</i> P.A.Dangeard <i>C. microporum</i> Nägeli <i>Cosmarium botrytis</i> Meneghinii <i>C.formosulum</i> Hoff <i>C. granatum</i> De Brébisson <i>C.subcostatum</i> Nordstedt <i>Cosmarium</i> sp. <i>Crucigenia vectangularis</i> (A. Braun) Gay. <i>Dactylococcopsis</i> sp <i>Eudorina elegans</i> Ehren <i>Geminella interrupta</i> (Turp.) Lagerheim <i>Gloeotrichia natans</i> (Hedwig) Rabenhorst <i>Kirchneriella contorta</i> (Schmidle) Bohlin <i>Meringosphaera spinosa</i> prescot <i>Oedogonium cardiacum</i> (Hass.) Wittrock <i>Pediastrum simplex</i> Meyen <i>Pediastrum</i> sp <i>Sceneds quadricauda</i> (Turp.) de Brébisson <i>Scenedesmus armatus</i> Chodat <i>S. abundans</i> var. <i>brevicauda</i> G.M. Smith <i>S. aculeolatus</i> <i>S. arcuats</i> (Lemmermann) <i>S.bijuga</i> (Turb.)Lagher <i>S. ecorins</i> <i>S. quadricanda</i> <i>S. quadricanda</i> var. <i>Longispina</i> (Chod.) G.M. Smith <i>S.dimorphus</i> (Turb.)Ktz. <i>S. quadricauda</i> var <i>westii</i> <i>Selanastrum</i> sp. <i>Schizochlamys gelatinosa</i> A. Braun <i>Spirogyra scrobiculata</i> (Stock.) Czurda <i>Tetrademus wisconsinense</i> G.M. Smith <i>Tetraedron hastatum</i> (Reisch)Hansg <i>T. minimum</i> (A. Braun Hansging

	<i>Trochiscicia granulate</i> (Reinsch) Hansging <i>Ulothrix</i> sp. <i>U. aequalis</i> Kuetzing <i>U. variabilis</i> Kuetzing <i>Volvox</i> sp. <i>Westella linearis</i> G.M. Smith <i>Zygnema</i> sp.
Cyanophyta	
	<i>Anabaena</i> sp. <i>Aphanocapsa rivularis</i> (Carm.) Rabenhorts <i>Arthrospira</i> sp <i>Calothrix</i> sp. <i>Chroococcus disperses</i> (keissl) Lemmerman <i>Chroococcus limneticus</i> Lemmermann <i>Chroococcus turgidus</i> (Ktz.) Naegeli <i>Gomphosphaeria aponina</i> Kützing <i>Lyngbya aestuarii</i> Lemmermann <i>Lyngbyanordgaardii</i> Wille <i>Lyngbya limnetica</i> Lemmermann <i>Lyngbya perelgeaus</i> Lemmerman <i>Lyngbya</i> sp <i>Merismopedia elegans</i> A. Braun <i>M. glauca</i> (Ehr.) Naegeli <i>M. punctata</i> Meyen <i>Nostoc</i> sp. <i>Oscillatoria agardhii</i> Gomont <i>O. amphibian</i>
	<i>O. chalybea</i> Mertens <i>O. Formosa</i> Bory <i>O. granulatan</i> Mertens <i>O. laete-virens</i> (Crouan) Gomont <i>O. limnetica</i> <i>O. ornate</i> (Ktz.) Gomont f. <i>planetonica</i> Elenkin <i>O. princeps</i> Agardh <i>O. sancta</i> (Ktz.) Gomont <i>O. subbervis</i> Schmidle <i>O. sublimformis</i> Kutz <i>O. tenuis</i> Agardh <i>O. terebriformis</i> Agardh <i>O. sp.</i> <i>Phormidium subfuscum</i> Ktz. <i>P. sp</i> <i>Spirulina laxa</i> G.M. Smith <i>Spirulina major</i> Kützing <i>S. princeps</i> (West and west) G.S. West <i>S. subsalsa</i> Oersted
Euglenophyta	
	<i>Euglena acus</i> Ehernberg <i>E. elongate</i> Schewiakoff <i>E. gracilis</i> Klebs <i>E. proxima</i> Dangeread <i>E. spirogyra</i> Ehernberg <i>Phacus caudatus</i> Huebner <i>P. sp.</i>

	<i>Trachelomonas acanthostoma</i> (Stoken) De Flander
Dinophyta	
	<i>Glenodinium pulvisculus</i> (Ehr.)Stein <i>G. quadriden</i> <i>G.sp.</i>

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