Some biochemical effects of *lupinus albus* L. seed oil on normal and alloxan induced diabetic mice

Fatima A.Mohammad

Department of Chemistry, College of Science, Mosul University, Mosul, Iraq (Received 10/6/2008, Accepted 15/12/2008)

Abstract:

In this research *Lupin albus* L. seed oil had been extracted and then identification of it is contents from the fatty acids and lipids which was compared with standard fatty acids and lipids by using thin layer chromotograph.

Also the research has been dealed with the effects of increasing concentrations of (*Lupin albus* L.) seed oil on some biochemical parameters in serum and tissue of normal and alloxan-induced diabetic mice. When administrated orally (50,100,200) mg/Kg body weight of lupin seed oil the results indicated a significant decrease of serum glucose levels at 100 mg/kg body weight in normal and alloxan-induced diabetic mice the percentage of decrement was (-25.51% and -29.85%) respectively and then determined copper concentration in lupin oil by using atomic absorption spectrophotometer.

Also noticed that the level of cholesterol in serum of normal and alloxan induced diabetic mice was significantly lowerd, the percentage of decreasing was (-22.39 and -23.09%) respectively.

This research refer to significant decrease in liver glycogen content in normal mice of (-23.29%) after 180 min of administration of oil relative to control value.

On the other hand, lupin seed oil has non significant difference on the concentration of liver glutathione and heart malondialdehyde relative to control value.

Key word: lupinus albus, diabetes, cholesterol, glycogen, glutathione, malondialdehyde.

Introduction:

Lupins have been cultivated for over 2000 years, originating around the Mediterranean and along the Nile valley where they were used for human consumption. Lupins have dijitate leaves [1], annual plant belonging to the class of leguminosae and of Rosales order, dicotyledons class and spermatophyta phylum, with flower terminating receme upto 1.5 m high [2]. Lupins whole seed contain protein, lipids fatty acids, ash, fibre, amino acids, carbohydrates, calcium, phosphorus, zinc, iron, copper and manganese [3]. Also the plant contains substantional amounts of antioxidants including vitamin E, vitamin C, thiamin, riboflavin and niacin [4,5,6].

Lupin hypoglycemic activity was described by (Trider, 1969)[7] which proposed it as a substitute for insulin in mild to medium diabetes mellitus. The hypoglycemic effect of lupin meal was described also in [8] which suggest the used of plums jam containing lupin meal for use as diabetic food for diabetics. Also lupins powdered seed in the form of cataplasma is used to treat ulcers and the seed decoction reduces blood pressure. Lupins is diuretic, seeds are used in veterinary medicine in treating skin disease [9]. When lupin seed was steeped in water for a day then biled, and added to the diet of rats , slightly superior growth [3]. Lupin seeds extracts by using phenolic compounds have antibacterial properties so that [10] suggest using lupin to inhibit bacteria growth. Its seeds used both for alimentary purposes (duo to their remarkable protein contents) and anti parasitic agents [2]. Lupins are thought to be a good substitute for soya beans due to its protein content, for incroporating into stock feed [11]. The main potential market for white lupin grain clearly lies in ruminat feed where its high oil and protein contents are of great value because of the absence of anti-nutritional factor [12].

The aim of the present study is to relates the used of lupin seed oil for the preparation of medicament, food supplements or foods for treatment of diabetes.

Tools and Methods:

Oil extraction:

Lupin seed oil which prepared using 750 g of lupin powder added to 2 L of petroleum ether 60-80 °C in soxhlet apparatus for 12 h period at 40°, the solvent was evaporated using rotatory evaporator to obtain the oil [13].

Oil identification by using thin layer chromatography (TLC):

The oil of lupin identificated by using plate of TLC (supplied by Merck) into groups according to their polarity and examples from each group are included as representative standards [14].

Copper determination in oil:

Copper was determined in oil the lupin using atomic absorption spectrophotometer [15].

Animals used:

Fourty male albino mice (1.5-2) months old weighting (25-30) g each were used. They were housed under standard conditions of temperature 27 °C, humidity, and had free access to food and water. Animals were divided into eight groups (5 mice of each group). The mice were administered orally with oil at increasing concentrations (50,100,200) mg/Kg body weight of normal fated mice. The control group was administered physiological salin solution only. The blood samples were collected from orbital sinus puncture[16] of an ether anesthesia mice 3h of administration. Then the tissues (Livers and Hearts) of these mice were rapidly removed after scarification and washed with ice-cold saline and then kepted in the deep freeze until the glutathione concentration(GSH) and glycogen content were determined in livers and malondialdehyde (MDA) concentration was determined in hearts. The serum was separated by centrifugation at 4000g. Then , blood serum glucose level was determined using Randox kit for glucose oxidase methods [17]. Also the enzymatic method was applied to determine serum cholesterol level using PAP kit [18].

The glycogen content in liver was determined by colorimetric method [14]. MDA in tissues was determined depending on the reaction between MDA and thiobarbutieric acid [19]. Hepatic glutathione were determined using modified Ellman method [20].

The induction of diabetes with alloxan was performed by subcutaneous injection of alloxan monohydrate, which was immediately dissolved in 0.02 ml sterile physiological saline solution at a dose of 150 mg/Kg body weight. The diabetic state was monitored by tested the glucosurea using tes-tape, Eli lilly and company,USA [16] . Then the resulting diabetic animals were grouped into groups (5,6,7,8) after fasting for a period of 16 h, then (6,7,8) groups were administered orally with an oil at 50,100 and 200 mg/Kg body weight. After 3h of administration blood collection and tissues were taken

and performed as indicated previously with fasted normal mice. All assays were done in duplicate.

Statistical :

Significance of differences between mean values was analyzed by Duncan (uses harmonic mean sample size=3.00 and "F" test) [21].

The statistical methods used to analyze the data include mean, standard deviation and maximum, while (F) test was used to compare between groups at ($p \le 0.05$).

Results and Discussion

TLC technique:

The fresh dried plant "Lupins Albus" was used for phytochemical analysis by using plate of TLC to inditificated oil of lupin from mixture of three different solvents (petroleum ether B.p: 60-80 °C): diethyl ether : and glacial acetic acid (80:20:1) ratio as a mobile phase[14].

	Carbon chain length	Compound	Rf value standard sample	Rf value of Lupin oil		
	C12	Lauric	0.69	0.0		
Staurated fatty	C14	Myristic	0.32	0.0		
Staurated fatty acid	C16	Palmitic	0.51	0.51		
aciu	C18	Stearic	0.65	0.0		
	C20	Arachidic	0.87	0.87		
Unsaturated fatty	C18:1	Oleic	0.47	0.47		
acid	C18:2	Linoleic	0.53	0.55		
Phospholipid		Lecithin phosphotidylecholin	0.73	0.72		
Vit A		Vit A	0.89	0.89		
Waxs		Muricul polmitodo	0.31	0.32		
Honey waxs		Myricyl palmitede	0.31			
Sterols		Cholesterol	0.64	0.64		

Table (1): (Rf) values of lupin oils comparing with standard sample of lipids and fatty acids

Detection of oils on TLC plates was normally carried out by spraying the plate with 2,7 dichloroflurescein.

In order to find the type of fatty acids and lipids. Indentification was based on comparing the relative retention (Rf) of lupin oils with relative retention (Rf) of standard sample analysed under the same experimental conditions as shown in table (1). These results were with agreement with other researchers [22] which conclude that the fatty acid composition of white lupin was useful for human consumption , the results indicated that oleic and arachidic acids in seed oil were very high in lupin [23].

On the other hand, Hansen and Czochanska and were found that the composition of lupin seed oil contains triglycerides, phospholipids, free sterols, glycolipids, sterol and wax esters while they were found that the main fatty acids in the total oil extract were linoleic, oleic and palmitic [24].

Effect of lupin seed oil on the some biochemical parameters:

The mean values of serum (Glucose and cholesterol) and liver [GSH concentration and glycogen content] and malondialdehyde (MDA) concentration in heart for control and lupin seed oil –treated normal and alloxaninduced diabetic mice were shown in Table (2).

The result indicated that the dose of oil which was response the experiment is 100 mg/Kg body weight and

there is a significant decrease of serum glucose levels in untreated and treated with alloxan diabetic mice the percentage of decrement was (-25.51%) and (-29.85%) respectively compared with control value as show in table (2) .This decrease may be due to an increase peripheral glucose utilization which increases the release of insulin and /or inhibition of the proximal tublar reabsorption meachanism for glucose in the kidney [25]. The result of the present study show that the lupin seed oil may be possess active constituents capable of lowering blood glucose in both diabetic and non diabetic animals, this effect of lupin oil may be due to it's ability to increase glycolysis or decrease glyconeogenesis or may then cause inhibition of glycogenolysis in the liver. These results were in agreement with other studies [26,7] which they suggest the use of lupin as therapeutical agent , in particular as hypoglycemic agent. Also lupin seed oil content of Cu at concentration (0.4 μ g/g) which was simlar to [3]. The presence of Cu at this concentration plays an important role in insulin synthesis [27] which lead to decrease in serum glucose level.

Also the results in Table (2) indicated that the lupin seed oil at a dose of 100 mg/Kg body weight decreases the level of cholesterol. The percentage of decrement was (-22.39%) relative to control and (-23.09%) relative to diabetic control. This result was similar to [28], this decrease may be due to inactivation of the hydroxy methyl glutaryl CoA reductase (HMG –CoA reductase), a thiol group enzyme that is necessary for cholesterol synthesis [16,29].

In the present study, the mice treated with oil of 100mg /Kg body weight showed a significant decrease in glycogen content in normal mice of (-23.29%) after 180 min of administration of oil relative to control group, as

shown in table (2), these results are similar to that results obtained by other [28] which observed that lupin diets increased plasma glucagons hormone levels, this hormone has an opposite effect for that of insulin.

On the other hand, lupin seed oil has non significant difference on the concentration of liver glutathione and heart malondialdehyde relative control value.

Table (2): Mean blood serum (glucose and cholesterol) level in mg/100 ml ± SE and liver (glycogen mg/g and				
GSH nomle/g) and heart MDA nmole/g after oral administration of different doses of lupin seed oils in normal				

Grou p No.	Treatment	Serum glucose level mg/100m l	Change %	Serum cholester ol level mg/100m l	Change %	liver Glycogen mg/g	Change %	liver GSH nmole/g	Chang e %	MDA heart nmole/g	Chang e %
1	Normal	96.73		162.1		2.82		4276.2		281.35	
	saline	±		±		±		± .		± _	
	contral	1.47 de		1.9 d		1.9 f		6.1 e		6.3 c	
2 Seed oil 5	Seed oil 50	83		144.4		2.69		4302.5		265.47	
	mg	±	-10.78	±	-10.92	±	-12.18	±	4.82	±	-4.1
	mg	2.13 c		2.7 c		0.01 b		12.7 e		3.5 ba	
3	Seed oil	65		125.8		1.93		4319.8		241.92	
	100 mg	±	-25.51	±	-22.39	±	-23.29	±	6.41	-	-9.3
	Too Ing	3 a		3.4 a		0.04 a		5 ef		5.5 a	
4	Seed oil	90		134.73		2.71		4278.86		253.42	
	200 mg	±	-4.6 ±		-16.88	±	-10.28	±	2.01	±	-7.21
	200 mg	2.8 f		2.36 b		0.02 e		3.2 f		3.4 b	
5	Diabetic	185.13		183.53		3.11		2890.8		456.77	
	control	±		±		±		±		±	
C	control	2.15 g		1.79 e		0.03 d		24.4 a		2.59 g	
б т	Diabetic	154.06		147.76		2.91		2873.3		439.89	
		±	-13.78	±	-14.35	±	-8.84	±	-11.31	±	-8.3
p	plus 50 mg	1.47 d		2.89 c		0.05 df		4 a		4.5 f	
7	Diabetic	129.86		132.7		2.88		2863.2		446.34	
	plus 100	±	-29.85	±	-23.09		-10.7	±	-15.4	±	-5.11
	mg	3.29 b		1.7 ab		۱e		4 ab		2.6 d	
8	Diabetic	158.6 ±		143.86		2.85		2884		452.2	
	plus 200	0.66 de	-11.63	±	-16.62	±	-13.09	±	-8.2	±	-2.0
	mg			3.7 c		0.04 fd		7.6 b		3.7 db	

• vertically different letter meaning that there is a significant difference at ($p \le 0.05$).

• The values are mean ±SE of 4 mice each group.

References

1.Clapham,A.R.,Tutin,T.G.and Moore, D.M.,(1987). Flora of british Isles. 3rd ed. Cambridge University press. :840-841.

2.Gill,N.T.andVear,K.C.,(1980).Agricultural botany. 3rd ed . London: Gerald Duckworth and Co. Ltd.: 1321.

3. Hove ,E.L.,(1974). Composition and protein quality of sweet lupin seed. 25(7):851-859.

4. Szczapa, E.L., Korczak, J., Kalucka, M. and Wojtasiak R.,(2003). Antioxidant properties of lupin seed products. J. Food Chemistry. 83(2):279-285.

5. Erbas, M., Certel, M. and slu, M.K.(2005).Some chemical properties of white lupin seeds (Lupinus Albus L.). J. Food Chemistry. 89(3): 341-345.

6. Frias, J., Miranda, M.L., Doblado, R. and Valverde, C.V. (2005). Effect of germination and fermentation on the antioxidant contents and antioxidant capacity of lupinus albus (L.var) Multolupa. J. Food Chemistry ,92(2):211-220.

7. Morazzoni ,Paolo, Duranti and arcello.(2008). Use of lupin conglutin for the treatment of type II diabetes <u>http://www.freepatentsonline.com/7323441.html</u>

8. Mario, V.(1996). Archives latinoamericanos de nutricion. 46 (3):.234-237.

9. Kotob F.T.,(1985). Medical plant in libya .Arab. Encyclopedia houses: 566.

10. Szczapa, E.L., Siger, A., Trojano Wska, K., Kalucka, M., Malecka, M. and Pacnolek, B.(2003). Chemical composition and antibacterial activities of lupin seeds extracts ,Wiley Inter Science Journal, 47 (5):286-290.

11. Sells, J.E.(1989).Combinable alternative crops Bedford: AFRC Institute of Engineering.

12. Milford, G.F.J. and Shield, I.F.,(1996). The potential of lupins for UK. Agriculture .J. of the Roxal Agricultural Society of England.:157.

13. Harbone, J.B., (1973).Phytochemical methods .1st. ed.Printed in Great Bitain by Cox and Wyman Ltd., London, pp. 92-95.

14.Plummer, D., (1978).An Introduction practical biochemistry 2nd ed., Mc Graw-Hill Book Co.:345-346.

15.Thomas, L.C. BSc. F.R.I.C.,G.J. CHIMBERLIN. (1980).Colorimetric chemical analytical, 9th ed., England. 16. Abd- Alrahman S.Y.(1995). Effect of starvation and experimental diabetes mellitus on tissue levels of glutathione and lipid peroxidation. Ph. D. Thesis, College of Veterinary Medicine, University of Mosul(In Arabic). 17. Trider, P.,(1969).Ann.Clin.Biolchem.,6:.24-27.

18. Richmod, W., (1973).Clin Chem., 19:1350-1356.

19. Volkan ,E., Nurperi, G. and Ahmet, B.(2001).N-acetyl cystine reduces cerebral lipid peroxidation in a rat

model of infantile hydrocephalus. J. Neurol. Sci., Issue :1302-1310.

20. James, R.C., Goodman, D.R. and Harbison, R.D., (1982). Hepatic glutathione and hepatoxicity, changes inducedloy selected marcortics. J.Pharmaco. Therap, 221: 708-714.

21. Steel and Torrie, J.(1960). Principles and procedures of statistics, Mc -Graw-Hill Book Co. :99-131.

22. Boschin ,G. Agostina, A.D. and Paolo (2008).Effect of genotype and environment on fatty acid composition of lupinus albus L.Seed Food Chemistry .18(2):600-606.

23. Uzun, B., Arslan, C., Karhan, M. and Toker, C.,(2007). Fat and fatty acids of with lupin (Lupinus albus L.) in comparison to sesame (sesamum indicum L.) Food Chemistry,102(1):45-49.

24. Hansen ,R.P. and Czochanska.(1974). Comosition of lipids of lupin seed (Lupinus angustiflius L. Var. "uniwhite") .J. of the Science of Food Agriculture ,25 (4):409-415.

25. Chattopadhyay, R.(1999).A comparative evaluation of some blood sugar lowering agents of plant origin. Ethnopharmacol, J. 3:373-376.

26. Mansour, H.A., Newairy ,A.A, Yosef .M.I. and Sheweita, S.A.(2002). Biochemical study on the effects of some Egyptian herbs in alloxan-induced diabetic rats Journal Toxicology.170(3):221-228.

27. McCarty, M.F.(1999). High dose biotin, an inducer of glocokinase expression and therapy of type II diabetes. Med. Hypotheisi. 52: 401-406.

28.chango, A. Villaume, C., Bau, H.M., Schwertz, A., Nicolas ,J. p. and Mejean, L.(1997).Effects casein, sweet white lupin and sweet yellow lupin diet on cholesterol metabolism in rats. Journal of the Science of food and agriculture.76(2):303-309.

29. Arivazhagan, S., Balasenthil, S. and Nagini, S., (2000). Modulatory effects of garlic and neem leaf extracts on N- methyl l- N- nitro - N - nitrosoguanidine (MNNG) -induced oxidative stress in wistar rats. Cell. Biochemi. Functions., 18 (1):17-21.

بعض التأثيرات الكيموحيوية لزيت ثمار الترمس على الفئران السليمة والمصابة بداء السكر فاطمة عبد الحميد محمد

قسم الكيمياء ، كلية العلوم ، جامعة الموصل ، الموصل ، العراق (تاريخ الاستلام: ١٠ / ٦ / ٢٠٠٨ ، تاريخ القبول: ١٥ / ١٢ / ٢٠٠٨)

الملخص:

تم في هذا البحث استخلاص زيت ثمار الترمس ومن ثم شخصت محتوياته من الأحماض الدهنيه والدهون مقارنة مع احماض دهنيه ودهون قياسية باستخدام تقنية كروماتوغرافيا الطبقة الرقيقة .

كما تناول البحث تأثير تراكيز متصاعدة لزيت ثمار الترمس على بعض المتغيرات البابوكيميائية في مصل وأنسجة الفئران السليمة والمصابة بداء السكري التجريبي المستحدث بالالوكسان، اذ ادى التجريع عبر الفم بتراكيز (٥٠، ١٠٠، ١٠) ملغم/كغم من وزن الجسم من زيت ثمار الترمس الى انخفاض معنوي في مستوى كلوكوز مصل الفئران السليمة والمصابة بداء السكري المستحدث بالالوكسان ال ومن ثم تم استخدام تقنية الامتصاص الذري اللهبي لايجاد تركيز النحاس في بواقع (– ٢٥,٥٠) و (– ٢٩,٨٥) على التوالي ومن ثم تم استخدام تقنية الامتصاص الذري اللهبي لايجاد تركيز النحاس في ريت ثمار الترمس كلى المستحدث بالالوكسان ورن الجسم من زيت ثمار الترمس الى انخفاض معنوي في مستوى كلوكوز مصل الفئران السليمة والمصابة بداء السكري المستحدث بالالوكسان بواقع (– ٢٥,٥٠) و (– ٢٩,٨٥) على التوالي ومن ثم تم استخدام تقنية الامتصاص الذري اللهبي لايجاد تركيز النحاس في زيت ثمار الترمس.كما لوحظ ان هنالك انخفاض معنوي في مستوى الكوليسترول في مصل الفئران السليمة والمصابة بداء السكر ريت ثمار الترمس.كما لوحظ ان هنالك انخفاض معنوي في مستوى الكوليسترول في مصل الفئران السليمة والمصابة بداء السكر ريت ثمار الترمس.كما لوحظ ان هنالك انخفاض معنوي في مستوى الكوليسترول في مصل الفئران السليمة والمصابة بداء السكر ريت ثمار الترمس.كما لوحظ ان هناك انخفاض معنوي في مستوى الكوليسترول في مصل الفئران السليمة والمصابة بداء السكر (- ٣٠,٠٩٠) و (- ٣٠,٣٠) و التجريبي المستحدث بالالوكسان بواقع (– ٣٠,٣٠) على التوالي ، وكما يشير البحث الى انخفاض معنوي في كلايكوجين الكبد في الفئران السليمة بواقع (– ٣٠,٠٠) مار الترمين معان معنوي في كلايكوجين الكبد في الفئران السليمة بواقع (– ٣٠,٠٠) بعد مضي ١٠ دقيقة من التجريع مقارنة بالقيم الضابطة.

من ناحية اخرى فان ليس لزيت ثمار الترمس تاثير على تراكيز كلوتاثايون الكبد ومالوتالديهايد القلب مقارنة بالقيم الضابطة..

الكلمات الدالة : الترمس ، السكري ، الكوليسترول ، الكلوتاثايون.