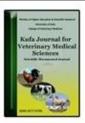
Kufa Journal for Veterinary Medical Sciences Vol. 5 No.1 (2014) 10-15



Kufa Journal for Veterinary Medical Sciences

www.vet.kufauniv.com



Rate of induction of resistance in Fecal *E-coli* against cefquinome as compare to ceftriaxone after continuous passage in *vivo*

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Abstract:

The study was conducted to evaluate the development of resistance for fecal E. coli against third and fourth generations of cephalosporins. This bacteria was chosen because they represent the pool of resistance elements that are available for transfer to other bacterial species including pathogens. Three groups of 5 (mice) were used. 89.25µg/Kg) of ceftriaxon and(12.5µg/Kg) of cefquinome, were given a sub therapeutic dose orally for the first and second groups respectively. While the third group were given distilled water only as a control. Then isolation, purification and identification of fecal E-coli from GIT was done. Morphological and biochemical tests had been used to make sure that isolated bacteria was E.coli. It has been observed that the mean value minimum inhibitory concentration (MIC) of the isolated bacteria from both of first and second groups was compared with the control group.MIC of both antibacterials in comparison with the control group which were 1.18, 1.37µg/ml for ceftriaxone ,cefquinome respectively. While the MIC values for ceftriaxone and cefquinome were 16.00 and $4.6~\mu g/ml$ respectively .this means that the significancy (P<0.05) was 13.55 folds and 3.35 folds in the third and fourth generation generations respectively in comparison with the control of each antibacterial .We concluded that antibiotic resistance may not be only a consequence in pathogenic bacteria but also in normal flora which could contribute this resistance to other microorganism.

Key word: MIC: Minimum Inhibition Concentration, PBPs: penicillin-binding proteins

معدل استحداث المقاومة في الاشيريشيا القولونية البرازيه ضد السفكوينوم مقارنة مع السفترياكزون بعد تكرار تمريرها داخل الجسم الحي

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الخلاصة:

أجريت هذه الدراسة لتقييم تطوير المقاومة في الاشيريشيا القولونية البرازيه ، في جسم الكائن الحي ضد الجيل الثالث والرابع من السيفالوسبورين الشائعة الاستعمال في الإنسان والحيوان ، وقد اختيرت هذه البكتيريا لكونها تمثل محيط من

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عناصر المقاومة والتي تكون متاحة إلى أنواع أخرى من البكتيريا وبضمنها المرضية. أخذت في البداية ثلاث مجاميع (فئران) كل مجموعة تحتوى على خمسه أفراد أعطيت (89.25 مكغم/مل) من السفترياكزون و (12.5 مكغم/مل) من سُفكوينوم كربع الجرعه العلاجية مره واحده يوميا ولمده أسبوع ، للمجموعة اللاولى والثانيه على التوالي ، وتم اخذ المجموعة الثالثة كمجموعة سيطرة وإعطائها ماء مقطر فقط ولنفس المدة المذكورة أعلاه ، وبعد انتهاء الفترة المحددة تم عزل الاشيرشيا القولونية البرازيه وبعدها تم إعادة تأكيد هوية الجرثومة وبعدها تم تحديد قيمه التركيز المثبط الأدنى MIC للبكتيريا لكل مجموعة (الأولى والثانية) ومقارنتها مع مجموعة السيطرة. وكانت القيم MIC للمضادات الحيوية هي لمجموعة السيطرة هي 1.18 مكغم/مل للسفترياكزون و 1.37 مكغم/مل للسفكوينوم وبنفس الطريقة تم تحديد قيم MIC للمجموعتين الاولى والثانية بعد استحداث المقاومة لهما وكانت قيم MIC ،16.00 MIC مكغم/مل للسفترياكزون والسيفكوينوم على التوالي وهذا يعني بأن الاهميه ضمن المستوى (P<0.05) كانت 13.55 ضعفا و 3.35 ضعفا للجيل الثالثُ والرابع على التوالي بالمقارنة مع مجموعة السيطرة لكلُّ مضاد حيوى ولقد استنتج من هذه الدراسة أن مقاومة المضادات الحيوية قد لا تكون فقط للبكتيريا المسببة للأمراض ولكن أيضا تحصل للفلورا الطبيعية التي يمكن أن تسهم في نفل هذه المقاومة إلى الكائنات الحية الدقيقة الأخرى.

Introduction:

The increase in the number of resistant and multiresistant strains of bacteria is a concern of health worldwide, particularly with the decline in the number of new antibiotics available for treatment. While much effort has been toward management directed and monitoring of antibiotic use and the prevalence of bacterial within communities (1,2). Furthermore, antimicrobial-resistant is spreading between different bacterial strains in different habitats has been demonstrated (3).

The normal flora contains antibiotic resistance genes to various degrees, even in individuals with no history of exposure to commercially prepared antibiotics. Several factors seem to increase the number of antibiotic-resistant bacteria in feces. One important factor is the exposure of the intestinal flora to antibacterial drugs. Antibiotics used as feed additives seem to play an important role in the development of antibiotic resistance in normal flora bacteria.. The knowledge **(4)**. prevalence and patterns of antimicrobial resistance in generic Escherichia coli might indicate the pool of resistance elements that are available for transfer to bacterial species other including pathogens(5). Generic E. coli are used to monitor changes in prevalence and patterns of resistance as these commensal bacteria are regularly found in the gastrointestinal tract of animals and humans (6).

Ceftriaxone third generation a cephalosporins P, N and C, 7-amino cephalosporinic acid, and with the addition of side chain, it became possible to semisynthetic produce compound, Ceftriaxone injectable is cephalosporin addition agent,in .Ceftriaxone are highly resistant to staphylococcal β- lactamases producers and has a good activity against Gram positive and Gram negative bacteria (7) .it is considered to be the drugs of choice for many infections caused by members of the Enterobacteraciae (8). Ceftriaxone, a betalactam antibiotic is mainly bactericidal. It inhibits the third and final stage of bacterial cell wall synthesis by binding preferentially specific to penicillin-binding proteins (PBPs) that are located inside the bacterial cell wall.

Cefquinome aminothiazolyl is an cephalosporin extended spectrum betalactam and a member of the fourth generation cephalosporins, Cefquinome has been used only in veterinary medicine and only for individual treatment, and is safe and well tolerated. Formulations for the US will be available for parenteral injection, either as multiple or as singledose products.(9). it is a zwitter-ionic compound with improved penetration into the periplasmatic space of Gram-negative bacilli and enhanced binding to penicillinbinding proteins .Aim of our study that is isolation and identification of fecal E-coli from mice and determine the sensitivity by measuring the (MIC) against cefquinome and ceftriaxone and determination possibility of induction of resistance of these microorganisms against the above mentioned drugs in *vivo* by continuous treatment with sub therapeutic doses(1/4) in mice.

Materials and Methods:

To determine the rate of antibiotic resistance of *fecal E. coli* from healthy mice and to infer it is induced by antibiotic use by giving orally sub therapeutic dose, In the present studies white Swiss BALB/C mice were employed as the test animals,15 mice were taken and divided three groups respectively as following:

- 1- group one, 5 mice were given a sub therapeutic dose orally $(89.25\mu g/Kg)$ of ceftriaxon, once daily for one week.
- **2-** Group two 5 mice were given sub therapeutic dose orally (12.5µg/Kg)for cefquinome, giving once daily for one week (10)
- 3- Group three served are control five mice were given distilled water only for the same period. At the end of 7 days isolation. purification period. identification of fecal E-coli from GIT was done by Fresh specimens of faeces (0.5 g) from 6 normal healthy mice collected in sterile universal containers. They were transported quickly to the laboratory and processed within an hour of collection, cultured first on the brain heart infusion broth, incubated at 37°C, for 24hrs, transfer to macConkey agar secondly, and transfer to E.M.B agar. The diagnosis depends on the color, shape, size of colonies, also the confirmatory tests had been completed to make sure that isolates bacteria was E.coli after that determined the mean MIC to bacteria for each group (first and second) which compare with control group.

Results and Discussion: Induction resistance in *vivo* by giving antibiotic orally at sub therapeutic dose(1/4) for one week.

An interesting observation in this study is the induction of resistance to third and fourth generation cephalosporins in vivo. In an experiment designed to induce resistance in vivo for fecal E. coli to cetriaxone and cefquinome, after exposure microorganisms to sub of susceptible therapeutic dose (1/4) of each of the two drugs for seven days .Then isolation, purification and identification of fecal Ecoli from GIT was done. Morphological and biochemical tests had been used to make sure that isolated bacteria was E.coli., The results of MIC of the two control group (after antibacterials for giving distilled water only) estimated by tube dilution method are summarized in table 1. These were 1.18, 1.37µg/ml for ceftriaxone, cefquinome respectively. This no significant results showed that difference in value of MIC with each drug because that third and fourth generation have been found to be effective against most major pathogens, including E. coli.

The same process after giving drugs orally (sub therapeutic dose) for induction of resistance the MIC values were set out in tables 2,3 and 4 they were 16.00 and 4.6 ug/ml for ceftriaxone and cefquinome respectively, which mean an increase of 13.55, 3.35 folds respectively. The third generation showed significantly highest increase (13.55) folds, while the fourth showed the lowest increase generation folds.These findings (3.35)agreement with (11) found reduction in the proportion of sensitive coliforms excreted in pigs after chlortetracycline fed at concentrations of 10 or 20 grams per ton. (12), found in their study when used clindamycin, in short-term studies, cause disturbances in the composition of the gut microbiota as well as to select for resistance. These studies have mainly been based on data from isolates and have indicated a normalization of the flora a few weeks following withdrawal of the treatment However, by using molecular focused on the approaches Bacteroides, it is found longterm shifts in composition of the intestinal microbiota of individual subjects after a short-term administration of clindamycin (13). As mentioned before exposure of microorganism to different levels of antibacterial drug may result in increase in degree of resistance as reported before by many workers. (14) reported the use of broad spectrum antibiotics creates selective pressure on the bacterial flora, thus increasing the emergence multiresistant bacteria, which results in a vicious circle of treatments and emergence of new antibiotic resistant bacteria. The

gastrointestinal tract is a massive reservoir of bacteria with a potential for both and transferring receiving antibiotic resistance genes, in the context that the reason for the development of resistance in vivo is due to replacement of sensitive serotypes by other already resistant serotypes.

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At the same time, it is possible to monitor the acquisition and persistence of resistance genes in the community. Together this information should help to provide knowledge of the natural dynamics of the normal microbiota and help us to understand the long-term consequences of antimicrobial treatment. This information great importance implementation of rational administration guidelines for antibiotic therapies.

Table 1: the result value of MIC of two antibacterials after giving distal water only for one week

No of mice	Value of MIC µg/ml for ceftriaxone	Value of MIC µg/ml for cefquinome	
Mice 1	1.25	1.25	
Mice 2	2.5	2.5	
Mice3	1.25	1.25	
Mice4	0.31	0.62	
Mice5	0.62	1.25	
* Mean ± stander error	1.18 ± 0.37	1.37 ± 0.30	

^{*}Number represent mean \pm stander error P>0.05

Table 2. show initial and final values of MIC of the ceftriaxone, after giving orally, contain sub therapeutic dose (1/4 therapeutic dose) for one week.

No of mice	Initial MIC µg/ml (giving D.W only)	final MIC µg/ml (giving sub therapeutic dose)	
Mice 1	1.25	20.00	
Mice 2	2.5	10.00	
Mice3	1.25	20.00	
Mice4	0.31	20.00	
Mice5	0.62	10.00	
* Mean ± stander error	1.18 ± 0.37	16.00 ± 2.44 a	

^{*}Number represent mean ± stander error

Different small letters mean significant (P<0.05) result between concentration within group.

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Table 3. show initial and final values of MIC of the cefquinome, after giving orally, contain sub therapeutic dose (1/4 therapeutic dose) for one week.

No of mice	Initial MIC μg/ml (giving D.W only)	final MIC µg/ml (giving sub therapeutic dose)	
Mice 1	1.25	3.12	
Mice 2	2.5	6.25	
Mice3	1.25	1.56	
Mice4	0.62	6.25	
Mice5	1.25	6.25	
* Mean ± stander error	1.37 ± 0.30 b	4.6 ± 0.98 a	

^{*}Number represent mean ± stander error

Different small letters mean significant (P<0.05) result between concentration within group.

Table .(4-13) show values of MIC of the control group and two antibiotics in vivo ,after giving sub therapeutic dose for one week.

ANTIBIOTICS	Value of MIC µg/ml of control groups (giving D.W only)	Value of MIC after giving sub therapeutic dose for one week	Folds of elevation
Ceftriaxone	1.18 ± 0.37 b	16.00 ± 2.44 Aa	13.55
Cefquinome	1.37± 0.30 b	4.60 ± 0.98 Bb	3.35

Numer represent mean \pm stander error.

Different small letters mean significant (P<0.05) result between within group. **Different** capital letters mean significant (P<0.05)result betwee different groups. L.S.D: 2.9.

References:

- Costanzo, S.D.; Murby, J. and 1. Bates, J. (2005). Ecosystem response to antibiotics entering the aquatic environment, Marine Pollution Bulletin. 51: 218-223.
- Batt, A.L.; Bruce, I.B. and Aga, D.S., (2006). Evaluating the vulnerability of surface waters to antibiotic contamination from varying waste water

treatment plant discharges. Environmental Pollution. 142: 295-302.

- Svarm, (2006). Swedish veterinary 3. antimicrobial resistance monitoring. The Institute (SVA), National Veterinary Uppsala, Sweden.: 23-24.
- Henning, S. Marianne. 4. and S.(2001) . Review article of the Resistance

- to antibiotics in the normal flora of animals. Vet. Res. 32, : 227–241.
- Sunde, M. and Sorum, H. (2001). Self-transmissible multi-drug resistance plasmids in Escherichia coli of the normal intestinal flora of healthy swine. *Microb*. Drug Resist. 7 (2): 191-196.
- Kim, S.; Hu, J.; Gautom, R. Kim, 6. J.; Lee, B. and Boyle, DS.(2007). CTX-M spectrum extendedbeta-lactamases, Washington State. Emerg Infect Dis. 13:513-514.
- Neu, H. C.; Aswapokee, N.; 7. Aswapokee, P. and Fu, K.P. (1979). HR576, a new cephalosporins active against Gram positive and Gram negative aerobic and anaerobic bacteira. Antimicrob. Agents Chemother. 15: 273-281.
- 8. Luke, C. H. (2011). Studies on Salmonella enterica and Escherichia coli with a focus on ceftiofur and the genetic resistance determinant blaCMY-2. PhD Dissertation. Graduate Program in Veterinary Preventive Medicine Ohio State University, US.
- 9. FDA/CVM(Food and Drug Administration Center for Veterinary Medicine) Guidance. (2006). Cefquinome formulations for parenteral injection for the treatment of bovine respiratory disease, :1-50.

Michael, L.; Dieter, I; Norbert, 10. K.; Astrid, M.; Karl, S.; Gerhard, S. and Elmar, S.(1991). Antibacterial Activities In Vitro and In Vivo and Pharmacokinetics of Cefquinome (HR IIIV), a New Broad-Spectrum Cephalosporin. Antimicrobial agents and Chemotherapy, Vol. 35, No. 1; : 14-19.

No. (1)

- 11. Finlayson ,M. and Barnum, D. A.(1973). The Effect of Chlortetracycline Feed Additive on the Antibiotic Resistance of Fecal Coliforms of Weaned Pigs Subjected to Experimental Salmonella Infection. Can. J. comp. Med. Vol. 37:63-69.
- 12. Sullivan, A.; Edlund, C. and Nord, C. E. (2001). Effect of antimicrobial agents on the ecological balance of human microflora. Lancet Infect Dis 1, 101–114.
- C.; Lo fmark, 13. Jernberg, Edlund, C. and Jansson, J. K. (2007). Longtermecological impacts of antibiotic administration on the human intestinal microbiota. ISME J 1, 56-66.
- S. 14. Susanne. and Karen. K.(2011). Assessment of **Bacterial** Antibiotic Resistance Transfer in the Gut. Review Article, International Journal of Microbiology; Volume 2011, Article ID 312956:1-10.