

Effect of Garlic Oil on Gentamicin Induced Hepatorenal Toxicity in rats

اثر زيت الثوم على السموم الكبدية الكلوية المستحثة بأعطاء دواء الجنتاميسين على الجرذان

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Abstract:

Gentamicin is an important aminoglycoside, natural or semisynthetic antibiotic commonly used against the life threatening infections in human. Nephrotoxicity is the major side effect of aminoglycosides. The aim of this study is to investigate the protective/ prophylactic roles of garlic oil against gentamicin-induced abnormalities in metabolic biochemical parameters in serum and tissues of female albino rats. In this study, we used 30 Wistar albino 230-250 g female rats. The animals were randomly divided into five groups. Each experimental group consisted of six animals. Control group, they were fed with only standard rat diet and tap water for 2 week. Garlic Oil: Rats were treated daily with a dose of 5 ml/kg Garlic Oil (GO) via gavage, and received was start of the 5 day of the experiment. Gentamicin: Rats were treated daily with a dose of 100 mg/kg gentamicin (G) as intraperitoneal (IP) at 5 day of the experiment. Gentamicin + Garlic Oil: Rats were treated with synchronised intraperitoneal gentamicin plus garlic oil 5 ml/kg every day via gavage at 5 day of the experiment. Garlic Oil (Pre) + Gentamicin: Rats were treated with 5 ml/kg/ every day via gavage garlic oil in start the first day of the experiment and then gentamicin received at the above-mentioned dose at 5 day of the experiment. A significant increase in serum levels of glucose, bilirubin, urea and creatinine as well as the activity of the AST, ALT, LDH, and ALP enzymes were observed in rats treated with gentamicin for a period of ten days. However, supplementation of gentamicin-intoxicated rats with garlic oil ameliorated the gentamicin adverse effects as evidenced by a significant increase of serum total protein content, and a decrease of serum glucose, bilirubin, urea and creatinine levels, as well as the activity of AST, ALT, LDH and ALP enzymes. Histopathological changes in liver and kidney were not observed on animals treated with Garlic oil when compared with the control group. However, animals treated with gentamicin showed fibrosis, necrosis, and fatty infiltrate in liver and showed necrosis and degeneration of glomerulus in kidney. This effect was significantly decreased in animals pretreated with garlic oil.

Keywords: Gentamicin, Hepatorenal toxicity, Garlic oil

المخلص :

الجنتاميسين هو عبارة عن مضاد حيوي شائع من عائلة الكلايكوسايد يستخدم ضد الامراض المعدية التي تهدد حياة الانسان طبيعته الكيميائية عبارة عن امينو كلايكوسيد وتعتبر هذه المجموعة الدوائية ذات اعراض سمية على الكلى و الكبد اذا اعطيت بجرعات متواصلة او عالية. الهدف من الدراسة تقييم كفاءة زيت الثوم الذي يعتبر ذو خصائص مضادة للاكسدة لتقليل سمية الجنتاميسين على الكلى و الكبد و معرفة الدور الوقائي و العلاجي لهذا الزيت. اجريت الدراسة على ثلاثون جرذا وتم تقسيمها الى خمسة مجاميع كل مجموعة تضمنت ستة جرذان. اعتمدت مجموعة الضبط على الغذاء المعياري و الماء و لم يتم اعطائها اي مادة علاجية , المجموعة الثانية تم اعطائها زيت الثوم و بجرعة مقدارها 5 ملم \ كغم \ يوم عن طريق الفم , بالنسبة للمجموعة الثالثة تم حقنها بمادة الجنتاميسين و بجرعة 100 ملغ \ كغم , المجموعة الرابعة تم اعطائها زيت الثوم بعده حقنها بالجنتاميسين اما المجموعة الخامسة فقد تم اعطائها زيت الثوم ولمدة خمسة ايام قبل حقنها بمادة الجنتاميسين. اسفرت نتائج التجربة بأن اعطاء مادة الجنتاميسين للجرذان و لمدة عشرة ايام تسبب زياده معنوية في مستوى الكلوكرز في الدم ,

البليروبين, اليوريا و الكرياتينين بالإضافة الى مستوى فعالية انزيمات وظائف الكبد بينما لوحظ اعطاء زيت الثوم للجرذان المحقونة بدواء الجنتاميسين ادت الى انخفاض معنوي في القياسات البايوكيميائية اعلاه. كما تم التوصل بأن هنالك تغيرات نسيجية مرضية في كل من النسيج الكبدي و الكلوي في الجرذان المحقونة بمادة الجنتاميسين في حين قلت التغيرات النسيجية المرضية بعد اعطائها زيت الثوم.

مفتاح البحث: الجنتاميسين, السموم الكبدية الكلوية, زيت الثوم

Introduction:

Gentamicin is an important aminoglycoside, natural or semisynthetic antibiotic commonly used against Gram negative bacterial infections. Nephrotoxicity is the major side effect of aminoglycosides, which may occur in 13-30 % of treated patients¹⁻³. Gentamicin is excreted through kidneys without degradation or metabolic changes, with 5 to 10 dose is concentrated in proximal tubules vastly exceeding the concurrent serum concentration⁴. The accumulation of gentamicin in kidney cortex can cause oxidative stress and lipid peroxidation⁵. Several studies have shown that reactive oxygen metabolites including reactive oxygen species like superoxide, hydroxyl radical anion and hydrogen peroxide are important mediators of tissue injury⁶⁻⁹. Gentamicin causes the activation of platelet activation factor ensuing local vasoconstriction and thus reduces the renal blood flow and glomerular filtration rate. Gentamicin induced renal injuries are mostly localized to the proximal tubules because of association of gentamicin with polyanionic inositol phospholipids and megalin, a receptor, for uptake of gentamicin. Gentamicin resists the degradation of phospholipids that compromised the lysosomal membrane integrity and eventually the leakage of enzymes. Gentamicin behaves as iron chelator and iron-gentamicin complexes are thought to be involved in free radical formation¹⁰⁻¹³.

Garlic is a member of the lily family, contains more than 200 chemical compounds. Some of its more important ones include volatile oil with sulphur containing compound (allicin, allin and ajone), and enzymes (allinase, peroxidase and myrosinase)^{14,15}. Ancient Egypt records mentioned that use of garlic as a remedy for a variety of disease¹⁴.

Garlic has antioxidant properties that have been shown to inhibit lipid peroxidation (LPO)¹⁶, and dose-dependent induction of endogenous antioxidants in rat kidney and liver¹⁷. And also have antimutagenesis, xanthine oxidase inhibitor, anticarcinogenesis, anti-inflammatory, antiviral, antifungal, antiatherogenic and antithrombotic effects¹⁸⁻²².

The aim of this study is to investigate the protective/prophylactic roles of garlic oil against gentamicin-induced abnormalities in metabolic biochemical parameters in serum and tissues of female albino rats.

Material and Methods:

Chemicals:

Gentamicin (80 mg/2ml) was obtained from the Essential Drug Company (Baghdad, Iraq), and given by intraperitoneally injection at a dose of 100mg/kg body weight as previously described¹⁰. Garlic oil was purchased from local market (Kerbala, Iraq). Garlic oil was given by gavage at a dose of 5ml/kg as described²³.

Animals :

In this study, we used 30 female Wistar albino (230-250 g) rats which were housed in wire bottom cages, free standard diet, tap water and with a 12 h light/ dark cycle for 2 weeks (the experimental period). The animals were randomly divided into five groups, each experimental group consisted of six animals.

Control group, they were fed with only standard diet and tap water for 2 weeks.

Garlic Oil group, rats were treated with garlic oil with a dose of 5 ml/kg daily via gavage started from the 5th day to last day of the experimental.

Gentamicin group, rats were treated with 100 mg/kg gentamicin daily by intraperitoneal injection started from the 5th day to last day of the experimental.

Gentamicin-Garlic Oil group, rats were treated with gentamicin with a ratio of 100 mg/kg by intraperitoneal injection at the same time garlic oil with ratio of 5ml/kg was given via gavage daily started from the 5th day to last day of the experimental.

Garlic Oil-Gentamicin group, in this group, rats were treated with garlic oil with a ratio of 5 ml/kg daily via gavage started from the first day to the last day of the experiment while gentamicin was given with a ratio of 100 mg/kg by intraperitoneal injection from the 5th day to last day of the experimental.

The end of the experiment rats were sacrificed 24h after the last garlic oil and gentamicin received, and blood samples were collected in centrifuge tubes. Serum was separated from coagulant blood by centrifugation at 860g for 20 min, and then frozen at -20°C for biochemical analysis. One gram of liver or kidney tissue samples were homogenized (Heidolph® homogenizer) with 4 ml 0.9% NaCl at 4000 rpm for 1.5 min and then extract by centrifuge at 2000 rpm for 20 min, then stored at -20°C for subsequent measurements. Liver and kidney were excised, tissue samples were rinsed with 0.9 cold NaCl solutions and stored at -80 °C, and embedded on paraffin for histopathologic research.

Biochemical analysis:

The serum glucose was estimated by glucose kit (Randox®)²⁴. Serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST) activities were estimated according to Reitman and Frankel methods²⁵. Creatinine levels were determined using Diamond Diagnostic kit® according to Owens *et al.* method,²⁶ also urea was estimated by the use Diamond diagnostic kit® as previously reported²⁷. Total protein and bilirubin were determined using Diamond diagnostic kit® as described²⁸. Alkaline phosphatase was determined according to Belfield and Goldberg method²⁹. Lactate dehydrogenase were determined using commercially available diagnostic kit (Biomerieux®) as described²⁸. The cholesterol and triglycerides were estimated by the use Biolabo® diagnostic kit as described^{30,31}.

Preparation of kidney and liver tissues for Histopathology:

The specimens were fixed in formalin 10 % solution for 72 hours. After fixation, the tissues were washed under running tap water for 24 h and dehydrated with 50, 60, 70, 80, 90, 96 and 100% concentrated ethanol. The specimens were then laid in a 1:1 ratio of immersion oil and absolute alcohol for 1h, followed by immersion oil overnight, for transparency. After the application of xylol, the specimens were made into paraffin blocks using a 1:1 xylol and paraffin mixture for 1h and paraffin for 6 hours in an incubator. 10 micron thick sections were rehydrated and dyed with Masson's trichrome (Bio-Optica Masson tricromica cat no 04-010802, Milano S.p.a, via San Faustino, 58, 20134 Milano, ITALIA) technique.

Statistical analysis:

The data was analyzed using the Statistical Package for Social Science program (SPSS 12). For comparison between different experimental rat groups, one way analysis of variance (ANOVA) was used followed by Tukey's test. The results were expressed as means \pm SE and $P < 0.05$ was considered to be statistically significant.

Results:

As shown in table 1, 2 and 3 a number of biochemical parameters were determined in the serum collected from each group. While rats fed on standard diet supplemented with garlic oil did not show any significant changes in the majority of the parameters examined, a significant increase in serum levels of glucose, bilirubin, urea and creatinine as well as the activity of the AST, ALT, LDH, and ALP enzymes were observed in rats treated with gentamicin for a period of ten days. The total protein content significantly decreased in serum. However, supplementation of gentamicin-intoxicated rats with garlic oil ameliorated the gentamicin adverse effects as evidenced by a

significant increase of serum total protein content, and a decrease of serum glucose, bilirubin, urea and creatinine levels, as well as the activity of AST, ALT, LDH and ALP enzymes. At the same time there was a significant increase in total protein and a significant decrease of serum glucose, bilirubin, urea and creatinine levels, as well as the activity of AST, ALT, LDH and ALP enzymes when supplementation of garlic oil with gentamicin intoxicated rats.

As shown in table 4,5 the same parameters were estimated in liver and kidney tissues. In the gentamicin-treated rats, a significant inhibition in total protein contents, and the activity of the enzymes ALT and AST as well as the levels of renal urea, creatinine. The activity of hepatic ALP was significantly increased in the gentamicin-treated rats. However, administration of garlic oil to the gentamicin-intoxicated rats significantly restored these parameters in the liver and kidney organs.

Histopathological changes in liver and kidney were not observed on animals treated with Garlic oil when compared with the control group (Figure 1 and 2 for liver) and (figure 5 and 6 for kidney), however animals treated with gentamicin (100mg/kg i.p.) showed fibrosis, necrosis, and fatty infiltrate in liver (Figure 4) and showed necrosis and degeneration of glomerulus in kidney (Figure 7). This effect was significantly decreased in animals pretreated with garlic oil (Figure 3 for liver and Figure 8 for kidney).

Table (1): Shows the serum liver function tests in different rat groups.

	ALT U/L	AST U/L	ALP U/L	Bilirubin mg/dl	LDH U/L
Control	21.00 ± 1.86 ^a	85.00 ± 2.89 ^a	20.17 ± 1.97 ^a	0.280 ± 0.007 ^a	50.17 ± 2.47 ^a
Gnt	39.83 ± 2.89 ^b	135.33 ± 1.54 ^c	30.17 ± 0.95 ^b	0.583 ± 0.023 ^b	71.17 ± 3.09 ^a
Gar	19.67 ± 1.26 ^a	84.50 ± 1.38 ^a	19.33 ± 1.15 ^a	0.273 ± 0.007 ^a	49.50 ± 1.59 ^c
Gnt-Gar	24.00 ± 1.15 ^a	113.50 ± 2.28 ^b	20.83 ± 0.83 ^a	0.330 ± 0.016 ^a	59.67 ± 1.58 ^b
Gar-Gnt	22.00 ± 1.86 ^a	109.83 ± 2.21 ^b	20.33 ± 0.71 ^a	0.320 ± 0.014 ^a	58.50 ± 1.69 ^{ab}

Gen Gentamicin; Gar garlic oil; ALT alanine aminotransferase; AST aspartate aminotransferase; ALP alkaline phosphatase; LDH lactate dehydrogenase; Values are expressed as mean ± SME (n=6) P values were calculated by student t-tests, mean with different superscripts (a, b, c) differ significantly, P<0.05

Table (2): Shows the serum biochemical parameters in different rat groups.

	Glucose	Cholesterol	Triglyceride
Control	101.17 ± 3.11 ^a	7.00 ± 0.86 ^a	13.33 ± 0.95 ^a
Gentamicin	251.00 ± 2.84 ^c	18.67 ± 1.09 ^b	22.50 ± 1.52 ^b
Garlic oil	99.50 ± 2.20 ^a	6.17 ± 0.65 ^a	11.83 ± 1.01 ^a
Gentamicin -Garlic oil	150.50 ± 2.45 ^b	9.50 ± 0.99 ^a	15.17 ± 1.14 ^a
Garlic oil -Gentamicin	148.67 ± 3.32 ^b	8.83 ± 0.95 ^a	14.00 ± 1.15 ^a

Values are expressed as mean ± SME (n=6) P values were calculated by student t-tests, mean with different superscripts (a, b, c) differ significantly, P<0.05

Table (3): Shows the serum kidney function tests in different rat groups.

	Creatinine	Urea	Protein
Control	1.44 ± 0.10 ^a	35.17 ± 1.35 ^a	35.50 ± 2.83 ^a
Gentamicin	2.25 ± 0.38 ^b	52.33 ± 1.93 ^b	61.33 ± 2.14 ^b
Garlic oil	1.42 ± 0.10 ^a	34.67 ± 1.45 ^a	35.17 ± 1.40 ^a
Gentamicin -Garlic oil	1.29 ± 0.05 ^a	37.50 ± 2.90 ^a	37.83 ± 0.65 ^a
Garlic oil -Gentamicin	1.27 ± 0.05 ^a	36.17 ± 2.79 ^a	36.83 ± 0.48 ^a

Values are expressed as mean ± SME (n=6) P values were calculated by student t-tests, mean with different superscripts (a, b) differ significantly, P<0.05

Table (4): Shows the hepatic tissue function tests in different rat groups.

	AST U/mg		ALT U/mg		ALP U/mg	
Control	11.70	± 0.38 ^b	2.80	± 0.39 ^b	171.62	± 2.56 ^a
Gentamicin	7.60	± 0.50 ^a	1.10	± 0.04 ^a	272.65	± 5.24 ^b
Garlic oil	12.20	± 0.32 ^b	3.00	± 0.39 ^b	169.27	± 2.58 ^a
Gentamicin -Garlic oil	11.98	± 0.59 ^b	2.70	± 0.42 ^b	234.68	± 26.08 ^{ab}
Garlic oil -Gentamicin	12.05	± 0.54 ^b	2.80	± 0.37 ^b	234.80	± 36.50 ^{ab}

ALT alanine aminotransferase; AST aspartate aminotransferase; ALP alkaline phosphatase;

Values are expressed as mean ± SME (n=6) P values where calculated by student t-tests, mean with different superscripts (a, b) differ significantly, P<0.05

Table (5): Shows the renal tissue function tests in different rat groups.

	Urea		Creatinine	
Control	33.73	± 3.72 ^b	1.04	± 0.01 ^b
Gentamicin	12.42	± 0.07 ^a	0.63	± 0.09 ^a
Garlic oil	34.50	± 3.46 ^b	1.06	± 0.01 ^b
Gentamicin + garlic oil	26.00	± 0.68 ^b	1.16	± 0.04 ^b
Garlic oil + gentamicin	27.93	± 0.69 ^b	1.15	± 0.01 ^b

Values are expressed as mean ± SME (n=6) P values where calculated by student t-tests, mean with different superscripts (a, b) differ significantly, P<0.05

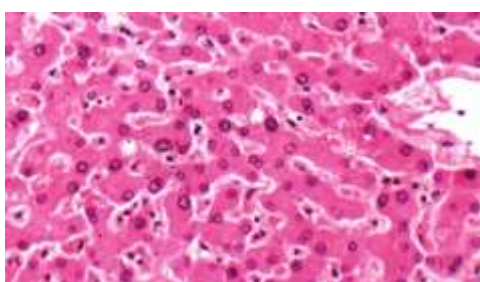


Figure (1) Liver section from a control rat showing normal morphology.

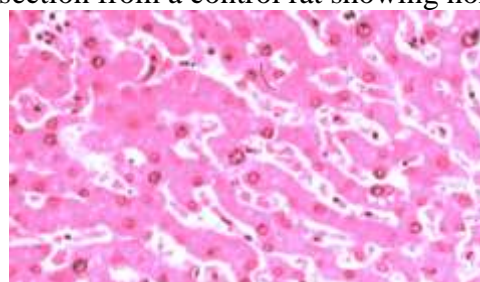


Figure (2) Liver section from rat treated with garlic oil showing normal morphology when compared with control rat.

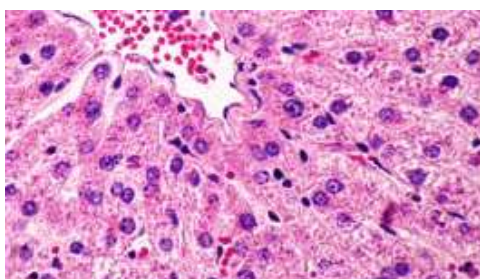


Figure (3) Liver section from rat treated with garlic oil prior to and during treatment with gentamicin, the section shown reduction of fibrosis, necrosis, and fatty infiltrate when compared with animals treated with gentamicin alone .

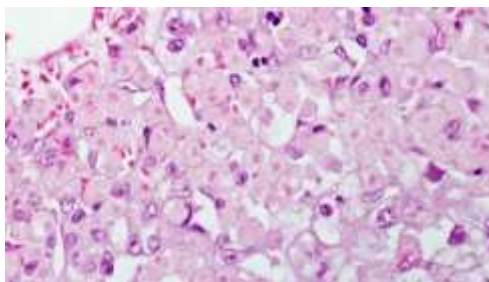


Figure (4) Liver section from rat treated with gentamicin showing fibrosis, necrosis, and fatty infiltrate.

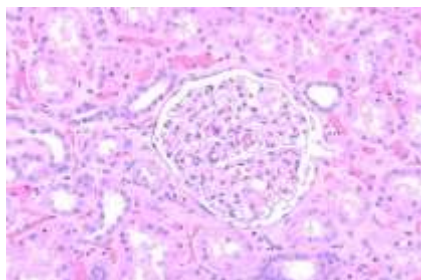


Figure (5) Kidney section from control group showing the normal structure of glomerulus.

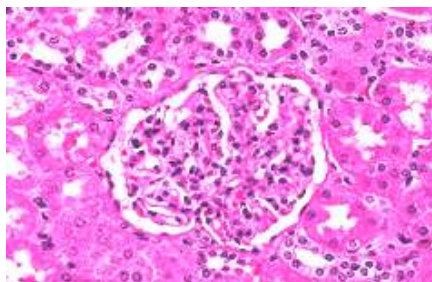


Figure (6) Kidney section from rat treated with garlic oil showing normal morphology when compared with control rat.

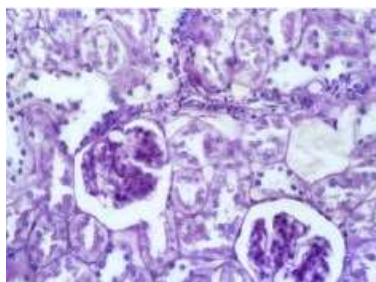


Figure (7) Kidney section from rat treated with gentamicin showing necrosis and degeneration of glomerulus.

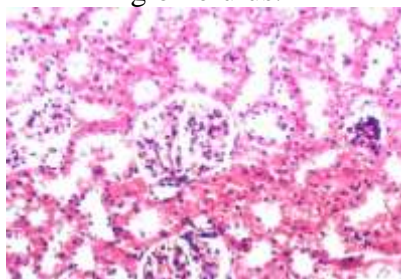


Figure (8) Kidney section from rat treated with gentamicin and garlic oil showing a reduction of necrosis and degeneration of glomerulus.

Discussion:

Gentamicin treatment causes significant increase in the serum level of liver and kidney function markers such as creatinine, urea, total cholesterol, triglycerides, bilirubin and glucose as compared to control indicating hepatorenal dysfunction. These injuries may be due to the production of free radicals and involvement of oxidative stress to hepatorenal toxicity caused by gentamicin treatment. Gentamicin may influence the various metabolic pathways of liver thereby enhancing the level of total cholesterol, triglycerides, glucose, bilirubin, urea and creatinine in serum.

This study evaluated kidney function by measuring serum creatinine and urea values. Gentamicin treatment is found to elevate creatinine and urea levels in serum, but decreased in the kidney, suggesting an impairment of kidney functions. These effects could also be attributed to the aminoglycoside induced nephrotoxicity is characterized by a decrease in the glomerular filtration rate and direct tubular injury. These observations are generally in agreement with other studies^{32,33}. Creatinine and urea are waste products of protein metabolism that need to be excreted by the kidney, therefore a marked increase of these parameters, as observed in this study, confirms an indication of functional damage to the kidney³⁴. Urea level can be increased by many other factors such as dehydration, antidiuretic drugs and diet, while creatinine is more specific to the kidney, since kidney damage is the only significant factor that increases the serum creatinine level³⁵. However, administration of garlic oil along with gentamicin caused significant decrease in urea and creatinine suggested the protective effects of garlic oil. This result agrees with Hassan et al³⁶, who reported the administration of garlic oil significantly reduced the concentration of urea and creatinine in sodium nitrite induced rats.

The present study showed that the injection of 100mg/kg gentamicin once daily for ten days to rats lead to a marked elevation in the levels of serum AST, ALT, LDH and ALP which is indicative of hepatocellular damage. This elevation may be attributed to the release of these enzymes from the cytoplasm into the blood circulation after rupture of the plasma membrane and cellular damage. Serum AST, ALT, LDH and ALP are biomarkers in the diagnosis of hepatic damage because they are released into the circulation after cellular damage³⁷. Increased level of LDH in serum indicated the toxic effects of gentamicin in rat. This result obtained in this study agrees with other reports¹⁰.

Garlic has been found to have an important dietary and medicinal role for centuries. Most of its prophylactic and therapeutic effects are ascribed to specific oil and water soluble organosulfur compounds. Thiosulfonates and other secondary metabolites of garlic, including steroids, terpenoids, flavonoids and other phenols, may be responsible for reported therapeutic effects of garlic. Reuter *et al*³⁸ have reviewed the therapeutic effects of garlic on cardiovascular system as well as its antibiotic, anticancer, antioxidant, immunomodulatory, anti-inflammatory, hypoglycemic and hormone-like effects. Garlic also increases anti-inflammatory monocyte IL-10 production and decreases proinflammatory cytokines such as TNF- α , IL-1 β , IL-6, IL-8, T cell interferon gamma, IL-2.³⁸⁻⁴⁰ Administration of garlic oil significantly reduced the activity of liver enzymes in gentamicin induced rats. Due to its ability to reduce free radical-induced oxidative damage in the liver,⁴¹ garlic extract has been shown to decrease liver enzymes in serum and prevent liver damage of rats with liver fibrosis⁴².

There was a significant decrease in the concentration of serum total protein in gentamicin group as compared to control group, might be depressed as a result of defective protein synthesis. However, administration of garlic oil along with gentamicin caused significant increase in total protein suggested the protective effects of garlic oil.

There was a significant increase in levels of serum ALT, AST, LDH, ALP, urea, creatinine, glucose, cholesterol, triglycerides and bilirubin in gentamicin group as compared to control group. However, administration of garlic oil for 5 days before treatment with gentamicin caused significant decrease in levels of serum of ALT, AST, LDH, ALP, urea, creatinine, glucose, cholesterol, triglycerides and bilirubin as compared to gentamicin groups suggested the prophylactic roles of garlic oil.

Conclusion

From this study we can conclude there are a protective and prophylactic effects of garlic oil against gentamicin induce damage, that return the abnormal value of the serum biochemical parameters of ALT,AST,ALP, bilirubin, lactate dehydrogenase, glucose, cholesterol, triglyceride, creatinine , urea and protein as well as hepatic tissue AST,ALT and ALP and renal tissue urea and creatininehistopathological changes in renal and hepatic tissue to normal.

References:

1. Ali BH. Gentamicin nephrotoxicity in humans and animals: Some recent research. *Gen Pharmacol*. 1995; 26: 1477-87.
2. Mathew TH. Drug-induced renal disease. *Med J Aust*. 1992; 156: 724-28.
3. Homes HD, Weinberg JM. Toxic nephropathies. In: *Kidney*. Brenner, BM, Rector FC Jr. (eds). Philadelphia, Saunders Co, 1986;1491-533.
4. Costa Silva VL, Gil FZ, Cavanal MF: Evaluation of distal tubules function in aminoglycoside induced nephropathy. *Brz J Med Biol Res* 1987, 20:833-836.
5. Ramsammy L, Ling KY, Josepovitz C, Levine R, Kaloyanides GJ. Effect of gentamicin on lipid peroxidation in rat renal cortex. *BiochemPharmacol*. 1985; 34: 3895-900.
6. Yang C, Du X, Han Y. Renal cortical mitochondria are the source of oxygen free radicals enhanced by gentamicin. *Renal Fail*. 1995; 17: 21-26.
7. Fantone JC, Ward PA. Role of oxygen-derived free radicals and metabolites in leukocyte-dependent inflammatory reactions. *Am J Pathol* 1992;107:397-418.
8. Weiss SJ, LoBuglio AF. Phagocyte-generated oxygen metabolites and cellular injury. *Lab Invest* 1982;47:5-18.
9. Fox RB. Prevention of granulocyte-mediated oxidative injury in rats by a hydroxyl radical scavenger, dimethylthiocarbate. *J Clin Invest* 1984;74:1456-64.
10. Khan M.BadarI,SiddiquahA.Prevention of hepatorenal toxicity with Sonchusasper in gentamicin treated rats.*BMC Complementary and A lternative Medicine* 2011,11:113.
11. Rodriguez-Barbero A, Lopez-Novoa JM, Arevalo M: Involvement of plateletactivating factor in gentamicin nephrotoxicity in rats. *ExpNephrol* 1997, 5:47-54.
12. Sastrasinh M, Knauss TC, Weinberg JM, Humes HD: Identification of the aminoglycoside binding site in rat renal brush border membranes. *J PharmacolExpTher* 1982, 222:350-358.
13. Moestrup SK, Cui S, Vorum H, Bregengard C, Bjem SE, Norris K, Gliemann J, Christensen EI: Evidence that epithelial glycoprotein 330/megalin mediates uptake of polybasic drugs. *J Clin Invest* 1995, 96:1404-1413.
14. Block E. The chemistry of garlic and onions. *Sci Am* 1985;252:114-9.
15. Daniel B. Mowrey. *The Scientific Validation of Herbal Medicine*. (New Canaan, Connecticut: Keats Publishing, 1986), 122.
16. Khanum F, Anilakumar KR, Viswanathan KR. Anticarcinogenic properties of garlic: a review. *Crit Rev Food SciNutr*2004; 44: 479-488
17. Banerjee SK, Maulik M, Manchanda SC, Dinda AK, Das TK, Maulik SK. Garlic-induced alteration in rat liver and kidney morphology and associated changes in endogenous antioxidant status. *Food ChemToxicol*2001; 39:793-797
18. Liu J, Lin RI, Milner JA. Inhibition of 7,12-dimethylbenz[a]anthracene-induced mammary tumors and DNA adducts by garlic powder. *Carcinogenesis* 1992;13:1847-51.
19. Amagase H, Milner JA. Impact of various sources of garlic and their constituents on 7,12-dimethylbenz[a]anthracene binding to mammary cell DNA. *Carcinogenesis* 1993;14:1627-31.
20. Song K, Milner JA. Heating garlic inhibits its ability to suppress 7, 12-dimethylbenz(a)anthracene-induced DNA adduct formation in rat mammary tissue. *J Nutr* 1999;129:657-61.
21. Hodge G, Hodge S, Han P. *Allium sativum* (garlic) suppresses leukocyte inflammatory cytokine production in vitro: potential therapeutic use in the treatment of inflammatory bowel disease. *Cytometry*2002; 48: 209-215.

22. Alhamami O,AL-Mayah J,AL-MousawiN,etal.Effects of garlic on haemostatic parameters and lipid profile in hyperlipidemicrats:antiatherogenic and antithromboiticeffects.EasternJournal of Medicin:2006;11:13-18.
23. Chen HW, Tsai CW, Yang JJ, Liu CT, Kuo WW and Lii CK. The combined effects of garlic oil and fish oil on the hepatic antioxidant and drug-metabolizing enzymes of rats. British J Nutr 2003; 89:189-200.
24. Trinder P. A colorimetric method for the determination of glucose. Ann ClinBiochem1969; 6:24-26.
25. Reitman S and Frankel S. A colorimetric method for the determination of serum glutamic oxaloacetic and glutamic pyruvic transaminase. Am J Clin 1957; 28:56-63.
26. Owen JA, Iggo B, Scandrett FJ, et al. The determination of creatinine in plasma or serum, and in urine: a critical examination. Biochem J 1954;58: 426- 437.
27. Chaney AL, Merbach EP. Modified reagents for determination of urea and ammonia. ClinChem 1962; 8: 130- 132.
28. Henry RJ. Principles and Techniques. Clinical Chemistry, 2nd Ed. Harper and Row 1974:525.
29. Belfield A, Goldberg DM. Revised assay for serum phenyl phosphatase activity using 4-amino-antipyrine. Enzyme 1971;12:561-73.
30. Zlatki A, Zak B, Boyle AJ. A new method for the direct determination of serum cholesterol. J Lab Clin Med 1953; 45: 486.
31. Foster LB, Dunn RT. Stable reagents for determination of serum triglycerides by a colorimetric Hantzsch condensation method. ClinChem1973; 19: 338-340.
32. Safa J, Argani H, Bastani B, Nezami N,Ardebili BR, Ghorbanihaghjo A, Kalagheichi H, Amirfirouzi A, Mesgari M, Rad JS: Protective effect of grape seed extract on gentamicin- induced acute kidney injury. Iran J Kid Dis 2010, 4:285-291.
33. Reiter M, Rupp K, Baumeister P, Zieger S, Harreus U: Antioxidant effects of quercetin and coenzyme Q10 in mini organ cultures of human nasalmucosa cells. Antican Res 2009, 29:33-40.
34. Panda, N.C. Kidney. In: Textbook of Biochemistry and Human biology. second ed., Prentise hall India, 1999 290-296.
35. Cheesbrough, M., Clinical chemistry tests. In: District laboratory practice in tropical countries. Cambridge new edition, part II.998, 331-363.
36. Hassan H,El-Agmy S, Gaur R. *In vivo* evidence of hepato- and reno-protective effect of garlic oil againstsodium nitrite-induced oxidative stress.International Journal of Biological Sciences 2009; 5(3):249-255.
37. Naik SR, Panda VS. Antioxidant and hepatoprotective effects of Ginkgo bilobaphytosomes in carbon tetrachloride-induced liver injury in rodents. Liver Int 2007; 27:393-9.
38. Reuter HD, Koch HP and Lawson DL. Therapeutic effects and applications of garlic and its preparation. In: Garlic: The science and therapeutic application of *Allium sativum*L and related species,2nd ed.(Koch,HP and Lawson,DL.eds), William and Wilkins, Baltimore MB, pp 135 – 212.
39. Hodge G, Hodge S, Han P. *Allium sativum* (garlic) suppresses leukocyte inflammatory cytokine production in vitro: potential therapeutic use in the treatment of inflammatory bowel disease. *Cytometry*2002; 48: 209-215.
40. Pal R, VaipheiK,SikanderA,etal.Effect of garlic on isoniazid and rifampicin-induced hepatic injury in rats.WorldJournal of Gastroenterology 2006;12(4):636-639.
41. Gedik N, Kabasakal L, Sehirli O, et al. Long-term administration of aqueous garlic extract (AGE) alleviates liver fibrosis and oxidative damage induced by biliary obstruction in rats. Life Sci 2005;76:2593-606.
42. Nakagawat S, Kasug S, Matsuura H. Prevention of liver damage by aged garlic extract and its components in mice. Phytotherapy Research 1989;3:50 - 53.