



# Animal hygiene assessment by spontaneous reactivation and aging kinetics of pesticide compounds

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## ABSTRACT

*The main aims of this paper were to investigate the rate of spontaneous reactivation of butyrylcholinesterase from plasma and serum of farm animals inhibited by the organophosphorus, chlorfenvinphos and trichlorfon. Organophosphorus are among the most toxic of all substances that cause poisoning in farm animals and are the most frequently encountered insecticides, commonly detected in agricultural products, animal-derived foodstuffs, environmental samples and home use and represent a significant potential health risk. The first-order rate constants obtained for spontaneous reactivation ( $k_s$ ) for chlorfenvinphos and trichlorfon for plasma and serum was ranged between 0.012 to 0.811h<sup>-1</sup>, while aging of phosphorylated butyrylcholinesterase follows the kinetics of a first-order reaction with rate constants aging ( $k_a$ ) ranged between 0.004 to 0.653 h<sup>-1</sup>. Half-time ( $t_{1/2}$ ) for spontaneous reactivation and aging are higher in trichlorfon compared to chlorfenvinphos and ranged from 1.1 to 28.9h (sheep), 2.1 to 63 h (cattle) and 2.8 to 53.3 h (goat), respectively.*

**Keywords:** Spontaneous reactivation, Aging, Farm animals, animal hygiene, pesticide compounds.



## تقييم صحة الحيوان بواسطة اعادة التنشيط الذاتي وديناميكية الهرم للمركبات المبيدة للحشرات

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### الملخص

ان الهدف الرئيسي من هذه الدراسة هو ايجاد معدل التنشيط الذاتي لانزيم *butrylcholinesterase* في بلازما ومصل الدم في الحيوانات الحقلية والمثبطة بواسطة المركبات الفسفورية العضوية، *chlorfenvinphos* و *trichlorfon*. والمعروف ان المركبات الفسفورية العضوية من بين اكثر المركبات السامة المسببة للتسمم في الحيوانات الحقلية ، ومن اغلب المبيدات الحشرية المألوفة حيث نجدها اكثر شيوعا في المحاصيل الزراعية، والمواد الغذائية المشتقة من الحيوانات وفي عينات البيئة والمنزل كما يمثل المخاطر الكامنة الموعثرة على الصحة . تم الحصول على ثابت معدل الرتبة الاولى لاعادة التنشيط الذاتي (ks) لمركب *chlorfenvinphos* و *trichlorfon* في بلازما ومصل الدم تراوحت بين 0.012 الى  $h^{-1}$  0.811 ، بينما هرم الانزيم *butrylcholinesterase* المفسفر يسلك تفاعل ديناميكية الرتبة الاولى مع معدل ثابت الهرم (ka) تراوحت ما بين 0.004 الى  $h^{-1}$  0.653 . نصف العمر (t1/2) لاعادة التنشيط الذاتي والهرم كانت اعلى في *trichlorfon* بالمقارنة مع *chlorfenvinphos* وتراوحت من 1.1 الى 28.9 h (الانعام) ، 2.1 الى 63 h (الابقار) و 2.8 الى 53.3 h (الماعز) ، على التوالي .



نستنتج من هذه الدراسة ، بان معدل الهرم كانت بطيئه جدا من اعادة التنشيط الذاتى لانزيم **butyrylcholinesterase** فى بلازما دم الحيوانات الحقلية (الابقار ، الاغنام ،والماعز ) والمثبته مع كل من **chlorfenvinphos** و **trichlorfon**. وهذه تنطبق ايضا لمصل الدم مع المركبين المذكورين على الرغم من وجود فرق ضئيل .

**الكلمات الدالة :** اعادة التنشيط الذاتى، الهرم، صحة الحيوان، الحيوانات الحقلية، المركبات المبيدة للحشرات.

## 1. INTRODUCTION

Organophosphorus insecticides still pose a main problem in animal hygiene. The use of organophosphorus compounds for pest control and endeavoured suicide reasons huge numbers of intoxications and several hundreds of thousands of fatalities per year mainly in developing countries [1-3]. In the current study, we have chosen chlorfenvinphos and trichlorfon. Both of them are routinely used in veterinary medicine. Organophosphorus compounds products are prevalent in animals destined for human consumption in the world with serious public health implications. Animal handlers are at risk of pollution and can serve as source of contamination to vulnerable hosts. Targeted pest control on farm animals, concerted veterinary efforts, professional health instruction, active attachment of farm animal careers and good health care systems are necessary for effective control [1, 4, 5].

The contamination with organophosphorus causes a generalized cholinergic crisis due to an irreversible inhibition of butyrylcholinesterase (acylcholineacylhydrolase, EC 3.1.1.8) by phosphorylation of their active site serine [1, 6] and successive accumulation of the neurotransmitter butyrylcholine. Phosphorylated butyrylcholinesterase is spontaneously hydrolysed, liberating phosphoric acid and the original active butyrylcholinesterase enzyme. This phenomena spontaneous reactivation, which proceeds very slowly and depends on the leaving group of the original organophosphorus inhibitor, but on the residual substituted groups on the phosphorus atom and the source of butyrylcholinesterase enzyme [1, 7].



However, organophosphorus-inhibited butyrylcholinesterase changes gradually into a non-reactivatable form on storage, then this phenomena is called aging. It was assumed that the aging might be caused by a migration of the phosphoryl group from an initial position to form more stable bond or by the elimination of serine phosphate to lose serine hydroxyl group. Generally is accepted that spontaneous reactivation and aging mechanism for alkoxy group of organophosphorus residue bound to butyrylcholinesterase[8, 9]. Recently, the standard treatment for organophosphorus poisoning comprises a mixture of atropine, an oxime, e.g. obidoxime. The oxime compounds that can reactivate organophosphorus-inhibited butyrylcholinesterase by attaching to the phosphorus atom of the organophosphorus compounds forming an oxime-phosphonate, which ruptures away from the butyrylcholinesterase molecule [1, 10].The main aims of this paper were to investigate the rate of spontaneous reactivation of butyrylcholinesterase from plasma and serum of farm animals inhibited by the chlorfenvinphos and trichlorfon. As well as, to investigate the time course of aging of organophosphorus-inhibited butyrylcholinesterase from plasma and serum of farm animals.

## **2. MATERIALS AND METHODS**

Samples were collected from healthy farm animals (sheep, cattle and goats) from local abattoirs were used in this paper. The samples (plasma and serum) were transported on ice to the laboratory for immediate processing. To obtain serum blood samples were allowed to clot for at least 2 h at 30 °C, after which they were centrifuged at 6000 g for 15 min. To obtain plasma 4 ml blood samples were added to anticoagulant heparin, 3.7 mg, 0.1% in 10 ml centrifuge tubes. Plasma was separated by centrifugation at 6000 g for 10 min [11-13].Butyrylcholinesterase activity was determined by the Ellman method [14], adapted for use with microtitre plates as described by Haigh et al. [15] using butyrylthiocholine iodide as substrate (2 mM final concentration) for measuring butyrylcholinesterase activity. Organophosphorus-inhibited butyrylcholinesterase was prepared by incubating samples with



appropriate organophosphorus concentrations for 45 min at 30 °C resulting in an inhibition of 90–98% of control activity. Organophosphorus-treated samples were stored in aliquots at -60 °C until use. In order to remove excess organophosphorus after affecting inhibition the samples by chlorfenvinphos and trichlorfon, the samples were filtrated and the absence of inhibitory activity was tested by incubation of organophosphorus-treated and control butyrylcholinesterase (20 min, 30 °C). Aliquots were taken after various time intervals for determination of butyrylcholinesterase activity ( $k_s$ ) and of the decrease of oxime-induced reactivation ( $k_a$ ). Organophosphorus-treated samples were incubated with 250 $\mu$ M obidoxime (45 min.) [16, 17].

#### *Statistical analysis*

The method was used to calculate the means and standard errors (SE). Differences with  $P < 0.05$  were regarded to have statistical significance. The pseudo-first-order rate for constants  $k_s$  and  $k_a$  were calculated by a non-linear regression analysis [1, 17].

### **3. RESULTS AND DISCUSSION**

This is the first study, which compares the  $k_s$  and  $k_a$  of organophosphorus inhibited butyrylcholinesterase in the plasma and serums of farm animals. Phosphorylated butyrylcholinesterase is susceptible to spontaneous hydrolysis of an alkyl-ester bond, resulting in a negatively charged residue which is resistant towards nucleophilic attack [18, 19]. The rate of  $k_{sto}$  inhibit erythrocyte butyrylcholinesterase by chlorfenvinphos has been observed to be 0.92 min<sup>-1</sup> for cattle [17, 20], 0.347 h<sup>-1</sup> for rat [21], while  $k_s$  values to trichlorfon was 0.012 h<sup>-1</sup> for human [21]. The  $k_s$  and  $k_a$  kinetics of butyrylcholinesterase inhibited by chlorfenvinphos and trichlorfon were determined in plasma and serum for sheep, cattle and goat using Ellman method as described in the section of Materials and Methods (**Tables 1 and 3**). Plasma  $k_s$  and  $k_a$  kinetic parameters for reaction between butyrylcholinesterase and two organophosphorus (chlorfenvinphos and trichlorfon) are shown in Table 1. The  $k_s$  for the three animals was decreased according to the rank order of sheep > cattle > goat for chlorfenvinphos and trichlorfon.



However,  $k_a$  values in plasma for animals was reduced in order of cattle >goat >sheep for chlorfenvinphos and trichlorfon. The relative activity between chlorfenvinphos and trichlorfon in cattle found highest  $k_s(4)$  and lowest in  $k_a$  (0.5). Kinetic  $k_s$  and  $k_a$  of all tested animals in plasma gave no correlation between  $k_a$  and  $k_s$  ( $R^2 < 0.24$ ) (Table 1).

**Table 1.** Rate constants for the spontaneous reactivation ( $k_s$ ) and aging ( $k_a$ ) of butyrylcholinesterase inhibited by chlorfenvinphos and trichlorfon from plasma of sheep, cattle and goat.

Inhibitor	Sheep	Cattle	Goat
<b>ks(h-1)</b>			
<b>Chlorfenvinphos</b>	0.645±0.19	0.324±0.05a	0.244±0.04a
<b>Trichlorfon</b>	0.188±0.06	0.081±0.021b	0.068±0.006b
<b>Ratioa</b>	3.4	4	3.7
<b>ka(h-1)</b>			
<b>Chlorfenvinphos</b>	0.028±0.013a	0.053±0.006a	0.045±0.014
<b>Trichlorfon</b>	0.004±0.002 b	0.150±0.007 b	0.067±0.014
<b>Ratioa</b>	7	0.5	0.67

Values in the table are mean±SE obtained from nonlinear regression analysis, each performed in triplicate (n=4 in each animal). Different letters in column is significantly different (ANOVA,  $P < 0.05$ ). aRatio (chlorfenvinphos vs. trichlorfon).

The half time ( $t_{1/2}$ ) from plasma for  $k_s$  and  $k_a$  kinetic parameters for reaction among butyrylcholinesterase and (chlorfenvinphos and trichlorfon) are shown in Table 2.

All farm animals studied in this research, the half time for trichlorfon was higher than chlorfenvinphos for spontaneous reactivation, in contrast with the values of aging (Table 2).



**Table 2.** Half times for the ks and ka of butyrylcholinesterase inhibited by chlorfenvinphos and trichlorfon from plasma of sheep, cattle and goat.

Inhibitor	Sheep	Cattle	Goat
<b>ks(t1/2)</b>			
<b>Chlorfenvinphos</b>	1.07 ± 0.16	2.14 ± 0.75a	2.83 ± 0.84a
<b>Trichlorfon</b>	3.69 ± 1.57	8.56 ± 0.98b	10.5 ± 2.87b
<b>ka(t1/2)</b>			
<b>Chlorfenvinphos</b>	24.75 ± 4.55	13.08 ± 1.675a	15.40 ± 1.36
<b>Trichlorfon</b>	28.88 ± 3.54	4.62 ± 0.346b	10.34 ± 3.87

The letters between the chlorfenvinphos and trichlorfon is significantly different (ANOVA, P< 0.05).

Serum ks and ka kinetics for reaction among butyrylcholinesterase and two organ phosphorus (chlorfenvinphos and trichlorfon). The values of ks for animals were decreased according to the rank order of goat >sheep > cattle for chlorfenvinphos while in range sheep >goat >cattle for trichlorfon. Instead, first-order rate constants for ka values was decreased according to the rank order of goat >cattle >sheep for chlorfenvinphos while decreased: sheep >goat> cattle for trichlorfon. Chlorfenvinphos/trichlorfon were 1.9, 8.9 and 18.9, respectively for ks and 0.8, 3 and 50.8, respectively for ka. The comparison between the ks and ka kinetics in the serums of all tested animals gave also poor correlation between ka and ks (R2 < 0.22) (**Table 3**).



**Table 3.** Rate constants for the spontaneous reactivation ( $k_s$ ) and aging ( $k_a$ ) of butyrylcholinesterase inhibited by chlorfenvinphos and trichlorfon from serum of sheep, cattle and goat.

Inhibitor	Sheep	Cattle	Goat
<b>ks(h-1)</b>			
<b>Chlorfenvinphos</b>	0.231±0.046	0.107±0.05a	0.811±0.012a
<b>Trichlorfon</b>	0.123±0.219	0.012±0.03b	0.043±0.05b
<b>Ratioa</b>	1.9	8.9	18.9
<b>ka(h-1)</b>			
<b>Chlorfenvinphos</b>	0.027±0.003	0.034±0.007a	0.653±0.004a
<b>Trichlorfon</b>	0.033±0.015	0.011±0.009b	0.013±0.02b
<b>Ratioa</b>	0.8	3	50.2

Values in the table are mean  $\pm$  SE obtained from nonlinear regression analysis, each performed in triplicate (n = 4 in each animals). Different letters in column is significantly different (ANOVA, P < 0.05). aRatio (chlorfenvinphos vs. trichlorfon).

Half time ( $t_{1/2}$ ) from serum for  $k_s$  and  $k_a$  kinetic parameters for reaction among butyrylcholinesterase and two organophosphorus (chlorfenvinphos and trichlorfon) are shown in Table 4. Again the half time in all farm animals, the trichlorfon was higher than chlorfenvinphos for  $k_s$ , while chlorfenvinphos for aging was much higher (5-times) than that of trichlorfon (Table4). Other than  $t_{1/2}$  for aging was nearly similar in trichlorfon than chlorfenvinphos in goat (Table4).



**Table 4.** Half times for ks and ka of butyrylcholinesterase inhibited by chlorfenvinphos and trichlorfon from serum of sheep, cattle and goat.

Inhibitor	Sheep	Cattle	Goat
ks(t1/2)			
Chlorfenvinphos	3.0 ± 0.654	6.48 ± 0.309a	6.24 ± 0.665a
Trichlorfon	5.64 ± 1.232	57.75 ± 5.87b	16.12 ± 1.765b
ka(t1/2)			
Chlorfenvinphos	25.67 ± 3.87	20.38 ± 1.24a	231 ± 9.46a
Trichlorfon	21.0 ± 1.82	63.0 ± 5.77b	53.31 ± 2.76b

The letters between the chlorfenvinphos and trichlorfon is significantly different (ANOVA,  $P < 0.05$ ).

Comparable to our work the chlorfenvinphos and trichlorfon was higher than above results. However, ks in the plasma proceeded substantially faster with goat chlorfenvinphos and trichlorfon compared with the sheep and cattle. While serum ks was faster with cattle exposed to chlorfenvinphos and trichlorfon compared with sheep and goat (Tables 1 and 3). Literature values of the half time (t1/2) of ks for butyrylcholinesterase are about 2 and 58 h for organophosphorus in human erythrocyte [21, 22]. The recovery of rate ka of reserved erythrocyte butyrylcholinesterase by organophosphorus was  $2.62 \times 10^4 \text{ min}^{-1}$  and  $7.77 \times 10^4 \text{ min}^{-1}$  for cattle and horse, respectively [16]. The serum trichlorfon-inhibited butyrylcholinesterase ka proceeded markedly later than ka of sheep, cattle and goats. In addition, t1/2 of ka for human erythrocyte butyrylcholinesterase is about 41 h for organophosphorus [21], concurring with our results which ranged between 21 to 231 h in serum and ranged between 4.62 to 24.75 in plasma (Table 4). This indicates that the reduction of chlorfenvinphos t1/2 in the plasma and serum might alter the use of oximes. In clinical research, the level and time course of ka is important, because it is the factor that limits the period for useful oximes administration after affecting farm animals with organophosphorus pesticides [1, 16, 23]. Finally, this paper establishes that values



of  $k_a$  and/or  $k_s$  could play a role on the administration of oximes in the plasma and serums of sheep, cattle and goats.

#### 4. CONCLUSIONS

This research is provided original data concerning an enzymological characterization of these inhibitors in these farm animals. As well as in this research, we surveyed recent developments in our understanding of the kinetic properties of  $k_s$  and  $k_a$  for sheep, cattle and goat butyrylcholinesterase is comparable in view of interactions with chlorfenvinphos and trichlorfon. In plasma from all three animals studied (sheep, cattle and goats), the rate of the aging process is much slower than spontaneous reactivation of butyrylcholinesterase inhibited with both of them (chlorfenvinphos and trichlorfon). This was also true in serum with chlorfenvinphos and trichlorfon, although the difference was less. Hence, in the case of  $t_{1/2}$  butyrylcholinesterase inhibited with trichlorfon the enzyme will tends to become much higher than chlorfenvinphos. The determination of reactivation and aging constants of chlorfenvinphos and trichlorfon organophosphorus pesticides with farm animals butyrylcholinesterase indicates that a structure activity relationship can be derived for inhibition as well as for spontaneous reactivation but not for aging and oxime-induced reactivation.

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