

The in vivo effect of some medicinal plant extracts on *Cryptosporidium* parasite.

Hiro M. Obiad*.. Tawfiq I. Al-Alousi**... Abdulla H. Al-Jboori***



* Foundation of Technical Education - Kirkuk Technical College

** Tikrit University - Veterinary Medicine College

*** Tikrit University - Education College.

ARTICLE INFO

Received: 14 / 9 /2011
Accepted: 24 / 4 /2012
Available online: 29/8/2013
DOI: 10.37652/juaps.2012.78251

Keywords:

in vivo ,
medicinal plant extracts ,
Cryptosporidium parasite.

ABSTRACT

Cryptosporidium is known as a parasite of humans especially children of both those who are immunodeficient and immunocompetent. The effect of prolonged diarrhea and dehydration can be dangerous, especially for young and immunodeficient persons. This study was designed to find out the watery and alcoholic effect of some medicinal plant extracts against the parasite. No significant differences between the watery and alcoholic extracts of the three medicinal plant used (*Corindrum sativum*, *Curcuma longa*, *Viscum album*) were noted. All the plants were dose related, *Curcuma longa* had the highest effect on *Cryptosporidium* oocysts shedding in laboratory infected Balb/c mice. With rate of 100% on the 7th day of treatment at 750 mg/kg and on the 5th day at 1000mg/kg in the watery extracts. And a rate of 100% on the 4th day at 1000mg/kg in alcoholic extracts, followed by *Viscum album* with rate of 48, 54% on the 7th day at 750, 1000mg/kg respectively for watery extract and 73, 76% on the 7th day at 750, 1000mg/kg respectively for alcoholic extract. The *Coriondrum sativum* had the lowest effect at all concentrations used in both watery and alcoholic extracts. No significant differences were seen with folic acid and potassium chloride adding to the watery and alcoholic plant extracts. Except with the antibiotic (azithromycin) in which its activity was increased to 100% on the 4th day of treatment whereas its effect was only 68% without them.

Introduction.

One of the most biologically intriguing, and clinically frustrating features of cryptosporidiosis is its resistance to antimicrobial drugs. Unlike many of its relative (*Toxoplasma*, *Eimeria*, and *Plasmodium*), there is no curative therapy for cryptosporidiosis, despite the *in vitro* and *in vivo* testing of many compounds. One possible explanation for this is that *Cryptosporidium* establishes a compartment within the host cell, which is morphologically different from the setting used by the related parasites. This unique parasitophorous vacuole may somehow shelter the parasite from antimicrobial drugs [1].

Because the clinical course of cryptosporidiosis depends largely on the immune status of the host, treatment options vary accordingly. In immunocompetent adults and children, no specific therapy is indicated, since the disease is self-limiting. However, as in any diarrheal illness, hydration must be carefully monitored. In immunocompromised hosts, particularly AIDS patients with CD4 cell counts below 200/mm³, cryptosporidiosis can be life-threatening and must be treated aggressively. Initially the nutritional hydration, and electrolyte status of the patient should be assessed and corrected with intravenous hydration, if necessary, antimotility agents such as opiates and somatostatin analogues may also be used. Several antibiotics that have some efficacy against *Cryptosporidium* have been reported, spiramycin and dicalzuril sodium have produced partial responses against the parasite (partial decrease in diarrhea and

* Foundation of Technical Education - Kirkuk Technical College.E-mail address:

decreases in stool oocyst number) [2]. Paromomycin has been shown to decrease the intensity of infection and improve intestinal function and morphology, paromomycin is a poorly absorbed broad spectrum antibiotic similar to neomycin [3]. The *in vitro* activity of nitazoxanide alone and in combination with azithromycin and rifabutin was investigated, nitazoxanide had showed moderate anti cryptosporidial activity (>50%), a parasite reduction of 79.8-83.9% was observed when nitazoxanide was combined with azithromycin and rifabutin [4]. When azithromycin was used alone for cryptosporidial diarrhea treatment in AIDS patients it showed good reduction in symptoms but didn't eradicate the oocysts from the stool [5].

. An experiment on more than 100 drugs *in vitro* only 40 drugs showed some affection on the parasite, no one of the antiparasitic drugs used (guinive, chloroquine, Pyrimethamin, difluoromethyl orinthine ,trimethoprim-sulfamethoxazol, diclazuril) had affected the parasite[6]. Fujikawa [7] believes that paromomycin sulfate is an effective drug against *Cryptosporidium* and because of difficulties for getting the drug it was replaced by clarithromycin.

.The infection may become chronic and life-threatening which can lead to death in some individuals specifically children, the elderly, and immnosuppressed patients. There is no completely satisfactory treatment for cryptosporidial enteritis has been successfully developed, and a wide variety of medications and components have been tested as possible treatments for the illness but there is currently no drug that can cure cryptosporidiosis therefore efforts are still needed to develop an effective drug [8, 9]. Our study is a trial for finding a treatment for the parasite.

Materials and methods.

Oocysts preparation and isolation.

. *Cryptosporidium* oocysts were obtained from human feces which were diagnosed by Modified Ziehl-Nelson (M.Z.N.) procedure[10].The positive stool samples were diluted with saline or D.W and sieved through stainless steel mesh (75 μ m). The sieved stool was distributed in centrifuge tubes (0.5ml of diluted stool + 10ml of D.W) and centrifuged at 2500rpm for 10min. The deposit was washed again with D.W. the second deposit was used for sucrose flotation isolation procedures. The purified oocyst was incubated with antibiotics (penicillin 5000 Iu/ml, streptomycin 5 mg/ml and amphotericin-B 50 μ g/ml) at 37C^o for 12hrs to kill microbial contaminants. The number of the oocysts were counted using neubar slide chamber and the dimensions of the oocysts were measured by ocular micrometer and stage micrometer slide. The oocysts were stored in

aqueous K₂Cr₂O₇ (2.5% wt/vol) at 4-8C^o until used later [11].

Treatment experiments: For trials to treating *Cryptosporidium* parasite three medicinal plants were chosen which have been used for treating intestinal disorders and diarrhea, the plants was *Curcuma longa* rhizomes, *Coriandrum sativum* and *Viscum album* fruits. The plant parts were obtained from local markets in Kirkuk and the species were confirmed in traditional Plant Center (T.P.C.) in College of Science / Tikrit University. For extracting the plant component Prabhakar [12] procedures were used with some modifications.

Alcoholic plant extraction.

.Medicinal herbs were grounded by rotary grinder to yield a fine powder,10 gm of plant powder was weighted and placed in a maceration jar with 200ml of 95% ethanol mixed thoroughly.The jar was closed tightly to prevent evaporation of alcohol and was left in a dark place for 72hrs at room temperature, shacked tightly at least twice for 3 min each day.After 72hrs, the jar was opened, the extraction was filtered with gauze then with filter paper (Whatmann No. 1).The solvent was evaporated in water bath at 50-60C^o. the dry extracts percentage yield were calculated and stored at 4C^o until used, (table1), [12].

Watery plant extraction.

. 10gm of powdered plant were weighted, added to pyrex beaker containing 200ml D. W. . The mixtuer was heated for 2hr on a hot plate at 60C^o. Heat source was removed and the beaker cooled at room temperature. Solution was filtered, dried, stored as in alcoholic extracts above[12]....

In vivo screening of anti-*Cryptosporidium* effects of plant extracts.

.Seventy seven groups (plant extract treating groups + control groups) of 3 Balb/c mice (total number of mice used were 231) 4-8 weeks old 11-16g weight were administered orally with 10³ oocysts isolated from human feces. The mice feces were examined daily for oocysts recovery [13].The infection was recovered after 3-4 days post infection. The infected mice were treated with watery and alcoholic plant extracts at concentrations of 250,750,1000mg/kg twice daily each 12 hrs. for 7 days by using stomach tube. The oocysts number in feces of infected mice were counted daily by collecting and weighting 1gm feces and diluting it in to known volume of saline. A drop of this dilution (0.1ml) was smeared on a slide and stained as in [10] procedure. The number of the oocysts were counted in the 0.1ml of

the feces and the total number of the oocysts in 1gm of feces were calculated [14].

The feces examination was continued for three days after disappearance of oocysts in the feces to confirm the disappearance of the oocysts. For control, two groups of mice were used, one treated with an antibiotic (azithromycin) 10mg/kg as a positive control and the other not treated with plant extracts or with the antibiotic as a negative control. The synergetic effect of folic acid 0.34mg/kg and potassium chloride 5mg/kg with the watery and alcoholic plant extracts and with the antibiotic were examined too by daily feces oocysts counting.

Toxicity testing of plant extracts.

The toxic effect of plant extracts were studied by weighting the mice and weighting their livers and kidneys, noting the daily activity of the mice, and measuring the enzymes (GOT, GPT) and urea levels in the blood serum for liver and kidney dysfunction recovery. For this test mice which were used for treating experiments at highest concentration (1000 mg/kg), positive control and negative control were sacrificed. Their blood were collected using insulin needle 29 gauge by heart puncher. The blood was left to clot at 4C° then centrifuged for 10 min at 3000 rpm. The serum was gently separated from clotted blood for GOT, GPT, and urea measuring, the morphologies and weights of the liver and kidney were noted and recorded.

Serum GOT(Glutamic-oxalo acetic transaminase) and GPT (Glutamic-pyruvic transaminase) measuring: For GOT and GPT assay kit were used from Randox com.

Procedure: 0.1ml of the sera were pipette into test tubes (0.1ml D.W in Reagent blank tube), 0.5ml of phosphate buffer 100mmol/L, PH7.4 was added to each tube, mixed, incubated for exactly 30min at 37C°, 0.5ml of 2.4-dinitrophenylhydrazine 2.0mmol/L was added to each tube. Mixed, allowed to stand for exactly 20min at 20-25C°. 0.5ml of sodium hydroxide 4.0mol/L was added. Mixed, the absorbance of samples was read at 546nm against the blank. GPT and GOT levels were obtained from standard absorbance tables in u/ml.

Serum urea measuring: Serum urea was measured using kit from biolabo reagents com.

Procedure: 1ml of working reagent (salicylate 31mmol/L, nitroprussiate 1.67mmol/L and urease ≥ 15 Kul/L) were pipette into test tubes (Blank, standard, assay), 5 μ l of demineralised water was added into blank tube, 5 μ l of urea 40mg/dL (6.66mmol/L) into standard tube and 5 μ l of sera into assay tubes were added too. Mixed and waited for 4min at room temperature, 1ml of base solution sodium hypochlorite

7 mmol/L and sodium hydroxide 62mmol/L were added to each tube. Mixed, and let stand for 8min at room temperature. Absorbance were read at 600nm against blank.. Serum urea levels were calculated by this equation:

$$\text{Urea} = \frac{\text{Abs(Assay)}}{\text{Abs(Standard)}} \times$$

standard concentration.

Different statistical tests were used for analyzing the results according to the data : Analysis of variance for factorial experiment with two factors and (F) tests. χ^2 (chi-square) test in style of independent and in style of homogeneous. Duncans multiple –range test style of comparison between the levels of the factors. The level of significance used was P<0.05.

Results.

In vivo anti- *Cryptosporidium* effects of watery plant extracts:

For *in vivo* testing of plant extracts on *Cryptosporidium* oocyst production , watery plant extracts of three medical plants with three concentrations were used and the results(table 2) showed that watery extracts at 250 mg/kg was significantly effective for *Curcuma longa* and *Viscum album* comparing with the control but not significantly effective when compared with azithromycin at $p < 0.05$ level. All the extracts were dose related and their effects were higher at 750, 1000 mg/kg. The *Curcuma* was the most significantly effective agent than the other plants and its effect was equal to azithromycin at 750 mg/kg. The *Viscum* and *Coriandrum* were significantly effective comparing with control but not with the *Curcuma* and azithromycin. At concentration of 1000 mg/kg, the *C. longa* was the most effective agent and it was even significantly higher than azithromycin followed by *V. album* which had equal effect with azithromycin. The *Coriandrum* was significantly higher comparing with control but not with azithromycin.

..The day of treating was significantly effective in increasing the plants effect at all concentrations used. The most effective one was *Curcuma* which caused the disappearance of oocysts on the 7th day with rate of 100% at 750 mg/kg and on the 5th day with rate of 100% at 1000 mg/kg. While *Viscum* and *Coriandrum* were effective with rate of 54, 41% at 1000 mg/kg on the 7th day for each one respectively comparing with azithromycin and control.

In vivo anti- *Cryptosporidium* effect of alcoholic plant extracts :

There was no significant differences between the alcoholic (table 3) and watery (table 6) extracts, except for *Viscum* at all concentrations especially on the 7th day of treating with rate of 50,73,76% for 250, 750, 1000

mg/kg respectively comparing with 26, 48, 54% for the same concentrations in the watery extracts. The *Curcuma* was significantly higher than the two other plants at 250 mg/ml comparing with the control but not with azithromycin, followed by *Viscum* comparing with the control. The *Coriandrum* was not significantly effective at 250 mg/kg.

.At 750 mg/kg the *Curcuma* effect was equal with the effect of azithromycin comparing the two with control, followed by *Viscum* and *Coriandrum* comparing with control but not with azithromycin. The *Curcuma* had the highest effect at 1000 mg/kg compared with the other plants and with the azithromycin and control, followed by *Viscum* compared with the azithromycin and control. The *Coriandrum* was not significantly effective compared with azithromycin but not with control. The increasing of treatment period was significantly effective on decreasing of the oocysts seen in stool, for all the plants and the antibiotic compared with the control. The most effective was the *Curcuma* which caused the disappearance of the oocysts with rate of 100% on the 4th day of treatment at 1000 mg/kg, and a rate of 82% at 750 mg/kg on the 7th day followed by *Viscum* with rate of 73% at 750 mg/kg and 76% at 1000 mg/kg on the 7th day. And the lowest was for *Coriandrum* with rate of 21, 38% at 750, 1000 mg/kg on the 7th day of treatment.

In vivo anti- *Cryptosporidium* effect of watery and alcoholic plant extracts with folic acid:

The folic acid had no synergetic nor antagonistic effect with both the watery or alcoholic extracts (table 4, 5), the effect of both plant extracts or the antibiotic were significantly not effected with adding of folic acid.

In vivo anti-*Cryptosporidium* effect of watery and alcoholic plant extracts with potassium chloride:

. The effect of potassium chloride on watery and alcoholic plant extracts (table 6, 7) were not significantly different from that of watery or alcoholic extract alone. Except with *Curcuma* at 1000 mg/kg which led to significantly decreasing the potency of the plant, and had changed the day of disappearance of the oocysts from the 4th day in alcoholic plant extract alone to the 7th day with adding of potassium chloride.

In vivo anti-*Cryptosporidium* effect of watery and alcoholic plant extracts with folic acid and potassium chloride:

The effect of adding potassium chloride and folic acid together to the watery and alcoholic plant extracts (table8, 9) were not significantly effective in decreasing the stool oocyst numbers at all concentrations used and at all days of treatment, compared with the watery and alcoholic extracts alone. But it was significantly

effective with the azithromycin, they increased its affectivity. On the 4th day the effect was increased to 100% with both the watery and alcoholic extracts.

The toxic effect of the plant extracts:

. For determining the toxicity of the plant extracts the daily activity, animal weight, liver and kidney morphology and weight, and serum GPT, GOT and urea were observed in the treated group. The activity, feeding and weight 11-16 g of the mice under treatment were not affected comparing with control nor any mortality were seen among them. No morphological changes were seen on the livers and kidneys of the treated mice comparing with the control mice (fig.2). The weights of the livers and kidneys were not affected by plant extracts and it was 1.1-1.3 g for livers and 0.2-0.4 g for the two kidneys in both treated and control groups. The serum GPT, GOT, and urea measuring (table10) showed no significant differences between the GPT, GOT and urea levels in both treated and untreated mice compared with un infected control mice.

Discussion.

In vivo anti *Cryptosporidium* effect of watery plant extracts:

.The effect of the plant extracts on the number of oocysts produced by infected mice and the day of disappearance of this oocysts was depended on the dose and the days of treatment. This may because the amount of the effective component in low concentrations may be not enough to effect the parasite, the doses of treatment must be continued for several days in order their active component to act properly. Perrucci [15] found significant affectivity of *Mangiferin* at 250 mg/kg only after the end of treatment (10 days).

.*C. longa* was the most effective plant extracts on the parasite especially at 1000 mg/kg which cause the disappearance of the oocysts on the 5th day of treatment exceeding the effect of the azithromycin which is now used for treating the parasite. This was similar to Kadappu [5] results who improved that azithromycin treated mice had become asymptomatic after 7days of treatment, but stool samples was positive for *Cryptosporidium* even after 7days of therapy. Curcumin, a natural polyphenolic compound, has been found to be active against a variety of diseases including anti carcinogenic, antimicrobial and antiprotozoal effect [16,17].

. Curcumin inhibits cell proliferation and induces cell cycle changes in colon adenocarcinoma cell lines by a prostaglandin-independent pathway and may inhibit or stop the parasite proliferation and development, several

studies had confirmed the effect of *C. longa*. A study of chicks infected with the *Eimera maxima* demonstrated that diets supplemented with 1% turmeric resulted in a reduction in small intestinal lesion and improved weight gain [18]. Among two drugs and two plant extracts, used by Sinch [19] *C. longa* showed maximum vermifuge activity at the concentration of 50 µg/ml on *Pheretima pothuma* model. The curative effect of oil extract of *C. longa* on *Schistosoma mansoni* infected mice was recorded by El-Ansary [20], they showed that the *C. longa* was normalized the concentration of protein, glucose, AMP-deaminase and adenosine deaminase which were changed by the worm infection, moreover it was more potent in reducing egg count.

The effect of *C. sativum* and *V. album* were not high and maximum effect of them was 40,54% at 1000mg/kg on the 7th day of treating respectively, this may because their active components are normal substances like proteins, fatty acids, carbohydrates, and minerals [21]. This result is identical to several other results in trials for treating the parasite with different plant extracts each with some effect but any was not curative. The effect of *Punica granitium* was 53% at 1000 mg/kg, *Thymus vulgaris* was 50% at 1000 mg/kg [22]. Al-Alousi [13] had used five medicinal plants at 500mg/kg which their effects were ranged from 34.5% to 70.9% and not completely effected the parasite. Perucci [15] had showed reduction rate of 80% for *Mangiferin* at 1000 mg/kg. Not identical results was found by Al-Abaasi [23] who showed high effect of *Viscum album* at 1000 mg/kg on cutaneous leishmaniasis, Uma [16] showed high effect of *C. sativum* against infectious bacteria causing diarrhea at 4mg/ml, but Qadir [24] showed no significant effect of aqueous and alcoholic extracts of *C. sativum* at both high and low doses he used (0.45, 0.9 g/kg) on *Haemonchus contortus* nematode in infected sheep. Same results were observed by Eguale [25] who detected no significant effect of *C. sativum* on the same worm for both watery and alcoholic extracts on 7 and 14 days post treatment.

In vivo anti *Cryptosporidium* effect of alcoholic plant extracts:

No significant differences has been noted between the watery and alcoholic plant extracts and the results of alcoholic extracts on oocysts shedding in the laboratory infected mice were similar to that of watery extracts. This may because very closely related compounds has been extracts by the two solvents or due to extraction method which include heating in watery extracts and only maceration in alcoholic extracts which may have lead to extract the same active components for both

solvents, this result was identical to results showed by Sinch, Eguale [19, 25] whom showed no significant differences between the watery and alcoholic extracts of *C. sativum* and *C. long*. Although the alcoholic extracts showed no significant difference with watery extracts but data showed that the alcoholic extracts were more effective with the *V. album* especially on the 7th day with rate of 50,73,76% for 250,750, 1000 mg/kg respectively, comparing with 26, 48, 54% for the same concentrations respectively in watery extracts. This agree with Al-Abbasi [23] who extracted 11 compounds in alcoholic extracts of *V. album* versus 9 compounds in watery extracts.

The effect of folic acid and potassium chloride on the plant extractes:

Folic acid, a water-soluble vitamin of the B-complex group when supplemented with the plant extracts had no significant effect on oocysts shedding per days of treatment. Same result was indicated by Khanna [26] who showed very little synergistic effect of folic acid when used with two medicinal plant extracts (*Ocimum sanctum*, *Commiphora mukul*) on lipid peroxidation in experimentally-induced hyperlipidemia mice. No significant effect was noted with using of potassium chloride with all plant extracts and azithromycin except with *C. longa* which decreased its effect, this may because of that the potassium chloride is interfere with the active components of the plants leading to reduction in its activity.

The using of potassium chloride with folic acid again had no significant effect with all plant extracts but they increased the azithromycin activity, this agree with that found by Al-Jarjary [27] in which she referred to the synergetic effect of vitamin E with azithromycin and spriamycin.

The toxic effect of the plant extracts.

The watery and alcoholic plant extracts had no toxic effect, the activity and weight of the treated mice were not affected comparing with control, this in agreement with Certad and Theodos[14, 28] whom not had recorded significant body weight change in mice infected with *Cryptosporidium*.

The weight or morphologic changes were not seen in the livers and kidneys of the treated mice, nor the serum GPT, GOT and urea were effected at the concentrations of the extracts which were used.

For the *Coriandrum* and *Curcuma* no toxic effect had been recorded by the previous studies [29, 30] but in *Viscum album* little toxicity was recorded in very high concentrations Al-Abbasi [23] had recorded 3000 mg/kg

as LD50 and Nawaygerae [31] had recorded 1525 mg/kg as LD50.

References.

- 1- Griffiths JK, Balakrishnan R, Widmer G. and Tzipori S. (1998). Pramomycin and geneticin inhibit intracellular *C. parvum* without trafficking through the host cell cytoplasm: implications for drug delivery. *Infect. Immune.* 66: 3874-83.
- 2- Flanigan T.P. and Soave R.(1993). cryptosporidiosis. *Prog. Clin. Parasitol.*1-20.
- 3- Goodgame RW. (1996). Understanding intestinal spore-forming protozoa: Cryptosporidia, Microsporidia, Isospora and Cyclospora. *Ann Intern Med.* 124:4 429-41.
- 4- Giacometti A, Cirioni O, Barchiesi F and others. (2000). Activity of Nitazoxanide alone and in combination with azithromycin and rifabutin against *C. parvum* in cell culture. *J. Anti-microb. Chemotherapy.* 45:4 453-456.
- 5- Kadappu KK, Nagaraja MV, Rao PV and Shastry BA.(2002). Azithromycin as treatment for Cryptosporidiosis in human immunodeficiency virus disease. *Dep. Med. Karhataka India* 48(3): 179-181.
- 6- Woods KM, Nesterenko M N and Upton SJ. (1996). Efficacy of 101 antimicrobials and other agents on the development of *C. parvum* in vitro. *Ann. Trop. Med. Parasitol.* 90:6 603-615.
- 7- Fujikawa H, Miyakawa H, Ignchi K, Nishizawa M and others.(2002). Intestinal Cryptosporidiosis as an initial manifestation in previously healthy Japanese patients with AIDS. *Gastroenterogy*, 37: 840-843.
- 8- Rueda, G, Fenoy S, Simon F, and Aguila C.(2008). Bobel-24-Activity against *C. parvum* in cell culture and in a SCID Mouse Model. *Anti- microb. Chemoth.* 52:3 1150-1152.
- 9- Klein P, Cirioni O, Giacometti A and Scalise G.(2008). In vitro and in vivo activity of aurintricarboxylic acid preparations against *C. parvum*. *J. Anti-microb. Chemoth.* 62:5 1101-1104.
- 10-Henriksen SA and Pohlenz JFL.(1981). Staining of *Cryptosporidium* by a Modified Ziehl-Nelson technique. *Acta. Vet. Scand.* 22: 594-96.
- 11-Arrowood M J.(2002). In vitro cultivation of *Cryptosporidium* species. *Clin. Microbial. Rev.* 15:3 390-400.
- 12-Prabhakar B, Uma K, Rajendran S and Sarayu LY.(2009).Antimicrobial Activity and phytochemical analysis of *Coriander sativum* against infectious diarrhea. *Ethno botanical leaflets* 13: 590-94.
- 13-Al-alousi TI.(2004). Prevalence of *Cryptosporidium spp.* In different resources with a trial treatment by using medical plant extracts. Ph. D. Thesis, Coll. Med. Tikrit. Univ.
- 14-Certed G, Ngouanesavauh T, Guyot K, Gantois N and others.(2007). *Cryptosporidium parvum*, a potential cause of colic adeno carcinoma. *Infec. Age. and Can.* 21.
- 15-Perrucci S, Fichi G, Buggiani C, Rossi G and Flamini G.(2006). Efficacy of manigiferin against *Cryptosporidium parvum* in a neonatal mouse model. *J.Para.Res.*99:2 184-188.
- 16-Uma B, Prabhakar K, Rajendran S and sarayu YL.(2009). Antimicrobial activity and phytochemical analysis of *Coriander sativum* against infectious diarrhea. *Ethno botanical leaf tets.* 13: 590-94.
- 17-Shahiduzzaman M, Dyachenko V, Khalafalla RE, Desouky AY and Dausgies A.(2009). Effects of curcumin on *Cryptosporidium parvum* in vitro. *J.Para.Res.*105:4 1155-61.
- 18-Allen PC, Danforth HD, Augustine PC.(1998). Dietary modulation of avian coccidiosis. *Int. J. parasitol.* 28: 1131-1140.
- 19-Sinch R, Meh TA A, Meh TA P, Shukla K.(2011). Antihelminthic activity of rhizome extracts of *Curcuma longa* and *Zingibar officinale*(*Zingiberaceae*). *Int. j. phar. & phar. Scie.* 3: 2 236-237.
- 20-El-Ansary A, Ahmed SA, Aly S A.(2007).Antischistosomal and liver protective effects of *Curcuma longa* extract in *Schistosoma mansoni* infected mice. *Indian J.Experi.Biolo.*45: 791-801.
- 21-Fraqiska M.(2005). Wild and cultivated vegetables, Herbs and spices in Greek Antiquity. *Envi. Archaeology* 10:1 73-82.
- 22-Al-Rifay O M S.(2006). Comparison between the diagnosis of Cryptosporidiosis by using the Eliza assay and the modified carbol fuchsin stain with the tese to discover the effect of the certain plant extracts

on the parasite. M. S. thesis. Tikrit .University. Arabic reference.

23-Al-Abbasi M LT.(2009). Comparison of watery and alcoholic extracts of *Artemisia aherba –alba* and *Viscum album* on cutenious leishmaniasis *in vitro* and *in vivo*. M. S. thesis. Tikrit University. Arabic reference.

24-Qadir S, Dixit AK and Dixit p.(2010).Use of medicinal plants to control *Haemonchus contortus* infection in small ruminants. *vet. wo.*3:11 515-518.

25-Eguale T, Tilhun G, Debella A, Feleke A and Makannen E.(2007). *In vitro* and *in vivo* antihelmintic activity of crude extracts of *Coriandrum sativum* against *Haemanchus contortus*. *J. Ethnopharmacol.* 110:3 428-33.

26-Khanna, N, Arora, D, Halder S and others. (2010). Comparative effect of Ocimum sanctam, Commiphora mukul, Folic acid and ramiprilon lipid peroxidatiion in experimentally-induced hyperlipidemia. *Indian . J. E. Bio.* 48: 299-305.

27-Al-Jarjary SA .(2006).Trials on treatment of Cryptosporidiosis in Balb/c mice. PH. D. thesis Mosul University. Arabic reference

28-Theodos, CM, Griffiths J K, D'onfro J and others.(1998). Efficacy of Nitazoxanide against *C. parvum* in cell culture and in animal models. *Am. Soc. Microbial.* 42:8 1959-1965.

29-Khanna S, Park HA, Sen C and others.(2009). Neuroprotective and Anti-inflammatory properties of a noves demethylated curcuminoid. *Antioxi. & Rewdox singaling.* 449-468.

30-Dawakhana H.(2007). Coriandrum: Cure from the kitchen, hashmi.com. <http://www.hashmi.com/coriander.html>.

31-Nawaygerae and others. (2000) . cited by Al-Abasi . (2009) .

<i>Viscum album</i>	0.51	5.1	0.58	5.8
<i>Coriandru m sativum</i>	0.88	8.8	1.22	12.2

Table(2):The means of oocyst numbers after different periods from giving the watery plant extracts.

Control without treatment	azithro mycin	Viscum album		Curcuma longa		Coriandrum sativum		Watery Plant Extract								
		means of oocyst numbers / days		means of oocyst numbers / days		means of oocyst numbers / days		means of oocyst numbers / days								
		05L	05C	0001	05L	05C	0001	05L	05C	1	2	3	4	5	6	7
5444	10	1000	5229	5048	5589	5511	5860	5038	5795	0	0	0	0	0	0	0
6154	16	7	4	8	0	0	0	7	0	6792	6560	6493	6231	6231	6231	6231
8587	36	20	2	51	23	8	0	0	0	5330	5095	6887	6170	6170	6170	6170
8791	2374	4858	6206	2150	4020	5608	5330	5095	6887	38	34	19	6170	6170	6170	6170
7850	72	43	28	75	53	33	38	34	19	3124	5310	6170	6170	6170	6170	6170
5029	65	54	32	97	65	55	65	40	30	65	40	30	6170	6170	6170	6170
5222	2779	3304	4019	0	1899	4010	3344	4889	6231	3344	4889	6231	6231	6231	6231	6231
	65	58	49	100	76	50	57	38	21	57	38	21	6231	6231	6231	6231
	2182	2360	4020	0	339	4100	3243	4049	5901	3243	4049	5901	5901	5901	5901	5901
	57	53	20	100	93	19	36	20	0	36	20	0	5901	5901	5901	5901
	1988	2388	3872	0	0	4002	3090	3870	5008	3090	3870	5008	5008	5008	5008	5008
	62	54	26	100	100	23	41	26	4	41	26	4	5008	5008	5008	5008

Table (1) Plant extracts weights yield with percentages.

Plant used	yield in gm. In alcoholic	Percentage %	yield in gm. In watery	Percentage %
<i>Curcum a longa</i>	0.63	6.3	1.56	15.6

Control without treatment	Azithromycin	Viscum album			Curcuma longa		
		1000	750	250	1000	750	250
.....	10	1000	750	250	1000	750	250
5444	4371	4562	4727	4799	3551	4551	4733
6154	20	16	13	11	35	16	13
	3583	4070	4401	4661	2878	4264	4892
	42	34	28	24	53	31	21
8587	3121	5234	5260	5312	1175	5010	5335
	64	39	39	38	86	42	38
8791	2665	4651	4909	4930	98	3792	5294
	70	47	44	44	99	57	40
7850	2892	3963	4216	4329	0	2421	4567
	63	50	46	44	100	69	42
5029	1924	2887	3254	3535	0	1017	3001
	62	43	35	30	100	79	41
5222	1948	1422	2992	3511	0	953	2678
	63	72	43	33	100	81	49

Table (6) : The means of oocyst numbers after different period from giving the watery plant extracts with potassium chloride.

Curcuma longa	Coriandrum sativum			Watery plant extract+p-otassium chloride 5 mg/kg	
	750	250	1000	750	250
1000	750	250	1000	750	250
4115	4431	4966	5360	5593	5651
24	19	9	2	0	0
4092	4290	4550	5960	6151	6544
34	30	26	3	0	0
3106	3602	5803	5363	6261	6443
64	58	32	38	27	25
1901	3125	5221	5294	5469	5991
78	64	41	40	38	32
1497	2290	4604	4430	4878	6232
81	71	41	44	38	21
316	1421	3112	3581	4372	4339
94	72	38	31	16	14
0	1119	3110	4203	4291	4872
100	79	40	16	15	7

Control without treatment	Azithromycin	Viscum album		
		1000	750	250
.....	10	1000	750	250
5444	4972	4900	5122	5663
6154	9	10	6	0
	3677	4560	5232	5982
	40	26	15	3
8587	3104	5501	5914	5916
	64	36	31	31
8791	2842	4905	5445	5675
	68	44	38	35
7850	2525	4197	5201	5116
	68	47	34	35
5029	2466	2908	3459	3513
	51	42	31	30
5222	2431	2725	3457	3513
	54	48	34	30

Table (7):The means of oocyst numbers after different period from giving the alcoholic plant extracts with potassium chloride.

Viscum album	Curcuma longa			Coriandrum sativum		Alcoholic Plant extract+ Potassium chloride 5mg/kg	
	750	250	1000	750	250	Con. mg/kg	Means of oocyst numbers/ days
750	250	1000	750	250	1000	1	
5013	5020	3951	3970	4151	5009	%	
8	7	27	27	24	8	2	
5592	5722	3106	4205	4290	5107	%	
9	7	50	32	30	17	3	
4800	5677	3101	4120	5018	5339	%	
44	34	64	52	42	38	4	
4451	4960	2662	4055	5016	5239	%	
49	44	70	54	43	40	5	
3096	4102	2001	3163	4072	4993	%	
61	48	75	53	48	36	6	
2075	3449	900	2802	3115	3223	%	
59	32	82	51	38	36	7	
2203	3331	0	1755	3027	3260	%	
58	36	100	66	42	38	28	

Control without treatment	Azithromycin	1000
.....	10	1000
5444	4972	4977
	9	9
6154	3677	4876
	40	21
8587	3104	4530
	64	47
8791	2842	4317
	68	51
7850	2525	3152
	68	60
5029	2466	2069
	51	59
5222	2431	1659
	54	68

Table (8) :The means of oocyst numbers after different period from giving the watery plant extracts with folic acid and potassium chloride.

Viscum album	Curcuma longa						Coriandrum sativum						Watery Plant extract+Folic acid 0.34mg/kg + Potassium-chloride 5mg/kg Con. mg/kg
	750		250		1000		750		250		5000		
	5102	5210	3173	4117	2204	2204	3173	4117	5000	5211	5201	5211	
11	6	5	42	24	8	5	5	5	5	5	5	5	
4639	4717	5100	3204	4603	5215	5236	5722	5215	5236	5722	5236	5722	
25	23	17	48	25	15	15	7	15	15	7	15	7	
4345	4595	4920	2224	4962	5620	5870	5922	5620	5870	5922	5870	5922	
49	46	43	88	74	35	32	31	35	32	31	32	31	
4109	4351	4781	182	4201	4306	4405	4794	4306	4405	4794	4405	4794	
53	51	46	98	52	51	48	45	51	48	45	48	45	
2855	3605	3825	0	2580	4391	4561	4887	4391	4561	4887	4561	4887	
64	54	51	100	67	44	42	38	44	42	38	42	38	
2822	2991	3199	0	1898	3588	3911	3980	3588	3911	3980	3911	3980	
44	41	36	100	62	29	22	21	29	22	21	22	21	
973	1142	2360	0	1802	3127	3373	3562	3127	3373	3562	3373	3562	
81	78	54	100	65	40	35	32	40	35	32	35	32	

Control without treatment	Azithromycin	10
.....	10	10
5444	2302	2302
	58	58
6154	1114	1114
	82	82
8587	989	989
	89	89
8791	0	0
	100	100
7850	0	0
	100	100
5029	0	0
	100	100
5222	0	0
	100	100

Table (9):The means of oocyst numbers after different period from giving the..alcoholic plant extracts with folic acid and potassium chloride.

Azithromycin	Viscum album						Curcuma longa						Coriandrum sativum						Alcoholic Plant extract+Folic acid 0.34mg/kg +Potassium-chloride 5mg/kg Con. mg/kg
	1000		750		250		1000		750		250		1000		750		250		
	4390	4505	4662	2932	4105	4420	4870	4801	5216	4870	4801	5216	4870	4801	5216	4870	4801	5216	
58	19	17	15	46	19	11	12	4	11	12	4	11	12	4	11	12	4	11	
1114	4014	4761	5139	2701	4117	4738	5519	5605	4117	4738	5519	5605	4117	4738	5519	5605	4117	4738	
82	35	23	16	59	33	23	10	9	33	33	10	9	33	33	10	9	33	33	
989	3209	3895	4670	1478	3449	4481	5555	5555	3449	4481	5555	5555	3449	4481	5555	5555	3449	4481	
89	63	55	46	82	59	48	35	35	59	48	35	35	59	48	35	35	59	48	
0	3408	3523	4506	204	3264	4452	5499	5499	3264	4452	5499	5499	3264	4452	5499	5499	3264	4452	
100	61	60	48	97	62	49	37	37	62	49	37	37	62	49	37	37	62	49	
0	1714	3048	4371	0	1659	4107	5185	5185	1659	4107	5185	5185	1659	4107	5185	5185	1659	4107	
100	78	61	44	100	78	48	34	34	78	48	34	34	78	48	34	34	78	48	
0	982	1804	3235	0	984	2896	4306	4306	984	2896	4306	4306	984	2896	4306	4306	984	2896	
100	80	64	36	100	80	42	21	21	80	42	21	21	80	42	21	21	80	42	
0	0	1035	2186	0	0	2248	2603	2603	0	2248	2603	2603	0	2248	2603	2603	0	2248	
100	100	80	58	100	57	48	26	26	57	48	26	26	57	48	26	26	57	48	

Control without treatment	5444	6154	8587	8791	7850	5029	5222
---------------------------	-------	------	------	------	------	------	------	------

Table (10) The toxic effect of plant extracts on the livers and kidneys function in the mice under treating.

Plant type	Extract	GPT Unit/ml	GOT Unit/ml	Urea Mg/dl
<i>Curcuma longa</i>	Alcoholic	28.6 ±2.3	36.5±1.7	3.5±0.05
	Watery	28.8±2.9	36.5±1.9	3.4±0.024
<i>Coriandrum sativum</i>	Alcoholic	29.0±2	36.5±1.2	3.5±0.03
	Watery	29.0±3	36.8±0.8	3.3±0.094
<i>Viscum album</i>	Alcoholic	28.3±2	36.6±0.8	3.5±0.03
	Watery	28.8±2	36.9±2.6	3.4±0.31
Control without plant extracts	----- -	29.2±1.3	36.5±2.5	3.4±0.095
Control without plant extracts without parasite	----- -	29.1±2.2	37.0±0.9	3.4±0.11

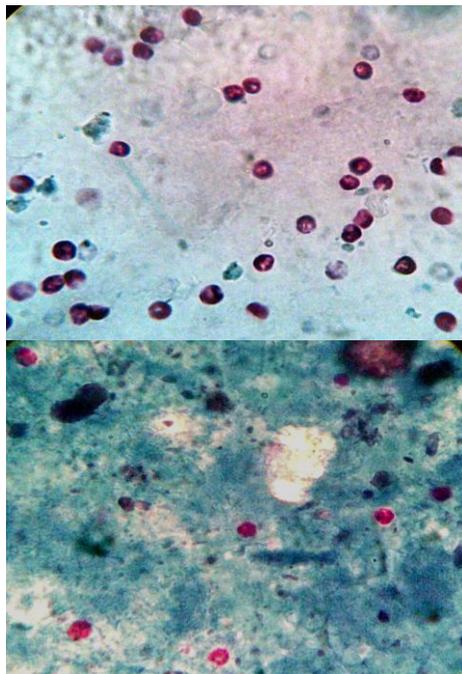
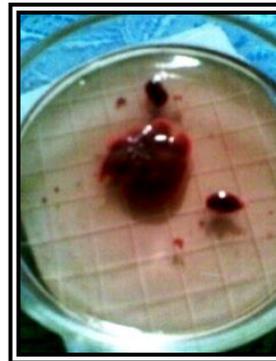


Fig.(1) Red or pink *Cryptosporidium* oocysts in human isolates stained by M.Z.N.1000x.



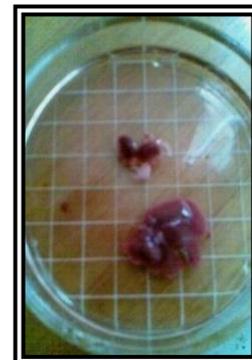
A



B



C



D

Fig. (2) Normal liver and kidney appearance of mice treated with the plants extracts: A (un treated) B, C, D (treated)

تأثير بعض مستخلصات النباتات الطبية على طفيلي داء البويغات الخبيثة في الحي.

هيرو محمد عبيد , توفيق ابراهيم الالوسي , عبدالله حسين الجبوري.

الخلاصة .

يعرف داء البويغات الخبيثة بأنه طفيلي يصيب الانسان و خاصة الاطفال من كلا الفئتين ذوي المناعة الطبيعية و ذوي المناعة القليلة . تأثير الاسهال المتسبب عن الطفيلي و فقدان السوائل الجسمية اثناء الاصابة قد يكون خطيرا جدا خاصة في الفئات العمرية الصغيرة و الاشخاص ذوي المناعة القليلة . صممت هذه الدراسة كمحاولة لأيجاد علاج للداء باستخدام المستخلصات الكحولية و المائية لبعض النباتات الطبية . لم يكن هناك فروق معنوية بين المستخلصات الكحولية و المائية للنباتات الثلاثة المستخدمة (الكركم ، الكزبرة ، الدبق)، وكل المستخلصات المستخدمة كانت ذو علاقة طردية مع التركيز حيث ازدادت تأثيراتها بزيادة التركيز . كان لنبات الكركم التأثير الاقوى لتقليل معدل الاكياس المطروحة في براز الفئران المعالجة نوع Balb/c, بمعدل تأثير 100% في اليوم السابع من العلاج عند تركيز 750 ملغم/كغم و اليوم الرابع عند تركيز 1000 ملغم/كغم في المستخلص المائي و معدل 100% في اليوم الرابع من العلاج عند تركيز 1000 ملغم/كغم مع المستخلص الكحولي. الدبق كانت في المرتبة الثانية بمعدل تأثير 54,48% في اليوم السابع عند تركيز 750, 1000 ملغم/كغم على التوالي للمستخلص المائي, ومعدل تأثير 76,73 % في اليوم السابع لنفس التركيزين السابقين على التوالي للمستخلص الكحولي. نبات الكزبرة كانت لها التأثير الاقل عند كل التراكيز المستخدمة ومع كلا المستخلصين الكحولي والمائي. لم يلاحظ فروق معنوية عند اضافة حامض الفوليك او كلوريد البوتاسيوم معا او بصورة منفصلة الى المستخلصات النباتية ماعدا عند اضافتهما معا الى المضاد الحيوي ازثرومايسين حيث سببا زيادة في تأثيره الى نسبة 100% في اليوم الرابع من العلاج بعد ان كان نسبة تأثيره 68% فقط دون المادتين .