











Organic Modification of Chitosan: An Overview

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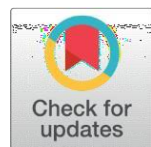
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ABSTRACT

Alkaline deacetylation of chitin yields chitosan, a naturally occurring polymer with remarkable biological qualities like immunological, antibacterial, and Wound healing capabilities. Recently, there has been an increase in interest in modifying chitosan chemically to increase its solubility and expand its uses. The usual method for obtaining chitosan is to deacetylate chitin in an alkaline environment. Chitin is one of the most common organic molecules, generated by biosynthesis second only to cellulose yearly. Chitin plays a significant role in the exoskeleton of animals, particularly insects, mollusks, and crustaceans. It is also the main fibrillar polymer found in some fungi's cell walls. Composed of glucosamine and N-acetyl glucosamine units connected by β (1–4) glycosidic linkages, chitosan is a linear polysaccharide. The degree of deacetylation refers to the amount of glucosamine present. Actually, in a broad sense. This study reviews the major chitosan chemical changes that have been suggested in the literature. Furthermore, a vast variety of derivatives with a wide range of uses are produced by these chemical alterations. Instances of recent and pertinent instances of the various uses are provided, with a focus on tissue engineering, medication delivery, and environmental applications. Looking ahead, the future of chitosan modification appears bright, driven by advancements in smart polymers, green chemistry, and nanotechnology. These developments are facilitating the creation of tailor-made materials with diverse applications, further expanding the potential utility of chitosan in various fields.

Keywords: Chitosan, Organic modification, Graft copolymerization, Biodegradability

INTRODUCTION:

Interesting characteristics of chitosan include biocompatibility and biodegradability¹, as well as non-toxic, non-immunogenic, and non-carcinogenic breakdown products^{4,5}. Chitosan hence has potential uses in a wider range of industries, including flocculation, waste water treatment, biomedicine, and functional membranes. However, the limited solubility of chitosan in weak acid solutions restricts its potential uses¹⁻⁶. The usual method for obtaining chitosan is to deacetylate chitin in an alkaline environment. Chitin is one of the most common organic molecules, generated by biosynthesis second only to cellulose yearly. Chitin plays a significant role in the exoskeleton of animals, particularly insects, mollusks, and crustaceans. It is also the main fibrillar polymer found in some fungi's cell walls. Chitosan is a linear polysaccharide consisting of glucosamine and N-acetyl glucosamine units connected by β (1–4) glycosidic linkages, as seen in Figure 1. The degree of deacetylation refers to the amount of glucosamine present. In fact, chitin is generally thought to become soluble in an aqueous acidic media when its deacetylation level exceeds around 50%^{2,4}. To increase chitosan's solubility and expand its uses, there has been a rise in interest in chemically modifying the material recently⁵. Chitosan's solubility at neutral and alkaline pH values can be significantly increased by derivatization, which involves adding tiny functional groups to the chitosan structure, such as carboxymethyl or alkyl groups without compromising the cationic character of the substance. Polymers having polyampholytic characteristics can be produced by substituting moieties containing carboxylic groups. Graft copolymerization has been the most popular modifying technique. Through the covalent attachment of a molecule, known as the graft, to the chitosan backbone, chitosan grafting enables the creation of functional derivatives^{7,8}. This study will explain the primary techniques that have been utilized to chemically alter chitosan. There will be a few examples of the chemical reactions and experimental setups that are indicative of the field. Lastly, the most significant uses of these modified chitosan-based polymers in various industries are also covered.

Chemistry of Chitosan

Deacetylated chitin is a naturally occurring that is converted into chitosan. A linear polysaccharide consists of β -(1→4)-linked N-acetylglucosamine (GlcNAc) and glucosamine (GlcN) units⁹. Reactive functional groups, mainly amino (NH₂) and hydroxyl (OH) moieties, are dispersed throughout the polymer backbone of the compound in its chemical structure as shown in Figure 1. These functional groups operate as important sites for organic alterations and give chitosan its distinct chemical reactivity¹⁰.

Amine Groups (NH₂): The glucosamine units of chitosan include the main amino groups, which give the polymer a basic character. A variety of changes are possible due to the amino groups' easy participation in nucleophilic substitution processes. The amine groups in chitosan are very reactive, which makes it possible to introduce a variety of functional entities including Schiff bases, amides, and esters¹¹.

Hydroxyl groups (OH): The hydroxyl groups that come from the GlcNAc and GlcN units help to make chitosan hydrophilic overall. These hydroxyl groups participate in hydrogen bonding interactions, especially those on the C-6 position of the glucosamine units. Furthermore, the hydroxyl groups have the ability to change chemically, introducing new functional groups like ethers and esters¹².

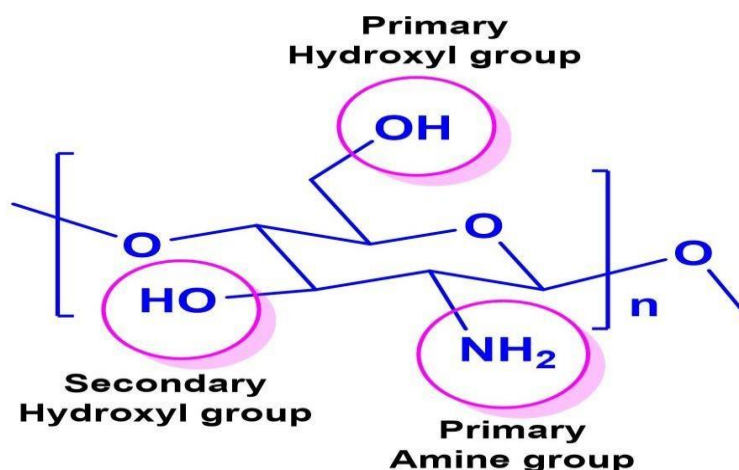
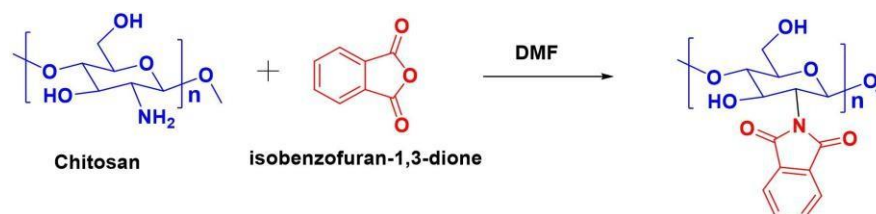


Figure 1 Structural representation of chitosan with active functional groups.

Chemical Reactivity and Protecting Groups

Chitosan's chemical reactivity results from the presence of reactive hydroxyl groups and nucleophilic amino groups. The production of amides and esters results from the interactions of the amino groups, which are nucleophiles, with electrophiles such as acylating agents. On the other hand, the hydroxyl groups can react with other substances to generate ethers, esters, and other derivatives¹³. N-phthaloyl-chitosan, an essential soluble N-protected precursor for further controlled modification processes of chitosan, may be produced using a straightforward and practical process for chemoselectively protecting the amino groups of chitosan. While partial hydroxy group phthaloylation was a side consequence of typical N-phthaloylation of chitosan in N,N-dimethylformamide, the O-phthaloylation was successfully reduced by the addition of a tiny amount of hydroxy-containing substances¹⁴. Water was one of the most appropriate chemicals to study; as a consequence, chemoselectively N-phthaloylated chitosan was formed, devoid of any noticeable O-phthaloyl groups. The resultant N-phthaloyl-chitosan had a bulky substituent, yet it was nevertheless found to be crystalline. According to a solubility test, N-phthaloyl-chitosan showed a strong affinity for organic solvents as shown in Scheme 1¹⁵.



Scheme 1: Protection group for the selective reaction of hydroxyl groups ¹⁴.

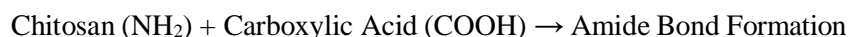
METHODS OF MODIFICATION

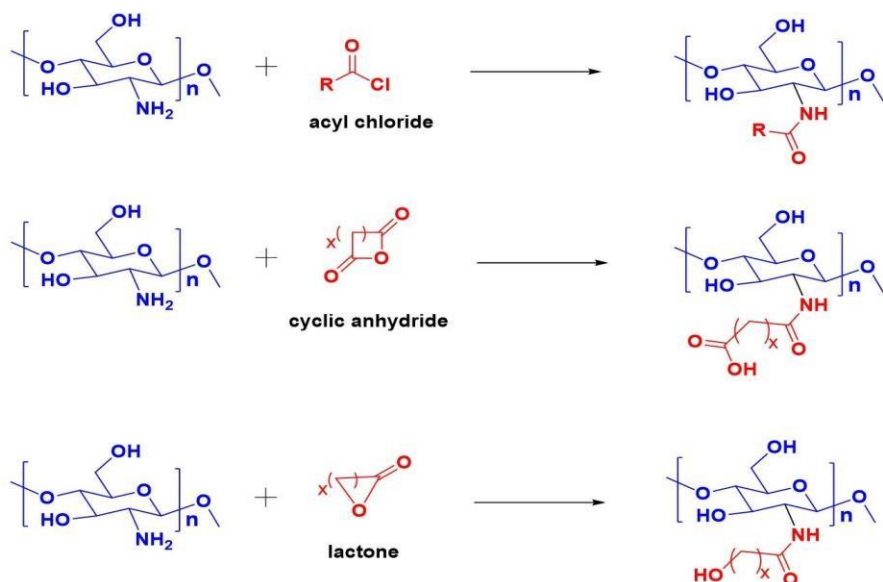
Chitosan's modification is significantly enhanced through various methods of attaching organic molecules. This allows researchers to tailor its properties for specific applications ¹⁶.

Acylation Reactions: The process of acylation reactions includes adding acyl groups to chitosan, usually by means of interactions with carboxylic acids, anhydrides, or acid chlorides as shown in Scheme 2¹⁷. Commonly, acylation processes result in the formation of esters and amides. These changes affect the solubility, biodegradability, and interaction properties of chitosan.

Schiff Base Formation: The condensation process between the amino groups in chitosan and aldehydes or ketones form Schiff bases. The azomethine (-C=N-) connections that Schiff bases introduce to increase the uses of chitosan in bioconjugation, drug administration, and sensing¹⁸ as shown in Scheme 3.

Amidation Coupling: Amide coupling involves the activation of carboxylic acid groups, typically facilitated by carbodiimide coupling agents, to enable their reaction with amino groups present in chitosan¹⁹. This process leads to the formation of amide bonds, resulting in the introduction of functional groups, as depicted in Scheme 4. The general reaction scheme for amidation coupling can be represented as follows:

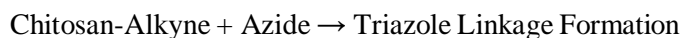




Scheme 2: Chitosan acylation with different acylating groups ¹⁷.

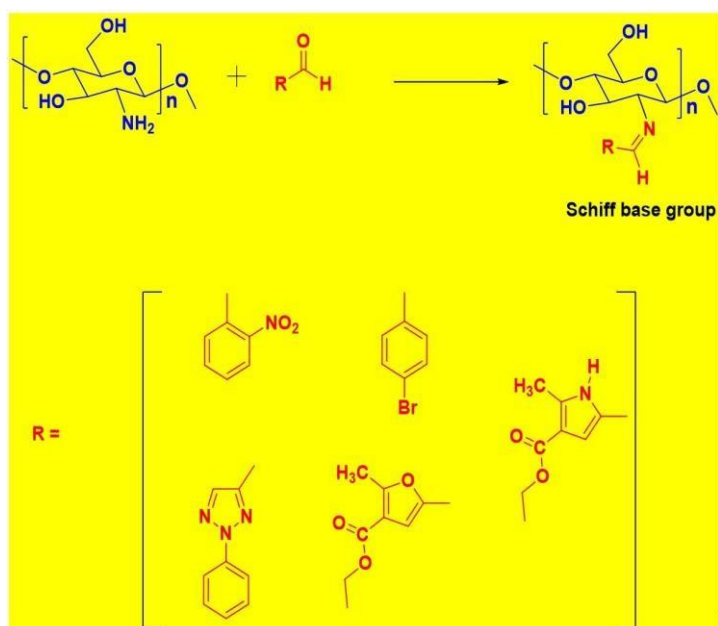
Carbodiimide coupling agents, such as N,N'-dicyclohexylcarbodiimide (DCC) or 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC), are commonly used to activate the carboxylic acid groups, facilitating their reaction with the amino groups of chitosan [20].

Click Chemistry: For the modification of chitosan, click chemistry—in particular, copper-catalyzed azide-alkyne cycloaddition, or CuAAC—offers a bioorthogonal method^{21,22}. Introduction of Functional Groups: Triazole linkages are created, which allow for exact control over the level of alteration while maintaining the original characteristics of chitosan as shown in Scheme 5²³. The general reaction scheme for CuAAC can be represented as follows:

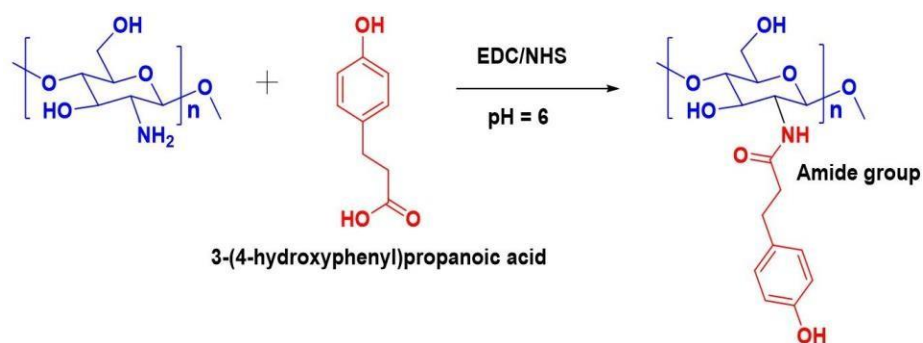


Enzymatic Modification: Using enzymes like lipases or transaminases to catalyze certain reactions on chitosan is known as an enzymatic technique. Introduced Functional Groups²⁴: The regioselective insertion of functional groups made possible by enzymatic modification minimizes unfavorable side effects and maintains the structural integrity of chitosan as shown in Figure 2²⁵.

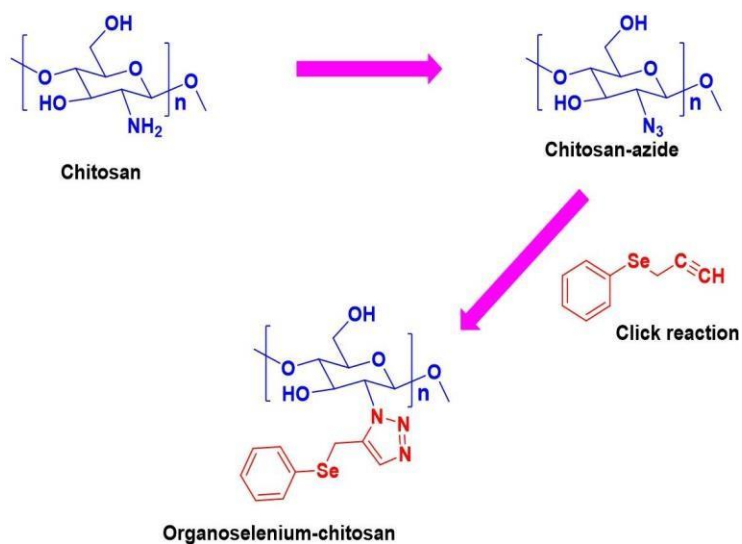
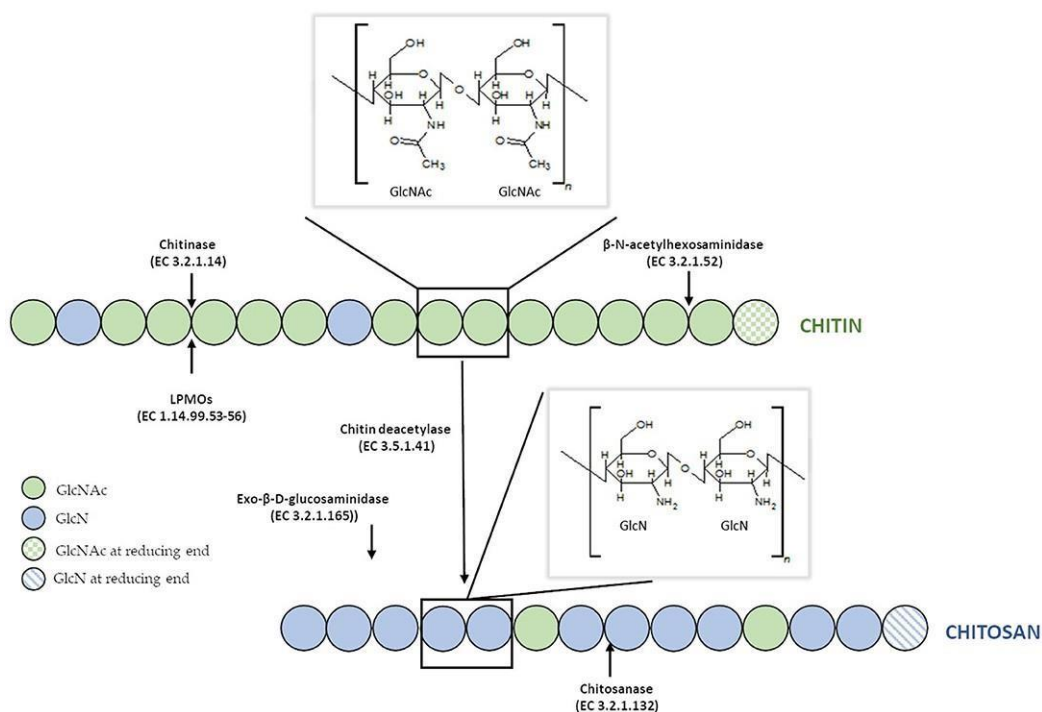
Polymer-Polymer Conjugation: There are several ways to conjugate chitosan with other polymers, such as mixing and graft copolymerization. Introduction of Functional Groups: By introducing various functionalities from the second polymer, this technique



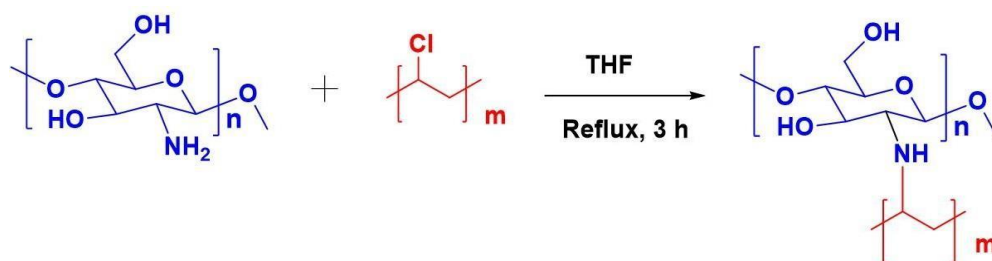
Scheme 3: Reaction route of synthesis Schiff base group with amine group of chitosan¹⁸.



Scheme 4: Chitosan amide formation with acid group²⁰.

Scheme 5: Click reaction of chitosan²³.Figure 2 Enzymatic modification of chitosan²⁵.

improves the mechanical, thermal, and biological characteristics of chitosan²⁶. Chitosan's adaptability is enhanced by polymer-polymer connections, which go beyond the addition of functional groups and enable the development of complex materials with specific characteristics. Through a variety of techniques, chitosan may be integrated with other polymers to improve its mechanical, thermal, and biological properties²⁷. By grafting polymer chains onto the chitosan backbone, a hybrid material is produced by graft copolymerization as shown in Scheme 6²⁸. Grafting can be accomplished by enzymatic or free radical polymerization processes. Different functions are added by grafted polymers, which change the mechanical strength, thermal stability, and material compatibility of chitosan. This approach finds use in the creation of functional coatings, tissue engineering, and drug delivery materials²⁹. After the preparation of the poly(vinyl chloride) (PVC)-chitosan thin films, the effect of filling nanoparticles on the lattice was examined²⁸. Tetrahydrofuran (THF) was used to dissolve PVC and 15% nano-chitosan in order to create thin films through the casting process. Using a diffusive reflectance apparatus, the optical characteristics of the thin films were measured in the approximate wavelength range of 250–1300 nm. In addition, calculations were made for the dielectric constant, Urbach energy, transmittance, reflectance, refractive index, absorption, optical conductivity, and skin depth. Both plain and NPs-filled thin film absorbance various values³⁰.



Scheme 6: Polymer-polymer modification of chitosan²⁸.

Functional Groups and Their Introduction

Mainly the modification of chitosan by adding several functional groups to its chemical structures. This gives the biopolymer specific characteristics. Here, we examine the insertion of important functional groups into chitosan by their synthesis³¹.

Synthesis Ester: When chitosan combines with derivatives of carboxylic acids, such as acid chlorides, anhydrides, or carboxylic acids, esters are frequently introduced through acylation reactions. Effect on Chitosan, the hydrophobicity, solubility in organic solvents, and film-forming properties of chitosan are improved by esters. Applications for modified chitosan esters include controlled release systems, medication administration, and wound healing³².

Synthesis Amides: When the amino groups in chitosan combine with the carboxylic

acid groups in a carbodiimide-mediated coupling process, amide groups result. Effect on the Chitosan, improvements in mechanical qualities, stability, and biocompatibility are all facilitated by amides. Biomedical applications such as medication delivery and tissue engineering frequently employ chitosan amides ³³.

Synthesis Schiff Bases: The amino groups in chitosan condense with aldehydes or ketones to generate Schiff bases. Effect on Chitosan, Schiff bases form azomethine linkages, which improve chitosan's reactivity to outside stimuli and open up new possibilities for imaging, biosensors, and drug administration ³⁴.

Synthesis Aldehydes and Ketones: Either Schiff base production or regulated oxidation of chitosan can introduce aldehydes and ketones. Effect on Chitosan: The presence of aldehydes and ketones increases the chitosan's potential for further chemical alterations and speeds up cross-linking events, which affects how it is used to make hydrogels and transport drugs ³⁵.

Synthesis Quaternary Ammonium Salts: Reactions involving alkyl halides, epoxides, or other quaternizing agents add quaternary ammonium salts. Effect on Chitosan: Chitosan's antimicrobial qualities are strengthened by the addition of positively charged quaternary ammonium groups, which makes it appropriate for use in water treatment, antibacterial coatings, and wound dressings ³⁶.

Synthesis Hydroxyl Derivatives: By reacting with different reagents, such as epoxides or isocyanates, hydroxyl derivatives can be introduced. Chitosan is impacted by hydroxyl derivatives because they improve hydrophilicity and provide more sites for chemical alteration. Tissue engineering, wound healing, and drug delivery are three areas where modified chitosan hydroxyl derivatives are useful ³⁷.

Synthesis Thiol Groups: Through reactions with reagents containing thiols, thiol groups can be added. Effect on Chitosan: The presence of thiol groups allows chitosan to take part in thiol-disulfide exchange processes, which promotes the creation of materials that respond to stimuli and bio conjugation techniques ³⁸. A thorough understanding of the synthesis and effects of various functional groups leads to a customized modification of chitosan. This information may be used by researchers to create chitosan derivatives with carefully adjusted characteristics, increasing the material's versatility in a variety of applications. Figure 3 shows possible reactions to form various functional groups with chitosan. ³⁹

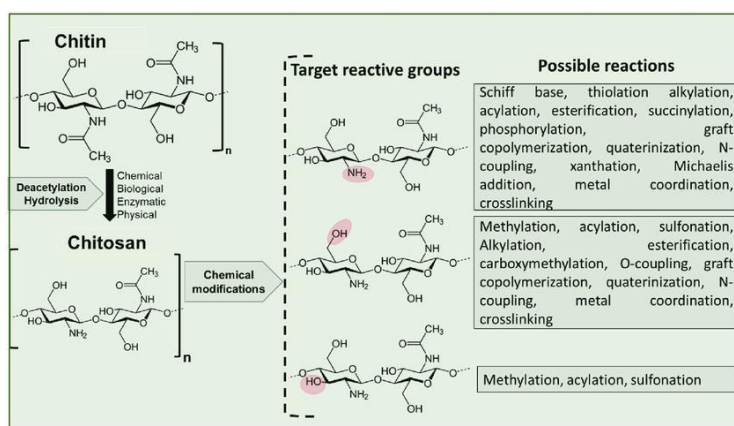


Figure 3 Possible reactions to form various functional groups with chitosan ³⁹.

Challenges, and Future Perspectives:

In both medical and industrial applications, it is important to address the toxicity of modified chitosan^{40,41}. This is because the original compound is biopolymer and biodegradable. However, this is not the case the produced modified materials have the same characteristics. Another important issue should be addressed is the solubility of chitosan in common organic solvents. Thus, chitosan is either insoluble or very slightly soluble in the most of organic solvent.

CONCLUSIONS:

In conclusion, the area of chitosan organic modification is greatly expanded the range of using it in both medical and industrial applications. The desire to chemically alter chitosan to make it more soluble and extend its applications has grown recently. Chitin is typically obtained by deacetylating it in an alkaline solution to produce chitosan. One of the most prevalent organic compounds is chitin, which is produced annually via biosynthesis second only to cellulose. Animal exoskeletons, especially those of insects, mollusks, and crustaceans, heavily depend on chitin. It is also the primary fibrillar polymer present in the cell walls of some fungi. Chitosan is a linear polysaccharide made up of glucosamine and N-acetyl glucosamine units joined by β (1–4) glycosidic links. The quantity of glucosamine present is referred to as the degree of deacetylation. Well, broadly speaking. With advancements in green chemistry, smart polymers, and nanotechnology ready to open up new applications in tissue engineering, drug delivery, and other fields, the future of chitosan modification seems bright. To fully utilize this extraordinary biopolymer in a variety of scientific and industrial contexts, chitosan modification methods must be further investigated and improved.

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DECLARATIONS

Authors' contributions

Both authors contributed equally to the design, execution, and interpretation of the research, as well as the drafting and revision of the manuscript. All authors approved the final version of the manuscript and are responsible for its content.

Conflict of interest

The authors declare that they have no potential conflicts of interest.

REFERENCES

1. Wang W, Meng Q, Li Q, Liu J, Zhou M, Jin Z, et al. Chitosan derivatives and their application in biomedicine. *International journal of molecular sciences*. 2020;21(2):487–487. doi.org/10.3390/ijms21020487.
2. 40-Hammood AJ, Al-Ameri SA, Al-Qaisi ZHJ. Synthesis, Characterization and Kinetic Study of poly (vinyl acetate)-g-Chitosan and its Use in Removal of Cadmium (II) ions from Water. *Al-Nahrain Journal of Science*. 2019;22(2):1–11. [10.22401/ANJS.22.2.01](https://doi.org/10.22401/ANJS.22.2.01).
3. Kumar MR, Muzzarelli R, Muzzarelli C, Sashiwa H, Domb AJ. Chitosan chemistry and pharmaceutical perspectives. *Chemical reviews*. 2004;104(12):6017–6084. doi.org/10.1021/cr030441b.
4. Ardean C, Davidescu CM, Nemeş NS, Negrea A, Ciopec M, Duteanu N, et al. Factors influencing the antibacterial activity of chitosan and chitosan modified by functionalization. *International Journal of Molecular Sciences*. 2021;22(14):7449–7449. doi.org/10.3390/ijms22147449.
5. 24-Aljawish A, Chevalot I, Jasniowski J, Scher J, Muniglia L. Enzymatic synthesis of chitosan derivatives and their potential applications. *Journal of Molecular Catalysis B: Enzymatic*. 2015;112:25–39. doi.org/10.1016/j.molcatb.2014.10.014.
6. Qu B, Luo Y. Chitosan-based hydrogel beads: Preparations, modifications and applications in food and agriculture sectors-A review. *International journal of biological macromolecules*. 2020;152:437–448. doi.org/10.1016/j.ijbiomac.2020.02.240.
7. Sashiwa H, Shigemasa Y, Roy R. Synthesis of dendronized chitosan– sialic acid hybrid using convergent grafting of preassembled dendrons built on gallic acid and Tri (ethylene glycol) backbone. *Macromolecules*. 2001;10(12):3905–3909. doi.org/10.1021/ma001832k.
8. Kaczmarek MB, Struszczyk-Swita K, Li X, Szczęśna-Antczak M, Daroch M. Enzymatic modifications of chitin, chitosan, and chito oligosaccharides. *Frontiers in bioengineering and biotechnology*. 2019;7:243–243. doi.org/10.3389/fbioe.2019.00243.
9. 41-Kumbhare M, Surana AR, Morankar PG. Nose-to-brain delivery of insulin nanoparticles for diabetes management: A review. *Baghdad Journal of Biochemistry and Applied Biological Sciences*. 2023;4(02):39–49. doi.org/10.47419/bjbabs.v4i01.178.
10. Kurita K, Ikeda H, Yoshida Y, Shimojoh M, Harata M. Chemoselective protection of the amino groups of chitosan by controlled phthaloylation: facile preparation of a precursor useful for chemical modifications. *Biomacromolecules*. 2002;3(1):1–4. doi.org/10.1021/bm0101163.
11. Mittal H, Ray SS, Kaith BS, Bhatia JK, Sharma J, Alhassan SM. Recent progress in the structural modification of chitosan for applications in diversified biomedical fields. *European Polymer Journal*. 2018;109:402–434. doi.org/10.1016/j.eurpolymj.2018.10.013.
12. Mourya VK, Inamdar NN. Chitosan-modifications and applications: Opportunities galore. *Reactive and Functional polymers*. 2008;68:1013–1051.

- doi.org/10.1016/j.reactfunctpolym.2008.03.002.
- 13 Bakshi PS, Selvakumar D, Kadirvelu K, Kumar NS. Chitosan as an environment friendly biomaterial-a review on recent modifications and applications. *International journal of biological macromolecules*. 2020;150:1072–1083. doi.org/10.1016/j.ijbiomac.2019.10.113.
- 14 26-Thakur VK, Thakur MK. Recent advances in graft copolymerization and applications of chitosan: a review. *ACS Sustainable Chemistry & Engineering*. 2014;2(12):2637–2652. doi.org/10.1021/sc500634p.
- 15 Chopin N, Guillory X, Weiss P, Bideau JL, Collic-Jouault S. Design polysaccharides of marine origin: chemical modifications to reach advanced versatile compounds. *Current Organic Chemistry*. 2014;18(7):867–895. dx.doi.org/10.2174/138527281807140515152334.
- 16 Ji, Wang J, Yu L, Chen H, Zhao Y, Zhang Y, et al. Chemical modifications of chitosan and its applications. *Polymer-Plastics Technology and Engineering*. 2014;53(14):1494–1505. doi.org/10.1080/03602559.2014.909486.
- 17 Argüelles-Monal WM, Lizardi-Mendoza J, Fernández-Quiroz D, Recillas-Mota MT, Montiel-Herrera M. Chitosan derivatives: introducing new functionalities with a controlled molecular architecture for innovative materials. *Polymers*. 2018;10(3):342–342. doi.org/10.3390/polym10030342.
- 18 5-Negm NA, Hefni HH, Abd-Elal AA, Badr EA, Kana MTA. Advancement on modification of chitosan biopolymer and its potential applications. *International journal of biological macromolecules*. 2020;152:681–702. doi.org/10.1016/j.ijbiomac.2020.02.196.
- 19 35-Cumpstey I. Chemical modification of polysaccharides *International scholarly research notices*. 2013; Available from: <http://dx.doi.org/10.1155/2013/417672>. doi.org/10.1155/2013/417672.
- 20 Abed RN, Sattar MA, Hameed SS, Ahmed DS, Al-Baidhani M, Kadhom M, et al. Optical and morphological properties of poly (vinyl chloride)-nano-chitosan composites doped with TiO₂ and Cr₂O₃ nanoparticles and their potential for solar energy applications. *Chemical Papers*. 2023;77(2):757–769. doi.org/10.1007/s11696-022-02512-6.
- 21 8-Aranaz I, Alcántara AR, Civera MC, Arias C, Elorza B, Caballero AH, et al. Chitosan: An overview of its properties and applications. *Polymers*. 2021;13(19):3256–3256. doi.org/10.3390/polym13193256.
- 22 Alves NM, Mano JF. Chitosan derivatives obtained by chemical modifications for biomedical and environmental applications. *International journal of biological macromolecules*. 2008;43(5):401–414. doi.org/10.1016/j.ijbiomac.2008.09.007.
- 23 Saleh TAK, Al-Tikrity ET, Yousif E, Al-Mashhadani MH, Jawad AH. Preparation of Schiff bases derived from chitosan and investigate their photostability and thermal stability. *Physical Chemistry Research*. 2022;10(4):549–557. doi.org/10.22036/PCR.2022.333808.2051.
- 24 Knidri H, Belaabed R, Addaou A, Laajeb A, Lahsini A. Extraction, chemical modification and characterization of chitin and chitosan. *International journal of biological macromolecules*. 2018;120:1181–1189. doi.org/10.1016/j.ijbiomac.2018.08.139.
- 25 Silva SS, Mano JF, Reis RL. Ionic liquids in the processing and chemical mod-

- ification of chitin and chitosan for biomedical applications. *Green Chemistry*. 2017;19(5):1208–1220. doi.org/10.1039/C6GC02827F.
- 26 Valdez O, Champagne-Hartley R, Saldívar-Guerra E, Champagne P, Cunningham MF. Modification of chitosan with polystyrene and poly (n-butyl acrylate) via nitroxide-mediated polymerization and grafting from approach in homogeneous media. *Polymer Chemistry*. 2015;6(15):2827–2836. doi.org/10.1039/C5PY00028A.
- 27 23-Nornberg AB, De Aquino TF, Martins CC, Luchese C, Wilhelm EA, Jacob RG, et al. Organoselenium-chitosan derivative: Synthesis via “click” reaction, characterization and antioxidant activity. *International Journal of Biological Macromolecules*. 2021;191:19–26. doi.org/10.1016/j.ijbiomac.2021.09.053.
- 28 32-Alqahtani NF. Functionalized imidazolium ionic liquids-modified chitosan materials: From synthesis approaches to applications *Reactive and Functional Polymers*. 2023;p. 105779–105779. doi.org/10.1016/j.reactfunctpolym.2023.105779.
- 29 Essawy AA, Hefni H, El-Nggar AM. Biocompatible and Biodegradable Chitosan Composites in Wound Healing Application. *Situ Novel Photo-Induced Skin Regeneration Approach Sustainable Polymer Composites and Nanocomposites*. 2019;p. 143–183. doi.org/10.1007/978-3-030-05399-4_5.
- 30 Sahariah P, Másson M. Antimicrobial chitosan and chitosan derivatives: A review of the structure-activity relationship. *Biomacromolecules*. 2017;18(11):3846–3868. doi.org/10.1021/acs.biomac.7b01058.
- 31 Vakili M, Rafatullah M, Salamatinia B, Abdullah AZ, Ibrahim MH, Tan KB, et al. Application of chitosan and its derivatives as adsorbents for dye removal from water and wastewater: A review. *Carbohydrate polymers*. 2014;113:115–130. Available from: <https://doi.org/10.1016/j.carbpol.2014.07.007>. doi.org/10.1016/j.carbpol.2014.07.007.
- 32 Abdelwahab HE, Hassan SY, Yacout GA, Mostafa MA, Sadek MM. Synthesis and biological evaluation of new imine-and amino-chitosan derivatives. *Polymers*. 2015;7(12):2690–2700. doi.org/10.3390/polym7121532.
- 33 24-Aljawish A, Chevalot I, Jasiewicz J, Scher J, Muniglia L. Enzymatic synthesis of chitosan derivatives and their potential applications. *Journal of Molecular Catalysis B: Enzymatic*. 2015;112:25–39. doi.org/10.1016/j.molcatb.2014.10.014.
- 34 Ji, Wang J, Yu L, Chen H, Zhao Y, Zhang Y, et al. Chemical modifications of chitosan and its applications. *Polymer-Plastics Technology and Engineering*. 2014;53(14):1494–1505. doi.org/10.1080/03602559.2014.909486.
- 35 Kurita K, Ikeda H, Yoshida Y, Shimojoh M, Harata M. Chemoselective protection of the amino groups of chitosan by controlled phthaloylation: facile preparation of a precursor useful for chemical modifications. *Biomacromolecules*. 2002;3(1):1–4. doi.org/10.1021/bm0101163.
- 36 Wang W, Meng Q, Li Q, Liu J, Zhou M, Jin Z, et al. Chitosan derivatives and their application in biomedicine. *International journal of molecular sciences*. 2020;21(2):487–487. doi.org/10.3390/ijms21020487.
- 37 Ardean C, Davidescu CM, Nemeş NS, Negrea A, Ciopec M, Duteanu N, et al. Factors influencing the antibacterial activity of chitosan and chitosan modified by functionalization. *International Journal of Molecular Sciences*. 2021;22(14):7449–7449. [10.3390/ijms22147449](https://doi.org/10.3390/ijms22147449).

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- 38 Essawy AA, Hefni H, El-Nggar AM. Biocompatible and Biodegradable Chitosan Composites in Wound Healing Application. Situ Novel Photo-Induced Skin Regeneration Approach Sustainable Polymer Composites and Nanocomposites. 2019;p. 143–183. doi.org/10.1007/978-3-030-05399-4_5.
- 39 Vakili M, Rafatullah M, Salamatinia B, Abdullah AZ, Ibrahim MH, Tan KB, et al. Application of chitosan and its derivatives as adsorbents for dye removal from water and wastewater: A review. Carbohydrate polymers. 2014;113:115–130. doi.org/10.1016/j.carbpol.2014.07.007.
- 40 Mittal H, Ray SS, Kaith BS, Bhatia JK, Sharma J, Alhassan SM. Recent progress in the structural modification of chitosan for applications in diversified biomedical fields. European Polymer Journal. 2018;109:402–434. doi.org/10.1016/j.eurpolymj.2018.10.013.
- 41 Knidri H, Belaabed R, Addaou A, Laajeb A, Lahsini A. Extraction, chemical modification and characterization of chitin and chitosan. International journal of biological macromolecules.2018;120:1181–1189. doi.org/10.1016/j.ijbiomac.2018.08.139.