Effect of the Biofertilizers and the Methods of its Addition on the Growth and Yield of Eggplant (*Solanum melongena* L.) Under Greenhouse Conditions

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Abstract

Research was conducted at Al-latifia Research Station of Plant Breeding and Improvement Center, Agricultural Research Directorate, during spring season of year 2016 to study the effect of some biofertilizers on growth and yield of Eggplant plant under greenhouse conditions. A factorial experiment was designed in Randomized Complete Block Design with three replicates and the means of traits compared by L.S.D. at 5% level. The main factor included methods of adding biofertilizers to seeds (A1) and transplants (A2). The secondary factor was biofertilizers included: without adding any Biofertilizer (control) (T0), *Pseudomonas fluorescence*, (T1), *Azospirillum brasilense* (T2), *Bacillus subtilis* (T3) and *Azotobacter chroococcum* (T4). The results showed that *Azotobacter chroococcum* treatment significantly increased the plant height (87.70 cm) compared with (57.70 cm) in the control treatment. Moreover, *Azotobacter chroococcum* treatment showed significant increases the yield per plant and the total yield which were, (1000.00 gm.plant⁻¹ and 899.00 Kg.greenhouse⁻¹) respectively in comparison with the control treatment which were (553.00 gm.plant⁻¹) and (498.00 Kg.greenhouse⁻¹) respectively.

Key Words: Azospirillum brasilense, Azotobacter chroococcum, Transplants and Grow.

تأثير المخصبات الاحيائيه وطرق اضافتها في نمو وحاصل الباذنجان Solanum melongena L. تحت ظروف البيت البلاستيكي

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لخلاصية

نفذ البحث في البيت البلاستيكي لمحطة ابحاث اللطيفية التابعة لمركز تربية وتحسين النبات للموسم الربيعي 2016 لدراسة تأثير بعض المخصبات الأحيائية في نمو وحاصل الباذنجان تحت ظروف البيت البلاستيكي و طبقت تجربه عامليه باستخدام تصميم القطاعات الكاملة المعشاة وبثلاث مكررات وقورنت المتوسطات الحسابيه للصفات المدروسه حسب أختبار LSD عند مستوى أحتمال 5 % اذ تضمن العامل الرئيسي طرق اضافة المخصبات للادور(A1) وللشتلات (A2) وتضمن العامل الثانوي المخصبات الاحيائية التي شملت: بدون المخصبات الاحيائية التي شملت: بدون المخصبات المعاملة المقارنة (T1) Pseudomonas fluorescence (T0) وتضمن العامل الثانوي المخصبات الاحيائية تأوق معاملة المعاملة المعاملة

Introduction

Eggplant (Solanum melongena L) is one of the main summer vegetables. It is an important vegetable crop grown worldwide. So, cultivation has spread throughout the world, and planting in most areas of Iraq on fields and also grown in greenhouses and tunnels (Almohammadi, 1990). It is planted for its fruits, which is a popular food for most developing countries. It is used in cooking, pickles and canned. Also, it kept freezing and drying (Salloum, 2012). Crop service under greenhouse conditions requires chemical fertilizers either by adding to the soil or spraying on plants to obtain a high yield (Al-zehawi, 2007). Chemical fertilization rates for vegetables have been increased relative to other crops because they can be planted more than one season per year, which led to increase the harmful effects on health and environment, especially the residual effect of nitrates which is considered one of the most dangerous compounds for human health. Therefore, the world is turning to Biofertilizers instead of chemical fertilizers to reduce environmental pollution. Biofertilizers are environmentally safe alternatives and have a big impact to get high production and avoid chemical pollution (Alhaddad, 1989) Biofertilizer is a natural product carrying living microorganisms derived from the root or cultivated soil. So, they don't have any ill effect on soil health and environment. Besides their role in atmospheric nitrogen fixation and phosphorous solubilization, these also help in stimulating the plant growth hormones providing better nutrient uptake and increased tolerance towards drought and moisture stress. (Anandaraj and Delapierre, 2010).

The beneficial effects of Biofertilizers such as *Azotobacter* and *Azospirillum* on plants are attributed mainly to an

improvement in root development, an increase in the rate of water and minerals uptake by roots, displacement of fungi and plant pathogenic bacteria and to a lesser extent, biological nitrogen fixation (Meena, et al., 2017). Moreover, inhibit and stop the growth of harmful microorganisms and they have the ability to stimulate, increase roots and shoot increase the absorption of growth, essential nutrients and increase plant ability to resist unsuitable environmental conditions (Allawi, 2013) Biofertilizers input in soil activated appropriate microbiological processes which can be more and better uniform supply of plants with nitrogen, phosphorus potassium, as well as some trace elements (Tosic, et al., 2016) (Aldulaimi, et al., 2003) explained that the use of the *Pseudomonas fluorescence* has increased growth of tomato plants under greenhouse conditions. The addition of Azospirillium sp. to peas plant was observed in giving a significant increase in plant height and number of leaves compared with control treatment (Thenmozhi, et al., 2010). While the addition of Biofertilizers (Azotobacter and Azosprillium) on tomato plants led to increase in plant height, number of leaves, relative content of chlorophyll, number of fruits and plant yield (Ramakrishnam and Selvakumar, 2012). Whereas (Rao, et al., 2014) found the treatment of cabbage with Azotobacter and Azosprillium increased the yield under plastic house conditions. Moreover, using Azotobacter Bacillus on lettuce gave the highest increase in plant yield (Tosic, et al., 2016).

In another study, using *Azospirillium* fertilizer with 25% FYM (Farm Yard Manure) gave increase in plant yield and total yield compared with control treatment (Meena, *et al.*, 2017). The use of suitable

combination of *Azotobacter* with 75% nitrogen fertilizer gave significant increases of plant height, number of branches and number of fruits and total yield of tomato plants (Shashi, *et al.*, 2018). The aim of this study is to determine the effect of biofertilizers on growth and development of eggplant in the greenhouses.

Materials and Methods

This experiment was carried out in the greenhouses at Al-latifia Research Station of Plant Breeding and Improvement Center, Agricultural Research Directorate during the Spring season of 2016 using Eggplant (Barcelona cv.).

Preparation of Seeds and Transplants 1. Treatment of Seeds

Seeds of eggplant were treated with biofertilizer by coating them with the bacterial suspension for 10 minutes and then let it to dry for 15 minutes. The treated and untreated seeds were planted on 7/10/2016 in transplanting trays which contain peat moss, and the trays were kept inside the green house for germination of seeds before transferring to the permanent place (Naeem, 2012).

2 - Treatment of Transplants

Seeds were sown in transplanting trays which contain peat moss on 7/10/2016. biofertilizers were added to eggplant seedlings when real leaf appeared at a rate of 5 ml of bacterial suspension for each transplant. The trays were placed in the green house until transferring to the permanent place (Naeem, 2012).

Preparing the Green House and Transplants

The green house was prepared with a distance of 9 * 50 m and solar sterilization was applied from 15 June until 1 September 2016 and then divided to five blocks (3 Blocks Were Used) with a length of 50 m and width of 0.80 m and a distance between raw and other 0.80 m with 1 m left on each side. Each block was divided to 9 sections of 5 m each

experimental unit, Eggplant transplants were planted with a distance of 0.60 m between plants on 12/11/2016. Chemical fertilizer was added two times, the first was 14 days after transplanting and included 120Kg N ha⁻¹ (Urea 46% N) and 160 Kg P ha⁻¹ (Super Triphosphate 21%) and 120 Kg K ha⁻¹(Potassium Sulphate 46% K) and the same amount added after one month from the first treatment (Al-naimi, 1999). Chemical and physical analysis were carried out according to Richards (1954) as shown in Table (1).

Table (1) Chemical and Physical Properties of Soil

Properties		Standard Unit	Value
(1:1) pH			7.2
(1:1)	Ec	dsm ⁻¹	3.3
Organic	Matter	gKg^{-1}	1.02
HC	O ₃	gKg-1	2.33
Available Nitrogen		mgKg ⁻¹	60.5
Avail Phospl		mgKg ⁻¹	11.4
Avail Potass		mgKg ⁻¹	156.3
Iona	Mg ++	mgL ⁻¹	73.00
Ions Soluble	Ca ++	mgL ⁻¹	199.1 0
Silt		gKg ⁻¹	465
Clay		gKg ⁻¹	370
Sand		gKg^{-1}	165
Soil Mixture		Silty Clay	

The factorial experiment was laid out in Randomized Complete Block Design (R.C.B.D) with three replicates. Means of traits were compared by L.S.D. at level 5% (Al-Sahuki and Wahib, 1990) using Statistical Analysis System (SAS, 2001).

 $Table\ (2)\ Effect\ of\ Biofertilizers\ and\ Methods\ of\ Addition\ on\ the\ Plant\ Height\ and\ Stem\ Diameter$

of Eggplant under the Greenhouse Conditions

Method of	P	lant Height (c	em)	St	em Diameter(cm)
Addition A Bio Fertilizers T	Seeds A1	Transplant A2	Mean of Bio Fertilizers	Seeds A1	Transplant A2	Mean of Bio Fertilizers
Т0	57.70	57.70	57.70	1.17	1.17	1.17
T1	75.30	96.00	85.70	1.45	1.52	1.48
T2	77.30	95.00	86.20	1.58	1.15	1.36
Т3	79.00	93.00	86.00	1.58	1.61	1.59
T4	78.70	96.70	87.70	1.45	1.78	1.61
Mean of a Method of Addition	73.60	87.70		1.44	1.44	
L.S.D 0.05	Method of Addition	Bio Fertilizers	Method of Addition Bio Fertilizers	Method of Addition	Bio Fertilizers	Method of Addition Bio Fertilizers
	9.37	14.81	20.94	N.S	N.S	N.S

T0 control T1 Pseudomonas fluorescence T2 Azospirillum brasilense T3 Bacillus subtilis T4 Azotobacter chroococcum

Biofertilizer Inoculation

Biofertilizers were obtained from Biotechnology Center/Agricultural Research Directorate.

Treatments Included

1- Methods of Addition inocula (A)

A1: Add the inocula to seeds.

A2: Add the inocula to the transplants.

2. The biofertilizer (T) included

T0: without Biofertilizer (Control)

T1: Pseudomonas fluorescence

T2: Azospirillum brasilense

T3: Bacillus subtilis

T4: Azotobacter chroococcum

Observation

The plant height, stem diameter (cm), average number of branches, fresh weight of plants and roots g, chlorophyll in the leaves was measured by

Chlorophyll meter device type SPAD - 502 (Minnotti, *et al.*, 1994) and the leaf area was measured with the portable leaf area meter (USACI-202) (Tekalign & Hammes, 2005) were measured after 55 days for 5 plants from each treatment. Furthermore, number of fruits per plant, weight of fruits, yield per plant (gm), total yield (Kg), fruit diameter and length were calculated.

Results and Discussion Plant Shoot Growth

The results in the Table (2) showed that methods of inocula addition were significantly differences in plant height, the transplants treatment (A2) gave the highest significant differences (P<0.05) in plant height at (87.70 cm) compared

with (A1) the seed treatment which reached (73.60 cm). So, the biofertilizers treatments, results showed that the T4 treatment (Azotobacter) significantly higher than (87.70 cm) and T2 followed it by treatment (Azospirillum) was (86.20 cm) compared with T0 treatment (without Biofertilizer), which gave (57.70 cm). The Interaction between the methods of adding (A) and (T) biofertilizers treatments had a significant effect with A2T4 (transplants + Azotobacter) treatment which gave of (96.70 cm), while A1T0 and A2T0 treatments gave (57.70 cm) for both.

Also, the results in the same Table showed non-significant differences between methods of addition biofertilizer and used biofertilizers and interaction between them in stem diameter.

Results in Table (3) indicated nonsignificant differences in the number of branches per plant due to methods of inocula addition and the biofertilizers used. The interaction between the methods of addition and biofertilizers treatments showed significant effect. The treatment combination of A2T2 gave the highest number of branches (9.67) while the lowest number of branches was recorded with A1T0 and A2T0 treatments which reached (4.33). The results also showed that non-significant effects were obtained from the methods of addition in shoot weight of plant. However, significant effect on shoot weight was found due to biofertilizers.

Table (3) Effect of Biofertilizers and Methods of Addition on the Number of Branches and Fresh Weight of Eggplant Shoots under the Greenhouse Conditions.

Method of		No. of Branch			Weight of Sh	noot (g)
Addition A Bio Fertilizers T	Seeds A1	Transplant A2	Mean of Bio Fertilizers	Seeds A1	Transplant A2	Mean of Bio Fertilizers
Т0	4.33	4.33	4.33	195.00	195.00	195.00
T1	7.07	7.00	7.03	319.00	441.00	380.00
T2	8.67	9.67	9.17	303.00	434.00	369.00
T3	7.33	8.33	7.83	307.00	383.00	345.00
T4	8.67	9.00	8.83	427.00	571.00	499.00
Mean of a Method of Addition	7.21	7.67		310.00	405.00	
L.S.D 0.05	Method of Addition	Bio Fertilizers	Method of Addition Bio Fertilizers	Method of Addition	Bio Fertilizers	Method of Addition Bio Fertilizers
	N.S	N.S	4.68	N.S	178.70	252.70

T4 treatment recorded the highest shoot weight (499.00 gm) while T0 treatment gave the lowest shoot weight (195.00 gm). The interaction between the methods of addition and biofertilizers showed a significant effect with A2T4 treatment which gave (571.00 gm), while A1T0 and A2T0 treatments gave (195.00 gm). Data of Table (4) showed nonsignificant differences in the roots fresh weight between the two methods of inocula addition whereas, biofertilizer had a significant effect, with the highest rate of (42.90 gm) in the T4 treatment compared with 27.40 gm for T0 treatment.

The interaction between the methods of addition and biofertilizer treatments had a significant effect in the highest weight reached (45.40 gm) with A2T4 treatment compared with the lowest weight of (27.40 gm) with A1T0 and A2T0 treatments. In addition to significant, the influence of methods of addition on chlorophyll content in leaves was recorded with transplants treatment A2 significantly superior which reached (47.39) SPAD units compared with the seeds treatment A1 which gave (44.86) SPAD units (Table 4).

Table (4) Effect of Biofertilizers and Methods of Addition on Fresh Weight of Roots and Relative

Chlorophyll Content of Eggplant Shoots under the Greenhouse Conditions.							
Method /	Fresl	Fresh Weight of Root (g)		Relative Chlorophyll Content			
of Addition A Bio Fertilizers T	Seeds A1	Transplant A2	Mean of Bio Fertilizers	Seeds A1	Transplant A2	Mean of Bio Fertilizers	
T0	27.40	27.40	27.40	44.00	44.00	44.00	
T1	36.80	44.40	40.60	46.17	48.27	47.22	
T2	37.70	44.70	41.20	44.73	49.00	46.87	
Т3	38.90	44.00	41.50	44.30	46.10	45.20	
T4	40.50	45.40	42.90	45.10	49.60	47.35	
Mean of a Method of Addition	36.30	41.20		44.86	47.39		
L.S.D 0.05	Method of Addition	Bio Fertilizers	Method of Addition Bio Fertilizers	Method of Addition	Bio Fertilizers	Method of Addition Bio Fertilizers	
	N.S	11.82	16.72	1.39	2.21	3.12	

T1 Pseudomonas fluorescence T0 control T2 Azospirillum brasilense T3 Bacillus subtilis T4 Azotobacter chroococcum

Table (5) Effect of Biofertilizers and Methods of Addition on Leaf Area of Eggplant under the Greenhouse Conditions

Method of	Leaf Area dcm ²						
Addition A Bio Fertilizer T	Seeds A1	Transplant A2	Mean of Bio Fertilizers				
T0	45.50	45.90	45.70				
T1	71.30	64.50	67.90				
T2	71.10	56.70	63.90				
T3	48.00	55.80	51.90				
T4	55.30	56.60	56.00				
Mean of a Method of	58.20	55.90					
Addition							
	Method of	Bio Fertilizers	Method of Addition				
L.S.D 0.05	Addition		Bio Fertilizers				
	N.S	N.S	N.S				
T0 control T1 Pseudomonas fluorescence							

T2 Azospirillum brasilense T3 Bacillus subtilis T4 Azotobacter chroococcum

The results of the same Table also showed significant differences in effect of Bio- fertilizers on chlorophyll content where the T4 treatment was significantly superior which reached (47.35) SPAD units and reduced significantly in T0 treatment to 44.00 SPAD units. The interaction between the methods of inocula addition and biofertilizers treatments had a significant effect with (transplants + Azotobacter) A2T4 treatment gave the highest amount of chlorophyll (49.60) SPAD units, while A1T0 and A2T0 treatments gave (44.00) SPAD units.

The results of Table (5) showed nonsignificant differences between methods of addition and the biofertilizers used and the interaction between them on the leaf area/dcm². The increases in most shoot characteristics of plants biofertilizers inoculation (Azotobacter and Azospirillum) attributed to the role of biofertilizers in the availability, absorption and concentration of nutrients phosphorus. such as nitrogen and a role in biofertilizers also play

stimulating the production of growth regulators, which are positively reflected in increased division, elongation and expansion of cells, which reflected on shoot growth (Salhia, 2010; Allawi, 2013; Shashi, et al., 2018), and the availability of elements in the leaves lead to increase management and activity of photosynthesis, which leads to increase CO₂ in the leaves, which is the basic unit for building carbohydrates, protein's structure and amino acids and therefore increasing vegetative growth. The results of the present study were in agreement with (Islah and El-Sayed, Dharmendra, 2014; Saeed, 2015 and Meena, et al., 2017).

Yield and It's Component

Table (6) showed that the methods of inocula addition gave a significant difference in the number of fruits per plant. A significantly higher number of fruits per plant was recorded in transplants treatment A2 was (5.13) fruit while the seed treatment gave (4.18) fruits per plant. While biofertilizers

varied significantly for the number of fruits per plant. biofertilizers application T2 and T4 treatments recorded higher fruits per plant reached (5.23 and 5.23) fruits respectively, compared with T0 treatment (without Biofertilizers) which gave (3.57) fruits. The interaction between the methods of inocula addition and biofertilizers application showed a significant effect with A2T2 and A2T4 treatments gave (5.69 and 5.81) fruits respectively, while A1T0 treatment gave (3.57) fruits. No significant differences in fruit weight were recorded among methods of inocula addition biofertilizers and the interaction between them. The methods of inocula addition showed significant differences in plant yield Table (7). The highest value of plant yield was obtained from inocula treatments of transplant A2 which was (942.00 gm.plant⁻¹) and significantly higher than the seed treatment A1 which gave (734.00 gm.plant⁻¹). The results showed that biofertilizer treatments, T4 treatment (Azotobacter)

significantly higher than other treatments which reached (1000.00 gm) while the lowest was for the control treatment T0 which reached (553.00 gm.plant⁻¹).

The interaction between the methods of inocula addition and biofertilizer treatments was found to be significant with the highest value for treatment A2T4 (1116.00 gm.plant⁻¹) and the lowest plant yield was recorded for A1T0 treatments which reached (553.00 gm.plant⁻¹).

Moreover, the results of Table (7) showed a significant effect in the total yield of greenhouse according to the methods of inocula addition which reached (848.00 Kg) in the treatment of transplants A2 compared with the treatment of seeds A1 which gave (660.00 Kg). In addition, biofertilizers varied significantly for plant yield with T4 treatment (Azotobacter) gave a higher than reached (899.00 value compared with T0 treatment (without biofertilizers) which gave (498.00 Kg).

Table (6) Effect of Biofertilizers and Methods of Inocula Addition on Number of Fruits and Fruit Weight of Eggnlant under the Creenhouse Conditions

Method of	gplant under the Greenhouse Conditions Number of Fruits Fruit Weight(
Addition A Bio Fertilizers T	Seeds A1	Transplant A2	Mean of Bio Fertilizers	Seeds A1	Transplant A2	Mean of Bio Fertilizers
Т0	3.57	3.57	3.57	153.50	153.50	153.50
T1	4.11	5.39	4.75	185.80	186.50	186.10
T2	4.56	5.69	5.13	182.40	201.30	191.90
Т3	4.00	5.18	4.59	173.60	186.10	179.80
Fertilizers T4	4.66	5.81	5.23	198.50	194.30	196.40
Mean of a Method of Addition	4.18	5.13		178.80	184.30	
L.S.D 0.05	Method of Addition	Bio Fertilizers	Method of Addition Bio Frtilizers	Method of Addition	Bio Fertilizers	Method of Addition Bio Frtilizers
	0.69	1.09	1.55	N.S	N.S	N.S

T1 Pseudomonas fluorescence

T2 Azospirillum brasilense T3 Bacillus subtilis T4 Azotobacter chroococcum

Table (7) Effect of Biofertilizers and Methods of Inocula Addition on Yield of Plant Height and

Method of	Ŋ	ield of plant	(g)	Total yield of house (Kg)		
Addition A Bio Fertilizers T	Seeds A1	Transplant A2	Mean of Bio Fertilizers	Seeds A1	Transplant A2	Mean of Bio Fertilizers
T0	553.00	553.00	553.00	498.00	498.00	498.00
T1	740.00	974.00	857.00	666.00	876.00	771.00
T2	848.00	1108.00	978.00	763.00	997.00	880.00
T3	647.00	958.00	802.00	582.00	862.00	722.00
T4	884.00	1116.00	1000.00	793.00	1005.00	899.00
Mean of a Method of addition	734.00	942.00		660.00	848.00	
L.S.D 0.05	Method of Addition	Bio Fertilizers	Method of Addition Bio Fertilizers	Method of Addition	Bio Fertilizers	Method of Addition Bio Fertilizers
	159.00	251.40	355.50	143.10	226.20	320.00

T0 control T1 Pseudomonas fluorescence

T2 Azospirillum brasilense T3 Bacillus subtilis T4 Azotobacter chroococcum

The Interaction between the methods of inocula addition and biofertilizer treatments found to be significant with the highest value recorded for treatment A2T4 which gave (1005.00 Kg) while the lowest yield was recorded by A1T0 and A2T0 treatments reached (498.00 Kg) for both of them.

The results of Table (8) showed significant differences in length of fruit by methods of inocula addition wherein the transplants treatment A2 reached (11.1 cm) compared with the seed treatment A1 which gave (8.9 cm), whereas biofertilizer had a significant effect, with the highest rate of (11.4 cm) was in the T4 treatment and the lowest was 7.3 cm in T0 treatment. The interaction between the methods of inocula addition and biofertilizer treatments showed a significant effect with A2T2 treatment gave (12.9 cm)

while reduced to (7.3 cm) in A1T0 treatment. The diameter of fruit in Table (8) was significantly different depending on the methods of inocula Addition. was recorded in transplants treatment A2 reached (4.68 cm) compared with seeds treatment A1 which gave (4.40 cm). Also, biofertilizer treatments showed a significant effect on fruit diameter and the results showed that T4 treatment (Azotobacter) was the highest of (4.88 cm), while the lowest was in control treatment T0 of (3.81 cm).

The interaction between the methods of inocula addition and biofertilizer treatments showed a significant effect on fruit diameter in A2T4 treatment which gave the highest value of (5.06 cm) while the lowest value of (3.81 cm) in A1T0 treatment.

The inoculation of eggplant with biofertilizers led to stimulation and

increase of growth through strategies which are used in this biological system, especially the availability of nutrients through phosphorus solubilization and nitrogen fixation in soil and increase resistance of plants to biotic and abiotic stresses and production of different growth regulators like IAA and GA3. All these factors contributed to increasing lengths and diameters of fruits, number of fruits and weight of fruit, which led to increasing of plant growth and total yield. (Saharan and Nehra, 2011, Dharmendra, 2014, Saeed, 2015 and Shashi, et al., 2018). The growth regulators also contribute to increasing shoot and root growth due to division and elongation of cells and tissues, which is

reflected positively on the increment of lengths, diameters and weight of fruits (Islah and Ei-sayed, 2011, Doifode and Nandkar, 2014, Saeed *et al.*, 2014 and Tosic, *et al.*, 2016).

conclusion

In this study, biofertilizers showed a significant effect on most traits. Azotobacter and Azospirillum applications, and showed positive results in increasing the fruit yield of eggplant. Also, results showed that treatment of with biofertilizers transplants superior on most traits compared with the treatment of seeds coating biofertilizers.

Table (8) Effect of Biofertilizers and Methods of Addition on length and Dameter of Fruit of

Eggplant under the Greenhouse Conditions

Method of	Length of Fruit(cm)			Diameter of Fruit(cm)		
Addition A Bio Fertilizers T	Seeds A1	Transplant A2	Mean of Bio Fertilizers	Seeds A1	Transplant A2	Mean of Bio Fertilizers
Т0	7.3	7.3	7.33	3.81	3.81	3.81
T1	7.3	11.6	9.50	4.39	4.63	4.51
T2	9.8	12.9	11.40	4.60	5.00	4.80
T3	9.8	11.3	10.58	4.50	4.93	4.71
T4	10.3	12.5	11.42	4.70	5.06	4.88
Mean of a Method of Addition	8.9	11.1		4.40	4.68	
L.S.D 0.05	Method of Addition	Bio Fertilizers	Method of Addition Bio Fertilizers	Method of Addition	Bio Fertilizers	Method of Addition Bio Fertilizers
	1.14	1.80	2.55	0.23	0.37	0.53

T0 control T1 Pseudomonas fluorescence

T2 Azospirillum brasilense T3 Bacillus subtilis T4 Azotobacter chroococcum

References

AL-dulaimi, A, J.; Samer. M, A. and Musa, SH. A. (2003). Development and Evaluating Formula of Pseudomonas fluorescens. Iraqi Journal Agriculture (Special Issue) 8 (3), 104 – 110.

Al-haddad, M. E. M. (1989). The Role of Biofertilizers on Reducing Agricultural Costs, Reducing Environmental Pollution and Increasing Crop Production. National Training Course on the Production and Use of Biological Fertilizers. College of Agriculture, Ain Shams University. Egypt.

Allawi, M. M. (2013). Impact of Bio Organic and Chemical Fertilization on the Roots Architectural and Growth and Yield of Pepper plant (*Capsicum annuum* L.) Ph.D. Dissertation, Horticulture and Landscape Department. College of Agriculture., University of Baghdad, Iraq. P:168.

Al-mohammedi, F. M. H. (1990). Protected Agriculture. University of Baghdad - ministry of Higher Education and Scientific Research.

Al-naimi, S. N. A. (1999). Fertilizers and Soil Fertility. Dar Al Kutub For Printing & Publishing, University of AL Mousl Ministry of Higher Education and Scientific Research, Iraq.

Al-sahuki, M. M. and Wahib. K. (1990). Applications in the Design and Analysis of Experiments. Dar Al Hekma for Printing & Publishing. Mosul. University of Al Mosul. Ministry of Higher Education and Scientific Research. Iraq.

Al-zehawi, S. M. A. (2007). Effect of Manures and Mulching on Growth, Yield and Guality of Potato (*Solanum Tuberosum* L.) MSc Thesis. Department of Horticulture. College of Agriculture. University of Baghdad.p:69.

Anandaraj, B. and Delapierre, L.R.A. (2010). Studies on Influence of Bioinoculants (Pseudomonas Fluorescens, Rhizobium sp., *Bacillus megaterium*) in Green Gram. J. Biosci Tech. 1(2),95-99.

Dharmendra; S. (2014). Effect of Organic Manures and Bio Fertilizers on Growth, Yield and Quality of Tomato (*Solanum lycopersicum* L.). Degree of Master of Science (Agriculture). Department of Horticulture, College of Agriculture. Jawaharlal Nehru Krishi Vishwa Vidyalaya, Jabalpur.India.139.

Dheyab, N. S. (2012). Utilization of Rock and Super Phosphate and Addition of Fungal and Bacterial Bio-fertilizer on Growth and Yield Potato. PhD Thesis. Horticulture and Gardening Engineering Department / College of Agriculture-university of Baghdad. Iraq .150

Doifode, V. D. and P.B. Nandkar, (2014). Influence of Biofertilizers on the Growth, Yield and Quality of Brinjal Crop. Int. J. of Life Sciences. Special Issue A2,17 -20.

Islah; M, M and El-sayed, M.A.M. (2011). Response of Sweet Pepper Plant Growth and Productivity to Application of Ascorbic Acid and Biofertilizers Under Saline Conditions. Australian Journal of Basic and Applied Sciences, 5(6), 1273-1283.

Meena, M. L. V. S. Gehlot; D.C.; Meena, S.; Kishor; Kumar and J. K. Meena (2017). Impact of Biofertilizers on Growth, Yield and Quality of Tomato (*Lycopersicon esculentum* Mill.) cv. Pusa Sheetal. Journal of Pharmacognosy and Phytochemistry. 6(4), 1579-1583.

Minnotti, P. L.; D.E. Halseth and J. B. Sieczka. (1994). Chlorophyll Measurement to Assess the Nitrogen Status of Potato Varieties. Hortscience. 29(12), 1497-1500.

Naeem, S. D. (2012). Utilization of Rock and Super Phosphate and Addition of Fungal and Bacterial Bio-fertilizer on Growth and Yield Potato. PhD Thesis. Horticulture and Gardening Engineering Department/College of Agriculture-university of Baghdad. Iraq .150

Ramakrishnan, K. and G. Selvakumar. (2012). Effect of Biofertilizers on Enhancement of Growth and Yield on Tomato (*Lycopersicum esculentum* Mill.) International Journal of Research in Botany 2(4), 20-23.

Rao, K. M. P. K.; Singh, H.; Babiang, K.; Ryingkhun and B. Maying. (2014). Use of Bio-fertilizers in Vegetable Production. Indian Horticulture Journal; 4(1), 73-76

Richards, L.F (1954) Diagansis and Improvement of Saline and Alkaline Soils. Agric. Handbook, U.S.A., 60.

Saharan, B. S. and Nehra, V. (2011). Plant Growth Promoting Rhizobacteria: A Critical Review. Life Sciences and Medicine Research, Volume 2011, LSMR-21, p:1-29.

Salhia, B. M. (2010). The Effect of *Azotobacter chroococcum* as Nitrogen Biofertilizer on the Growth and Yield of Cucumis Sativus, Degree of Master of Biological Sciences/Botany the Islamic University, Gaza.

Salloum, Y. F. (2012). Effect of Adding Organic Matter on Growth and Production of Eggplant Plant and Accumulation of Anthocyanin Pigment in Fruits. MSc Thesis. Horticulture and Garden landscspe Engineering Department. University of Baghdad.111.

SAS, (2001). User Guide Statistic (Version 6-12). SAS Inst. Inst. Cary, N.C.USA.

Saeed, F. H.; Hadi M. A. and Kazem, D. H. (2014). The Detection of Auxin and Cytokine Hormons in Leachate Growth

of Some Biogenic Fertilizers. Women's Scientific Conference (Special Issue): pp. 186-192.

Saeed, F. H. (2015). Integrated Management of Chemical, Organic and Biological Fertilizers and Their Impact on the Growth and Productivity of Some Genotypes of Cucumber Plant. PhD Thesis. Horticulture and Gardening Engineering Department/College of Agriculture - university of Baghdad. Iraq .138

Shashi, K; M. Kumar. R. kumar and M. Raghav (2018). Effect of Biofertilizers on Growth and Yield of Tomato (*Lycopersicon esculentum* Mill). International Journal of Current Microbiology and Applied Sciences 7 (2), 2542-2545.

Tekalign, T and S.P. Hammes. (2005). Growth and Biomass Production in Potato Grown in the Hot Tropics as Influenced by Paclobut Razel. Plant Growth Regulation. Springer Netherland 45(1), 37-46.

Thenmozhi, R. K.; Rejina, K. Madhusudhanan and A. Nagasathya. (2010). Study on Effectiveness of Various Biofertilizers on the Growth & Biomass Production of Selected Vegetables. Research Journal of Agriculture and Biological Sciences, 6(3), 296-301.

Tosic, I; Z. Golic and A. Radosavac. (2016). Effects of the Application of Biofertilizers on the Microflora and Yield of Lettuce (*Lactuca sativa* L.) Acta Agriculturae Serbica, Vol. 21(42), 91-981.