## The Role of An Oil-Based Formulation for Protecting the Biocontrol Fungus *Beauveria bassiana* from Some Chemical Fungicides Under Labortory Conditions

Hussein Maktouf Diwan Iyad Qahtan Wahid Majed Ibrahim Abdullah Hussein Naima Kashmer Azhar Abbas Obaid

Ministry of Science and Technology/Agricultural Research Directorate
Baghdad-Iraq

E mail: hayderrashid70@gmail.com

#### **Abstract**

The lab study included evaluation of oil formulation to protect the conidia of Beauveria bassiana from two fungicides; beltanol and basten 50 and four insecticides; decis, quality 5, dynamec and supredimethoate after mixing and incubated for 1 and 9 days at relative humidity (RH) 0.0, 6, 33 and 75% and temperatures; 25 and 35 °C. Vitality percentage of conidia in aqueous suspension was negatively affected by recommended doses of the pesticides either the fungicides in which the vitality % of conidia kept at the lowest level (0.0%) over their exposure periods (1 and 9 Days) to the 25 and 35 °C under RH 0.0 and 6%. Not only that, but the insecticides had a negative effect under the same environmental conditions in which conidia recorded a vitality ranged 0.0-34.5%. On the other hand, vitality of the formulated conidia was more degraded under influence of extreme drought conditions (RH 0.0 and 6%) after mixing with a both fungicides over 9 days of exposing to the 35°C and was recorded 70 and 83.3% for 9 days of exposing to the 35 °C under RH 33% in compared with vitality of the fungicide-free formulated conidia. All the formulated conidia whether treated or untreated with chemical pesticides maintained their vitality in completely (100%) at all temperatures under RH 75%. These results have confirmed that it was not benefitted to use the oils only in protecting conidia from adverse physical and chemical environmental conditions, unless appropriate moisture was available, even at a low level (RH 33%).

Key words: Beauveria bassiana, Pesticides, Biocontrol, and Oil Formulation

دور المعاملة بالزيوت في حماية عامل المكافحة الحيوية الفطر Beauveria bassiana من بعض مبيدات الفطربات الكيميائية في ظروف المختبر

حسين مكطوف ديوان اياد قحطان وحيد ماجد ابراهيم عبد الله حسين نعيمة كشمر ازهار عباس عبيد وزارة العلوم والتكنولوجيا/دائرة البحوث الزراعية بغداد –العراق

الخلاصة

الكلمات المفتاحية: Beauveria Bassiana والمبيدات والمكافحة الاحيائية والمعاملة بالزيوت

## Introduction

The entomopathogenic fungus Beauveria bassiana (Deuteromycetes) has infected various types of insects. Lepidoptera (Moths and Butterflies) and Coleoptera (Beetles) were the most susceptible to this fungus and include many insects of agricultural economic importance inhabiting the soil or crops. The number of insects infected with this fungus have been estimated at more than 200 insects which were distributed among nine insect species (Storey and Gardner 1986; Li and vang, 1988). The fungus has been developed commercially produced to control many agricultural insect pests (Amatuzzi, et al., 2017). B. bassiana has not cause any harmful effect on the environment components in compared with chemical pesticides (Sayed, et al., 2019), and it has been in a wide range of use due to its parasitic ability to most harmful insects that belong to different insect groups. The effectiveness of B. bassiana and its tolerance to environmental conditions were determined by its direct use as spore aqueous formulations against a specific insect pest (Prior, et al., 1988; Moslim, et al., 2004). It is worth noting that these formulations, which include biological control fungi, among them B. bassiana, remain in the place of developing and improving their biological performance based on the results achieved and the various environmental conditions including chemical ones that were exposed to them as these bio-pesticides were measured by the stability of their commercial use, and as far as provides positive results and rich economic returns. The presence of B. bassiana naturally in agricultural fields (Uma Devi, et al., 2008) or artificially released for biological control or integrated pest management made their spores susceptible to the chemical pesticides, either at the beginning of spraying or after that (Challa and Savinda, 2014) which leads to kill or inhibit their germination (Alves, 1998; Anderson and Roberts, 1983; Loria, et al., 1983; Benz, 1987), this eventually leads to the failure of the entire biological control. The success of using entomopathogenic fungi biocontrol or integrated programs for controlling the pest insect depended on the long survival of spores in the environment conditions (Benz, 1987) adapted from (Reddy, et al., 2018). Undoubtedly, the use of environmentally friendly oils or those compatible with bio-control fungi in the bio-products is one of the main means of protecting them from harsh environmental conditions (inglis, et al., 2002; Sangamithra, et al., 2015). Although the positive properties of the lacking water by the toxicity. availability, cheap price, potential for using in bio-preparation and spraying by simple hydraulic pumps, a several previous studies have emphasized importance of an oils in bio-preparations in compared with the water enhancement the work of fungi for insect pest infection, including the fungus B. bassiana (Prithiva, et al., 2017; Sridevi, et al. 2018; Sangamithra, et al., 2015). Chemical pesticides are still the first line in combating insect pests and fungi cause plant diseases in the short and medium term, as the use of chemical pesticides have been remained main strategy for agricultural pest control, giving farmers an opportunity to obtain large quantities of agricultural crops in good quality and at a lower cost, as the use of chemical pesticides may have inhibitory effects on natural enemies but in often leads in killing it (Bueno. et al..2017). Fungicides and some insecticides have toxic effects on large numbers of their entomopathogens (De Oliveira and Neves, 2004).

This study objectived to evaluate the effect of oil formulation on protecting the

fungus *B. bassiana* from two fungicides; beltanol and basten 50 and four insecticides decis, quality 5, dynamic and superdimethoate at 25, 35°C under RHs; 0.0, 6, 33 and 75% after 1 and 9 days of exposure.

## **Materials and Methods**

## The Fungus Beauveria bassiana

The fungus B. bassiana isolate X5 obtained from the bio-pesticides department, former Iraqi atomic energy organization is still used by us in the bacterial pesticide laboratory, biological control department of plant diseases, the integrated pest control center, the agricultural research directorate. The fungal isolate was refreshed by planting it in the medium potato extract dextrose agar (PDA), which was previously prepared and sterilized at the autoclave at a 121 °C/1.5 Kg/cm<sup>2</sup> for 20 minutes and then placed inside sterile 9 cm petri dishes and incubated at 25 °C for seventeen days to form colonies and complete growth and production of many spores. The dishes were kept in the refrigerator for two weeks for the purpose of using them in carrying out the experiments.

## **Inoculum Preparation of The Fungus** *B. Bassiana*

The inoculum of B. bassiana (Isolate 5x) was prepared for many times as needed. Several glass bottles (size 200 ml) were used in this preparation. A rice boiled with distilled water was added into these bottles at 30 g/ bottle and were plugged with cloth stoppers. The glass bottles were sterilized in an autoclave at 121 ° C/ 1.5 Kg/ cm<sup>2</sup> for 15 minutes. After sterilization, the glass bottles were cooled and inoculated with plugs of the fungal culture was kept on refrigerator using one plug/ bottle. The bottles were incubated for two weeks at 25 °C with continuous manual shaking until completing fungal growth. The bottles were left two weeks under lab conditions for more sporulation and fungal cultures were dried. In order to collect the flying spores' dust, an empty and sterilized glass bottle of the same size was placed inverted so that the openings of the two bottles coincided, as they were shaken together vigorously and simultaneously to stir up the spore dust, and it took five minutes to collect the dust. To obtain a suitable aqueous spore suspension, 100 ml of sterile distilled water was added. Several spores were counted using a haemocytometer and a light compound microscope. The concentration of the spores was calculated using the following equation.

# Evaluation of the Efficiency of Oil Formulations in Protecting *B. Bassiana* Spores from Chemical Pesticides Under Temperatures, and Different Exposure Periods

To evaluate the efficacy of some oil formulations in protecting B. bassiana spores from some chemical pesticides, the two fungicides were used betanol (Chinasol, Spanish) and basten 50 SC Chinese) (Carbendazim, and four insecticides; Quality 5, Superdimethoate (a.i. is Dimethoate, Ugandan), decis 2.5 EC (a.i. is Deltamethrin, German) and dynamec (a.i. is Abamectin, Manufacturer is Syngenta). We followed the methods described by Wyss (2004), Lopes, et al., (2014), Sangamithra, et al., (2015), Perez-Gonzalez and Sanchez-Pena (2017). The corn oil (20%) and Tween 20 (2%) were used with distilled water and a number of formulations were prepared after mixing with B. bassiana spores (6.6 x 105 spores/ml) that were previously prepared and dried in the sterile dishes supplied with hand-made sterile cardboard covers at rate 1ml / dish as above-mentioned method, and three replicates (Dishes) were used for each treatment. These formulations contained distilled water with Tween 20 (TW), corn oil with Tween 20 (COT), and spores with distilled water. pesticides were prepared by concentration (X) according to the instructions of the companies produced. while a double concentration (2X) was prepared, the pesticides as concentration (X) were added to the aforementioned formulations at 1: 1 (v/v) in each dish base, they were mixed under sterile conditions using sterile microscopic slides, and the following treatments were obtained : corn oil +tween20+ spores + beltanol (Co +Tw+ Sp + Belt), corn oil +tween20 +spores + basten 50 (Co + Tw + Sp + Bast), corn oil + tween20+spores + Quality 5 (Co+ Tw + Sp + Qual), corn oil+tween20 + spores + superdimethoate (Co + Tw+Sp + Supe), corn oil +tween20+ spores + decis (Co + Sp + Dec), corn oil+tween20 + spores + dynamec (Co + Tw+ Sp + Dena), It is also the pesticides with double concentration (2X) were put onto dishes containing distilled water at a rate of ml/ plate. In addition, pesticides untreated formulations; corn oil + Tw 20 + spores (Co + Tw + Sp), spores + distilled water + Tw, and spores + distilled water were used as control treatments, those combinations were added onto dishes at a rate of 2 ml/dish. Three replicates (Dishes) were used for every treatment and the dishes were distributed in the two incubators at 25 °C, 35 °C. These dishes were divided into two groups, where the first group was incubated for 1 day, and another for 9 days of exposing. After the exposing period ended, the dishes were removed from the incubators and the viability percentage of spores was calculated as described by Lacey (1997).

## Statistical analysis

The obtained results were statistically analyzed using the Ready-genstatistical Program. In the statistical software package, a calculation of the mean

vitalities of three replicates of each treatment, standard error of each mean (S.E.M.) and standard deviation (SD) of the tested variables were determined. A T-test was used to assess individually the significant difference of vitality% between chemical treated and untreated formulated conidia within one column under the 25 or 35 °C, RH, and one period of exposure. We used analysis of variance (ANOVA) at one-way ANOVA (No Blocking). On the other hand, a Ttest was applied to assess the significant the difference of vitality% between every treated chemical and untreated formulated spores within every row which reflected the impact of interaction of all different temperatures, RHs after periods of exposure using Two-way ANOVA (No Blocking). Lowest significant difference (LSD) was used at (P = 0.05) assessment between the treatments, whereas the value had probability less < 0.05 was more significant.

Evaluation of the Efficacy of Oil-Formulations in Protecting the *B. Bassiana* Spores from Some Chemical Pesticides Under Different Temperatures and Different Exposure Periods

Table (1) reveals the results of all the pesticides that killed spores of fungus B. bassiana in which spores recorded vitality 0.0% except superdimethoate vitality of spores showed superiority (28%) after one day of treatment at 25 °C. Spores showed high vitality (34.5%) after 9 days of treatment with the decis at 25 °C in compared with the other chemical treatments. Spores were treated with qality 5 registered high level of vitality (19.7%) after one day of incubation at 35 °C related to other chemical-treated spores.

In the Table (2), the results discovered that the formulated spores were superior (p = 0.05) in their keeping vitality in

which ranged 91.1% (178.1 Colony) to 100% (193.3 Colony) after 1 day of exposing to the 25 ° C under RH 0.0 % either with or without pesticides presence in compared with their vitality which ranged 0.0% (0.0 Colony) - 80% (154.64 Colony) after 9 days of exposing to the 25 ° C, 9.1% (17.6 Colony) - 98% (189.4 Colony) after 1 day of exposing to the 35 °C, and 0.0% (0.0 Colony) to 72.4% (139.94 Colony) after 9 days of exposing to the 35 °C. The spores of control treatment (Spores Only + Distilled Water) recorded a verv significant reduction in their vitality was ranged 0.0% (0.0 Colony) to 42.5% (Colony) over all the heat exposure periods. Spores of the formulations (Co+Tw) showed non-significantly differences in their vitality after 1 and 9 days of exposing to the 25 °C related to the vitality of all chemical-treated spores except to the formulated spores treated with beltanol and superdimethoate in which vitality amounted 91.1 and 0.0%, respectively. Formulated spores recorded a high percentage of vitality under RH 6%. Vitality revealed a relative rise after 9 days of exposing to pesticides at 25 °C which ranged (71.4 – 100 %), but it decreased when it was exposed to the 25 °C for the same period, whereas reached 36.7 - 77.1 %. Vitality was significantly (p = 0.05) declined after 1

day of exposing to the beltanol and basten 50 at 35 °C and the percentage was 25.9 and 39%, respectively. While the vitality remained steadfast after 1 day of exposure to the insecticides at 25 °C (93.4 – 100%). Formulated spores were recorded full vitality (100%) under RH 33% after 1 and 9 days of exposing to the two temperatures, but they recorded 70 and 83.3% after 9 days of exposinf to the beltanol and basten 50, respectively at 35°C.

Table (1) The effect of fungicides and insecticides on vitality% of fungus Beauveria bassiana under two exposure periods.

A RH 6% caused increase in the superdimethoate-treated vitality of formulated spores after 1 and 9 days of an exposing to the 25 °C, as it reached 96 and 66.6%, respectively. On the other hand, the results showed that a raising the RH to 33% led to an increase in the level of vitality (100%) of the formulated spores after 1 and 9 days of treating with pesticides and exposing to 25 and 35 °C, beltanol-treated formulated spores that recoded 90 and 75 % after 9 days of exposure to the 25 and 35 °C, respectively. All the pesticides-treated formulations provided completely protection from thermal and chemical effects using RH 75% in which spores achieved vitality 100%.

Table (1) The Effect of Fungicides and Insecticides on Vitality% of Fungus *Beauveria bassiana* Under Two Exposure Periods

	Temperature					
Treatment	25	o <sub>C</sub>	35 °C			
	1 Day	9 Days	1 Day	9 Days		
Distilled Water + Conidia (Control)	42.5	0.0	37.5	0.4		
Beltanol	0.0	0.0	0.0	0.0		
Basten 50	0.0	0.0	0.0	0.0		
Decis	0.0	34.5	3.7	2.4		
Quality5	0.0	0.0	19.7	0.0		
Dynamec	0.0	2.5	0.0	0.0		
Superdimethoate	28.3	0.0	0.0	0.0		

Table (2) Vitality Percent of Formulated and Unformulated Spores of Fungus *Beauveria Bassiana* at Different Chemical and Thermal Conditions

Treatment	Standard Number of Colonies = 180 Colony or Standard Vitality% = 100 RH 0.0%				Standard Number of Colonies = 170 colony or Standard Vitality% = 100 RH 6%			Standard Number of Colonies = 122.5 Colony or Standard Vitality% = 100 RH 33%				
												25 °C
	1 Days	9 Days	1 Days	9 Days	1 Days	9 Days	1 Days	9 Days	1 Days	9 Days	1 Days	9 Days
		Vitality	Vitality	Vitality	Vitality	Vitality	Vitality	Vitality	Vitality	Vitality	Vitality	Vitality
	%	%	%	%	%	%	%	%	%	%	%	%
Co+tw	100	80	80	72.4	100	100	100	100	100	100	100	100
Co+tw+bel	91.1	72.9	9.3	5.1	82.3	83.5	25.9	36.7	100	100	100	70
Co+tw+bas	92.4	68.9	9.1	4.6	100	75.5	39	41.42	100	100	100	83.3
Co+tw+dec	100	67.8	98	58.5	94.3	100	96.6	61.8	100	100	100	100
Co+tw+qu a	100	67.6	95	59	100	83.3	100	73.3	100	100	100	100
Co+tw+dy n	99.2	78	85.7	78	81.7	100	93.4	77.1	100	100	100	100
Co+tw+sup	96	0.0	76.6	0.0	100	71.4	95	66	100	90	96.7	80
1.s.d* at P = 0.05	8.23	20.26	11.16	8.24	4.98	5.18	8.4	13.53	0.0	6.62	2.82	10.12
1.s.d1 at P = 0.05	10.1	25.7	26.9	12.8	12.1	10.3	27.6					
1.s.d2 at P = 0.05	7.19	11.8	12.5	16.7	16.4	10.2	35.3					
l.s.d3 at P = 0.05	7.2	35.6	34.3	11	7.2	8.2	9.5					

- L.s.d\* between treatments under thermal period and RH
- L.s.d1 between periods of thermal exposure under different RHs
- L.s.d2 of thermal -humidity interaction for every exposure period
- L.s.d3 between periods of humidity exposure under every temperature

## **Results and Discussion**

Generally, all the tested fungicides and insecticides used in this study showed their negative effect on the viability of B. bassiana spores after treated directly with them according to the recommended concentrations; 1.5, 2.5, 1.25, 1, 1, and 2 ml/ L. for beltanol, basten 50, desic 2.5 ES, quality 5, dvnamic. and superdimethoate, respectively. The fatal effect (100%) of the basten 50 on spores was into line with the findings of Deb, et al., (2017) during their study the effect of some chemical fungicides, one of them was bavistin (Carbendazim 50 WP) germination of B. bassiana spores as it caused significant reduction percentage (0.0%) after 7 days of treatment with all concentrations of the fungicide at a 25 °C. In relevant to this study, (Challa and Savinda, 2014) explained that the complete inhibition (100%) in germination spore was resulted of susceptibility of the spores of B. bassiana to the active ingredient (Carbendazim), which constitutes 50% of this fungicide, as the spindle

formation disrupted in one stage of the mitosis, so it prevents the emergence of germination tube and the appresorium adhesion and thus the mycelium of the fungus was not formed. As for the negative effect of beltanol that had contributed to the viability declining (0.0%) did not differ from counterpart (Basten 50), only mechanism for killing the spores of the fungus B. bassiana differed in its influential role due to chinosol (8-Hydroxyquinoline Sulfate; 71.42% Hydroxyquinoline and 24.15% Sulfate) which was believed to have a chelating role in extracting minor essential elements from the surface of any fungus and thereby negatively affecting the balance of the mineral elements of this organism which inhibits their growth (EMEA, 1998), and this may applied to the fungus B. bassiana. In fact, there are no previous studies that dealt with the effect of beltanol on entomopathogenic fungi but confined to phytopathogenic fungi. The treatment of spores with the insecticide superdimethoate, significantly caused in their vitality reduction which

amounted to 28.3% after 1 day of exposure to the temperature 25 °C. These results were disagreed with findings of some previous studies that showed the compatibility of some insecticides with the spores of the fungus B. bassiana at different concentrations depending on the isolates were used. De Oliveira and Neves (2004) had studied the effect of three concentrations (three concentrations, half MC and twice MC) of 12 acaricides; Abamectin (Dynamec), Dimethoate (Superdimethoate) incorporated in different formulations on some growth parameters of B. bassiana, found that the Abamectin was had moderate compatibility with the spores. where germination was (53.9%), after 1 day of incubation at the 25 °C, while the dimethoate was incompatible with the spores as they attributed compatibility variation to the different chemical nature of these pesticides and to the concentrations were used. On the whole, many of authors have explained in their previous studies the main possible reasons behind the inhibition of some insecticides for the B. bassiana spores, which may be due to the complete obstruction of the biological pathways responsible for metabolism of fungal spores (Moore-Landecker, 1990) or to the action of some chemical (Ionic or Molecular) pesticides in the equation of the electrostatic charges of the mucosal coated layer of spores or the removal of those charges from them, which an imbalance in the communication of instructions or signals responsible for stimulating germination (Boucias, et al., 1988). Our results were also consistent with the findings of (Filho, et al., 2001) they have confirmed the incompatibility of Decis® 25 CE (it's a.i. is), as the fungus recorded a significant decrease in colonies form units (0.0 Colony) after 1 day of treatment at 30 ° C, they gave no explanation for the reason for the

deltamethrin decline in this growth parameter except that they noted what Morris (1977) showed that the pesticides belonging to the pyrethrins group had an highly bacteriostatic of the bacteria parasiting insects. RH and temperature are important environmental factors that determine longevity ofentompathogenic fungi and efficiency for infection the insect pests (Lazzarini, et al., 2006; Inyang, et al., 2000). Corn oil has fallen short in efficacy under the conditions of interference between RH 0.0% and two periods (1 and 9 Days) of exposing spores to the temperature 35 °C in the presence of chemical pesticides. The benefit of corn oil in protecting B. bassiana spores from temperature 35 °C and RH 0.0% may take a few hours followed by the damage spores, whereas a one-day exposure period to those adverse environmental conditions was sufficient to kill or inhibit their vitality. Adverse effect of pesticides on the spore's vitality, may be increased due to the rising in the pesticides concentrations as a result of water evaporation at a high temperature (35  $^{\circ}$ C) and low RH (0.0%). interpretation supported This Franklin, 2007 article explained that the high temperature and low RH will make the water in the pesticide droplets evaporated quickly, reducing the droplet size of the bush pesticide 2, 4-D, and the ester volatilization rate of the pesticide will had been accelerated three times after heat rising from 60 to 80 Fahrenheit. The corn oil had more instrumental when the RH was available. even if it was at a scarce level (6%). The long-term exposure (9 Days) has a bad effect on the spore vitality, despite the temperature was ideal (25 °C) at RH 0.0%. With the presence of these pesticides, as perhaps this period under these conditions is sufficient to volatile the hydrocarbons of the oil, which leads to an increase in its density and viscosity, and consequently, the dissolution of the

oil or its cracking into smaller units (Degrémont, 1991), which may allow the pesticides to enter the spores . This study showed that the benefit of the corn oil is limited to a specific time and that its positive role depends on the availability of RH. Increasing RH 0.0% to 6% to 33% helped spores for keeping their survivalability at highest attainable standard in all thermal and chemical periods, as the RH can alleviate a high temperature by evaporative cooling (Schroeder, 2000), lowering the amount of oil evaporation, and retardation its dehydration, as the oil has a slowest evaporation rate of water (Fingas, 2012). With this mechanism the RH at different levels maintained the physical properties of the corn oil and thus the oily film coated the spores acted as a barrier preventing the pesticides from crept into the spores. Besides the positive role of corn oil in maintaining vital of B. bassiana spores from extreme conditions (RH 6% and High Temperature 35 °C), water vapor surrounded spores worked as enhancement factor in mitigation of heat Intensity (Arauz and Sutton, 1989). Concerning the reduction in spores vitality of the control treatment (Only + Distilled Water), it was much more likely to the adversely impact of the water imbibition on the dry spores, as (Faria, et al., 2010) found that a high percentage of spores of B. bassiana that had already been exposed to environmental stress showed very slow germination, but lethal significant impact, as a result of hypersensitivity to the imbibition, in compared with the non-stressed spores (Meaning Spores That Were Prepared in the Form of an Aqueous Suspension from Fungal Colonies and Used Without Going Through The Drying Stage). showed a large germination rate on the PDA, only 2% were destroyed after 24 and 72 hours of incubation at 25 °C, while stressed- spores recorded a low percentage of germination (14 and 44%)

after 24 and 72 hours of incubation at a 25 °C, respectively, and the shattered of them reached more than 40%. On the contrary, spore of the formulation corn oil + tween 20 achieved a vital percentage of 100% in the first period (One Day Only) of exposure to the 25 °C at RH 0.0%, and the first and second periods of exposure to the 25 and 35 °C at RH 6% spores were also recorded 100% vitality. Mixing the oil- treated dried spores with the added distilled water has staved them off direct contact with water, avoiding the spores from destruction effect of imbibition. Paraffin oil prevented spores of imbibitional damages, and only 40% of them were destructed. This result consistent with the Faria's finding (2009), that the oil-based formulation supplied with the paraffin oil protected the spores from imbibition damage. Formulation of conidia in pure (Non-emulsifiable) paraffinic provided considerable protection from imbibitional damage. This underscores a need for establishing standard protocols for preparing aqueous suspensions of sensitive fungi for both research and commercial. Referring to these results, corn oil was the better oil in inducing the B. bassiana spores to germination and forming fungal colonies. And this vegetable oil has an alleviate the suffering of these spores from the chemical fungal pesticides hazards used in this study and mitigate the early drought as a result of exposing spores to the high temperature. Vitality of the fungus spores is directly proportional to the increasing RH exposed it. Because of corn succession in protecting the B. bassiana spores from the used chemical pesticides, we recommend expanding the test to the largest possible number of chemical pesticides preferably those are more important and commonly used in Iraq.

## References

**Alves,** S. B., and Lecuona, R. E. (1998). Epizootiologia Aplicada Ao Controle Microbiano De Insetos. Controle Microbiano De Insetos, 2, 97-169.

Amatuzzi, R. F.; Cardoso, N.; Poltronieri, A. S.; Poitevin, C. G.; Dalzoto, P.; Zawadeneak, M. A., and Pimentel, I. C. (2017). Potential of Endophytic Fungi as Biocontrol Agents of Duponchelia Fovealis (Zeller) (Lepidoptera: Crambidae). Braz. J. Biol. 78, 429-435.

Anderson T. E and Roberts D. W. (1983). Compatibility of *Beauveria bassiana* Isolates with Insecticide Formulation Used in Colorado Potato Beetle (Coleoptera: Chrysimelidae) Control. J Econ Entomol. 76, 1437-1441 Arauz, L. F. and Sutton, T. B. (1989). Influence of Temperature and Moisture on Germination of Ascospores and Conidia of Botryosphaeria Obtusa. The American Phytopathological. 79, 667-674.

**Benz**, G. Environment. (1987). In: Fuxa, JR; Tanada, Y. (Eds), Epizootiology of Insect Diseases, New York, Wiley, 177-214.

**Boucias**, D. G.; Pendland, J. C.; Latge, J. P. (1988). Nonspecific Factors Involved in Attachment of Entomopathogenic Deuteromycetes to Host Insect Cuticle. Appl. Environ. Microbiol., 54,1795-1805.

**Bueno**, A. F; Carvalho, G. A.; Santos, A. C.; Sosa-Gomez, D. R.; and Silva, D. M. (2017). Pesticide Selectivity to Natural Enemies; Challenges and Constraints for Research and Field Recommendation. Cien. Ciência Rural, 47 (6), 1-10.

Challa, M. M. and Savinda, S. k. (2014). Compatibility of *Beauveria bassiana* (Bals.) Vuill. Isolates with Selected Insecticides and Fungicides at Agricultural Spray Tank Dose. Innovare

Journal of Agricultural Science; 2(3),7-10

**Deb**, L.; Rajesh, T.; Majumdar, D., and Tombisana R. K. (2017). Evaluation of Biological Compatibility of *Beauveria bassiana* with Fungicides and Botanicals. J. Pharmacol. and Phytoche. SP1, 1120-1124.

**Degremont**, Water Treatment Handbook, 6th ed., Halsted Press, New York, NY, (1991). Fair, Geyer and Okun, Water and Wastewater Engineering, John Wiley & Sons, New.

**De Oliveira**, R. C.; P. M. O. J. Neves. (2004). Compatibility of *Beauveria bassiana* with Acaricides, Neotrop. Entomol. 33(3), 353–358.

**EMEA**/MARL/464/98-FINAL. (1998). The European Agency for the Evaluation of Medicinal Products. Veterinary Medicines. Committee for veterinary medicina products 8-hydroxyquinoline, pp.1-4.

Faria, M.; Hotchkiss, J. H.; Hajek, A. E., and Wraight, S. P. (2010). Debilitation in Conidia of the Entomopathogenic Fungi *Beauveria bassiana* and Metarhizium Anisopliae and Implication with Respect to Viability Determinations and Mycopesticide Quality Assessments, J. Invertebr. Pathol., 105(1), 74-83.

Faria, M. R. (2009). Studies on Entomopathogenic Fungi: Evaluation of Germination Protocols for Assessing Conidial Quality and Modified Atmospher Packaging for Enhancing High-temperature Shelf Life. Ph.D. dissertation, Faculty of the graduate school of Cornell University, pp.160.

**Filho**, B. A.; ALMEIDA, J. E. M.; and LAMAS, C. (2001). Effect of Thiamethoxam on Entomopathogenic Microorganisms. Neotrop. Entomol. 30(3), 437-447.

**Fingas**, M. F. (2012). Studies on the Evaporation Regulation Mechanisms of

Crude Oil and Petroleum Products. Advan. Chem. Eng. Sc., 2, 246-256.

**Inglis**, D. G.; Jaronski, S. T.; Wraight, S. P.; Beattie, A.; Watson, D.; Stevens, M.; and Rae, D. (2002). Use of Spray Oils with Entomopathogens. Spray Oils Beyond 2000: Sustainable Pest and Disease Management, 302-312.

Inyang, E. N.; McCartney, H. A.; Oyejola, B. A. and Ibrahim, L. (2000). Effect of Formulation, Application and Rain on the Persistence of the Entomogenous Fungus Metarhizium Anisopliae on Oilseed Rape. Mycolo. Res., 104 (6), 653-661.

Lacey, L. A. (1997). Manual of Techniques in Insect Pathology. Academic Press, New York, 410pp.

Lazzarini, G. M. J.; Rocha, L. F. N.; Luz, C. (2006). Impact of Moisture on in Vitro Germination of Metarhizium Anisopliae and *Beauveria bassiana* and their Activity on Triatoma Infestans. Mycol. Res. 110, 485-492.

Li, Y. W., and Yang, J. H. 1988. Prospects for the Use of Entomopathogenous Fungus Against Forests Pests. In: Study and Application of Entomogenous Fungus in China, Li, Y. W., Li, Z. Z., Liang, Z. Q., Wu, J. Wu, Z. K., and Xu, Q.F. ed.). Periodical Press, Beijing, 10-14 pp.

**Lopes**, R. B.; Pauli, G.; Mascarin, G. M. and Faria, M. (2014). Protection of Entomopathogenic Conidia Against Chemical Fungicides Afforded by an Oil-Based Formulation. Biocon. Sci. and Technol., 21(2), 125–137.

**Loria**, R.; Galaini, S.; and Roberts, D. W. (1983). Survival of Inoculum of the Entomopathogenic Fungus *Beauveria bassiana* as Influenced by Fungicides. Environ. Entomol. 12,1724-1726.

**Moore**-Landecker, E. (1990). Fundamentals of the Fungi (3rd Edition). Prentice Hall, Englewood Cliffs, new Jersey, p. 300.

Moslim, R.; Wahid, M. B.; Ali, S. R. A. and Kamarudin, N. (2004). The effect of Oil on Germination of *Beauveria bassiana* (Balsamo) Vuellimen and its Infection Against the Oil Palmbag Worm, Metisa plana (Walker) J. of Oil Palm Res. 16 (2), 78-87.

**Perez-** Gonzalez, O. and Sanchez-Pena, S. R. (2017). Compatibility in Vitro and in Vivo of the Entomopathogenic Fungi *Beauveria bassiana* and Hirsutella Citriformis with Selected Insecticides. Southwestern Entomologists, 42(3), 707–718.

**Prior**, C.; Jolland, P. and Patourel, Le. (1988). Infectivity of Oil and Water Formulations of *Beauveria bassiana* (Deuteromycotina: Hyphomycetes) to the Cocoa Weevil Pest Pantorhytes Plutus (Coleoptera: Curculionidae). J. Invertebrate. Pathology, 52, 66-72.

**Prithiva**, J. N.; Ganapathy, N. and Jeyarani, S. (2017). Efficacy of Different Formulations of *Beauveria bassiana* (Bb 112) Against Bemisia Tabaci on Tomato. J. Entomol. Zool. Stud., 5(4), 1239-1243.

**Reddy**, D. S.; Reddy, M. and Pushpalatha, M. (2018). Interaction of Fungicides with Bio-control Agents. J. Entomology and Zoology Studies.6(4), 545-551.

**Roberts**, D. W., and Campbell, A. S. (1977). Stability of Entomopathogenic Fungi. Misc. Publ. Entomol. Soc. Am.10, 19-76.

**Sangamithra**, S.; Jeyararani, S. and Ramaraju, K. (2015). Compatibility of Different Oils with *Beauveria bassiana*, A Potential Entomopathogenic Fungus. The Biocs. 10 (3), 1113–1117

**Sayed**, S. M.; Ali, E. F. and Al-Otaibi, S. S. (2019). Efficacy of Indigenous Entomopathogenic Fungus, *Beauveria bassiana* (Balsamo) Vuillemin, Isolates Against the Rose Aphid, Macrosiphum Rosae L. (Hemiptera: Aphididae) in Rose Production. Egyp. J. Biol. Pest Control., 29(19), 2-7.

Schroeder, D. V. (2000). An Introduction to Thermal Physics. Weber State University. Copyright, 2000, San Francisco, CA: Addison- Wesley Publishing Company, 214pp. P36.

**Sridevi**, K. M.; Jeyarani, S. J.; Ramaraju, K. (2018). Evaluation of Oil-based Formulation of Beauveria bassiana (Bb 112) (Bals.) Vuill. and Delivery Methods for the Management of Chilli Thrips, Scirtothrips dorsalis Hood. J. Biol. Control; 32(1), 62-67.

**Storey**, G. K., and Gardner, W. A. (1986). Sensivity of the Entomopathogenous Fungus *Beauveria bassiana* to Selected Plant Growth Regulators and Spray Additives. Appl. Env. Microbiol., 52(1), 1–3.

Uma Devi, K.; Padmavathi, J.; Uma, M. R. C.; Akbar A. K. P. and Murali M. C. (2008). A Study of Host Specifity in the Entomopathogenic Fungus, Beauveria bassiana (hypocreales: Clavicipitaceae). Biocontrol Sci. Tech. 18, 975–989.