

# Role of curcumin in prevention of cisplatin-induced nephrotoxicity in rat

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## Abstrac

Cisplatin is a platinum anticancer drug approved by the FDA in 1978. It is used to treat a broad spectrum of malignancies but the amount of drug necessary to produce a significant reduction in tumor burden usually produces significant nephrotoxicity. the purpose of this study was to evaluate the role of curcumin in the prevention of cisplatin-induced nephrotoxicity in rats. Thirty male albino rats weighing (200-250 gm), they were equally divided into three groups: Normal group (Isotonic saline group), control group "Cisplatin group" (12 mg/kg cisplatin, single dose I.P.) and curcumin group (50 mg/kg I.P.) prophylactically one day prior to cisplatin administration and continued for further 3 days. Cisplatin administration resulted in a significant increase in serum creatinine, BUN and MDA while serum glutathione level was significantly decrease compared to that of normal group. The Sections of kidneys rats treated by cisplatin were showed degenerative and necrosis of the proximal and distal convoluted tubules. Curcumin administration resulted in a significant decrease in serum creatinine, BUN and serum MDA while serum glutathione level was to be significantly increased. There were very minimal degenerative changes in kidney tissues of rats comparing to that of control group.

## الخلاصة

السيبيلاتين هو علاج كيميائي فعال يستخدم في معالجة طيف واسع من الاورام السرطانية لكن المشكلة في استخدامه تكمن في ان الجرعة العلاجية قريبة من الجرعة التي تسبب التسمم وان كمية الدواء الضرورية لاجداث تاثير علاجي عادة تؤدي الى حدوث تلف كلوي. اجريت هذه الدراسة على الجرذان لتقييم دور الكركمين في حماية الكلى من التلف الكلوي الذي يسببه دواء السيبيلاطين. تم استخدام 30 جرذ اوزنهم تتراوح بين ( 200-250غم) وقسموا بالتساوي على ثلاث مجاميع: مجموعة النورمال سلاين اعطيت عن طريق البريتون ( 0,2 مليلتر نورمال سلاين) و مجموعة السيبيلاطين (اعطيت جرعة واحدة 12 ملغم/كغم) من السيبيلاطين

عن طريق البريتون ومجموعة الكركمين والتي حقنت لاربعة ايام بالكركمين وحقنت بجرعة واحدة من السيبيلاطين في اليوم الثاني ( 12 ملغم/كغم) عن طريق البريتون . اظهرت النتائج ان اعطاء السيبيلاطين نتج عنه زيادة ملحوظة في مستوى الكرياتنين واليوريا والمالوندايديهايد وحصول نقصان ملحوظ في مستوى الكلوتاثيون واظهرت النتائج عدم حدوث تغيير ملموس في مستوى البروتين (الالبومين و الكلوبيولين) في المصل بالمقارنة بمجموعة النورمال سلاين، كما ان المقاطع النسيجية لكلية الجرذان اظهرت تلف كلوي ناتج عن اعطاء السيبيلاطين. كما اظهرت النتائج ان اعطاء الكركمين نتج عنه نقصان ملحوظ في مستوى الكرياتنين واليوريا والمالوندايديهايد وحصول زيادة ملحوظة في مستوى الكلوتاثيون. بالاضافة لذلك فان اعطاء الكركمين قلل التلف الكلوي الذي يسببه السيبيلاطين بشكل ملحوظ وكبير. واظهرت النتائج عدم حدوث تغيير ملموس في مستوى البروتين (الالبومين و الكلوبيولين) في المصل بالمقارنة بمجموعة السيبيلاطين. يمكن الاستنتاج ان الكركمين ضمن الجرعة المستخدمة له دور في حماية الكلى من التلف الكلوي الذي يسببه السيبيلاطين.

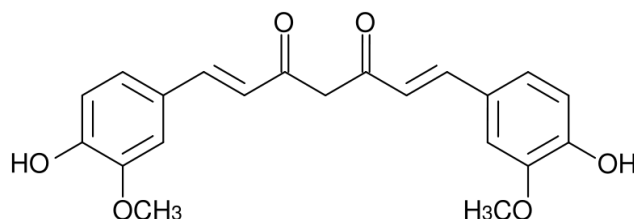
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## Introduction

One of the great success stories in the field of cancer chemotherapy is that of cisplatin a curative treatment for testicular tumors, in testicular cancer, the drug reaches greater than 90% cure rates, approaching 100% in early stage cases <sup>(1)</sup>. Cisplatin usually in combination with other drugs is commonly used as first line chemotherapy against cancers of the testis, lung, head-and-neck, esophagus, stomach, colon, bladder, ovaries, cervix, uterus and as second line treatment against most other advanced cancers, such as cancers of the breast, pancreas, liver, kidney, prostate as well as against glioblastomas, metastatic melanomas, and peritoneal or pleural mesotheliomas <sup>(2)</sup> and it has synergistic cytotoxicity with radiation and other chemotherapeutic agents <sup>(3)</sup>. The therapeutic effects of cisplatin are significantly improved by dose escalation. However high-dose therapy with cisplatin is limited by its cumulative nephrotoxicity and neurotoxicity <sup>(4)</sup>. The mechanism of action of cisplatin is similar to that of the alkylating agents; cisplatin enters cells by diffusion and by an active  $\text{Cu}^{2+}$  transporter and inside the cell, the chloride atoms of cisplatin are replaced by water, yielding a positively charged molecule that reacts with nucleophilic sites on DNA and proteins. Aquation is favored at the low concentrations of  $\text{Cl}^-$  inside the cell and in the urine. The platinum complexes can react with DNA, forming both intrastrand and interstrand cross-links <sup>(5)</sup>. The resulting cytotoxic lesion inhibits both DNA replication and RNA synthesis <sup>(3)</sup>. Severe and irreversible damage to the kidney remains the most important complication of cisplatin treatment as it may limit further treatment or even threaten life <sup>(6)</sup>. The efficacy of cisplatin is dose dependent, but the significant risk of nephrotoxicity frequently hinders the use of higher doses to maximize its antineoplastic effects <sup>(2,4 and 7)</sup>. Cisplatin gets accumulated in the tubular epithelial cells of proximal kidney tubule, causing nephrotoxicity, characterized by morphological destruction of intracellular organelles, cellular necrosis, loss of microvilli, alterations in the number and size of the lysosomes and mitochondrial vacuolization, followed by functional alterations including inhibition of protein synthesis, glutathione depletion, lipid peroxidation and mitochondrial damage<sup>(8)</sup>.

Proximal tubular damage appears acutely after administration of cisplatin as the result of impairment of cell energy production, possibly by binding to proximal tubular cellular proteins and sulfhydryl groups with disruption of cell enzyme activity and uncoupling of oxidative phosphorylation<sup>(9)</sup>. The initial proximal tubular damage is followed by a progressive loss of glomerular filtration and impaired distal tubular function<sup>(10)</sup>.

Curcumin [1,7-bis(4-hydroxy-3-methoxyphenyl)-1,6-heptadiene-3,5-dione] is the principal curcuminoid of the popular Indian spice turmeric, which is a member of the ginger family (Zingiberaceae) <sup>(11)</sup>.



## Curcumin structure <sup>(12)</sup>

Curcumin has been shown to have multiple anticancer effects, including inhibition of proliferation<sup>(13,14)</sup>, induction of apoptosis, inhibition of angiogenesis<sup>(13,15)</sup>, and inhibition of DNA topoisomerase II <sup>(13)</sup>.

## Materials and methods

Thirty male rats (*Rattus norvegicus*) weighing (200-250 gm), were selected. The animals were housed in the animal house of Institute of embryo researches and infertility treatment / Al-Nahrain University under condition of controlled temperature, allowed free access to water and food. The rats were equally divided into three groups (10 rats/ group). Normal group (Isotonic saline group) rats were received 0.2 ml of isotonic saline intraperitoneally (I.P.) for four days and sacrificed one day after. Control group (Cisplatin- induce nephrotoxicity group) rats were received 0.2 ml of isotonic saline (I.P.) one day prior to cisplatin administration (single I.P. injection 12mg/kg for induction of nephrotoxicity) and repeated isotonic saline administration daily, then rats were sacrificed after 3 days of cisplatin administration. curcumin group rats received curcumin (50 mg/ kg I.P.) prophylactically one day prior to cisplatin administration (single I.P. injection 12mg/kg) and repeated curcumin administration daily for further three days. The levels of serum creatinine<sup>(16)</sup>, BUN <sup>(17)</sup>, serum total protein <sup>(18)</sup>, serum albumin <sup>(19)</sup>, serum glutathione <sup>(20)</sup>, serum Malondialdehyde (MDA) <sup>(21)</sup> were measured and histopathological study by staining with hematoxylin and eosin <sup>(22)</sup>, then examined under light microscope were done.

## Statistical methods:

In this study, the obtained quantitative data were presented as [mean  $\pm$  standard error of mean (S.E.M.)]. Student (unpaired) t-test for independent data was used to test the significance of the differences between the results of the any two groups <sup>(23)</sup>.

## Results

Cisplatin administration resulted in a significant increase in serum creatinine ( $P = 1.923 \times 10^{-07}$ ) (fig. 1), serum BUN ( $P = 3.9 \times 10^{-09}$ ) (fig. 2) and serum MDA ( $P = 0.000339$ ) (fig. 4) while serum glutathione level was significantly decrease ( $p = 5.314 \times 10^{-11}$ ) (fig. 5) compared to that of normal group. There were no significant changes in serum total protein, albumin and globulin when

compared with normal group (fig. 3). Section of the kidneys rats treated by Cisplatin showing degenerative and necrosis of the proximal and distal convoluted tubules (fig. 7) when compared with normal group (fig. 6). Curcumin administration resulted in a significant decrease in serum creatinine ( $p = 9.97 \times 10^{-07}$ ), serum BUN ( $p = 4.1 \times 10^{-08}$ ) and serum MDA ( $p = 0.00025$ ) while serum glutathione level was to be significantly increased ( $p = 2.61 \times 10^{-6}$ ). There were very minimal degenerative changes in kidney tissues of rats comparing to that of control group (fig. 8)

## Discussion:

The usefulness of cisplatin is limited by its toxicity to normal tissues, including cells of the kidney proximal tubules. It has been reported that cisplatin induced nephrotoxicity is closely associated with an increase in lipid peroxidation in the kidney. In addition, cisplatin has been found to lower the activities of antioxidant enzymes and to induce depletion of GSH<sup>(8)</sup>. Since the induction of nephrotoxicity by cisplatin is assumed to be a rapid process and renal damage occurs within 1 hour so it is important that the protective agent is present in renal tissue before damage occurs<sup>(24)</sup> thus, in this study, curcumin was administered one day before cisplatin injection. The most common manifestation of cisplatin induced nephrotoxicity is a decline in glomerular filtration rate (GFR) leading to a rise in serum creatinine and BUN. The onset of toxicity in hospitalized, acutely ill patients is most often recognized by routine laboratory monitoring of these two chemistries<sup>(25; 26)</sup>. Serum creatinine is considered as a marker of acute nephrotoxicity<sup>(6)</sup>. In the present study, using an experimental model of cisplatin-induced nephrotoxicity in rats (single dose of 12mg/kg I.P.) was characterized by alterations in renal function as a significant increase in serum creatinine and BUN levels compared to normal group and this result is compatible with those observed by many others<sup>(8,26, 27)</sup>.

In all groups of this study; there were no significant changes in serum total protein, serum albumin and serum globulin comparing to that of normal group. Yao and co-workers (2007)<sup>(28)</sup> mentioned that in cisplatin induced nephrotoxicity there is a little change in protein being excreted in urine. In the present study, cisplatin caused elevation of MDA, reduction of glutathione which indicated severe kidney damage. Histological results of this study indicated that severe degenerations at the proximal tubular cells could be correlated with the harmful effects of cisplatin parallel to high MDA and low GSH levels. The increase in thickness of the glomeruli basement membrane could be a result of membrane disturbance due to cisplatin administration. Lipid peroxidation mediated by oxygen free radicals causes destruction and damage to cell membranes<sup>(29)</sup>.

Curcumin (50 mg/kg I.P.) administration resulted in highly significant decrease in serum creatinine and BUN levels comparing to that of control group.

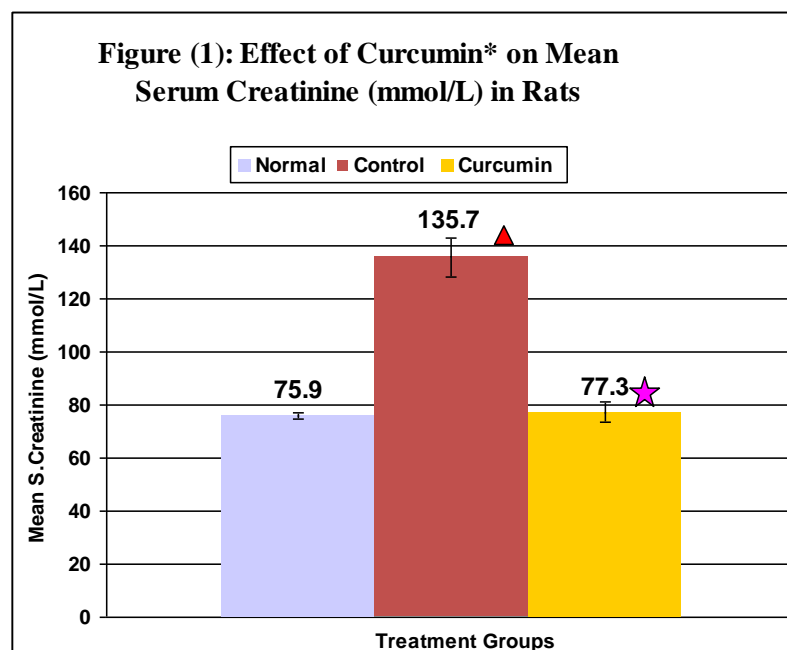
There was high significant decrease in serum MDA and high significant increase in serum glutathione comparing to that of control group.

Curcumin has strong antioxidant and anti-inflammatory properties <sup>(30, 31)</sup>. Extensive scientific researches on curcumin demonstrated its antiinflammatory action. Curcumin was found to inhibit arachidonic acid metabolism, cyclooxygenase, lipoxygenase, cytokines (Interleukins and tumor necrosis factor) and release of steroidal hormones. Curcumin was reported to stabilize lysosomal membrane and cause uncoupling of oxidative phosphorylation besides having strong oxygen radical scavenging activity, which was responsible for its anti-inflammatory property. Also curcumin was found to be a potent scavenger of superoxide <sup>(32)</sup>.

The role of curcumin is supported by a number of scientific evidences that have confirmed its anti-inflammatory and antioxidant actions both in vivo and in vitro. Many activities of curcumin can be also explained by its ability to suppress acute and chronic inflammation by scavenging reactive oxygen and reactive nitrogen species and enhancing antioxidant defense (i.e. by increasing glutathione level). However, curcumin is not only a simple antioxidant, but it plays a key role in activating antioxidative enzymes <sup>(33)</sup>. It has been proposed that antioxidants which maintain the concentration of reduced glutathione may restore the cellular defence mechanisms, block lipid peroxidation and thus protect against the toxicity of a wide variety of nephrotoxic chemicals <sup>(34)</sup>. Priyadarsini et al., (2003) <sup>(12)</sup> tested the antioxidant activity of curcumin by radiation-induced lipid peroxidation in rat liver microsomes and found that the efficiency to inhibit lipid peroxidation is 82%. P53 is a key regulatory protein in the cell cycle and initiator of apoptosis <sup>(35)</sup>. Jiang et al. (2007) <sup>(36)</sup> suggested a role of p53 in renal cell injury by cisplatin and it is activated during cisplatin treatment. Curcumin has an unprecedented number of molecular targets justifying its chemopreventive, antioxidant and anti-inflammatory activities; one of these targets is p53 <sup>(33)</sup>.

Histological result of the rat treated with curcumin showed very mild degenerative changes of the epithelial cells of the proximal tubules. The glomeruli look like normal, this correlated to strong antioxidant and anti-inflammatory properties of curcumin. One of cisplatin- induced nephrotoxicity mechanisms is elevated renal expression of TNF- $\alpha$  <sup>(37)</sup>, and as mentioned above curcumin has the ability to inhibit TNF- $\alpha$  <sup>(32)</sup> and this may be one of the protective mechanism of curcumin against cisplatin nephrotoxicity.

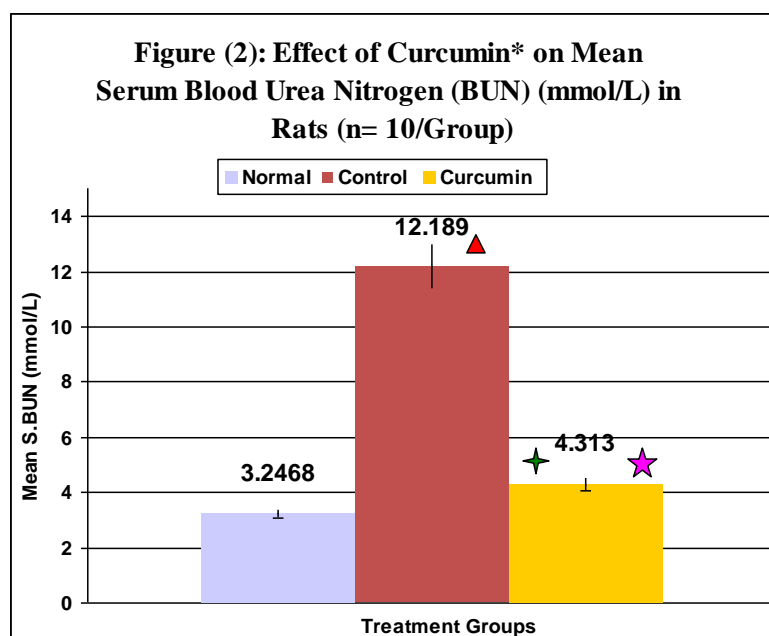
**Conclusion:** curcumin (50 mg/ kg, I.P.) had protective effects against cisplatin induced nephrotoxicity in rats.



\* = Curcumin (50mg/kg) I.P.

▲ = Significant difference ( $P = 1.923 \times 10^{-07}$ ) when compared with corresponding value in normal group.

★ = Significant difference ( $p = 9.97 \times 10^{-07}$ ) when compared to corresponding value in control group.



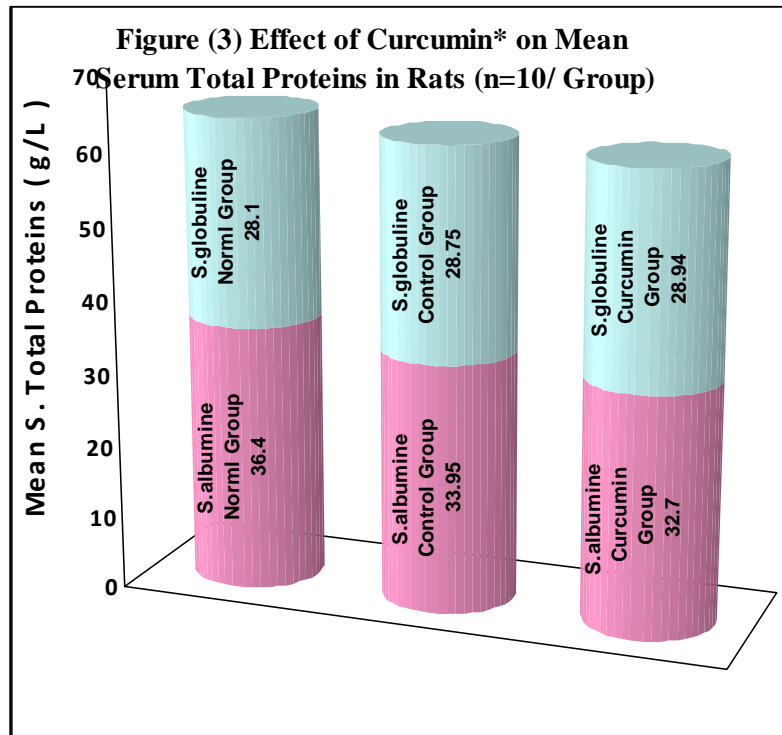
\* = Curcumin (50mg/kg) I.P.

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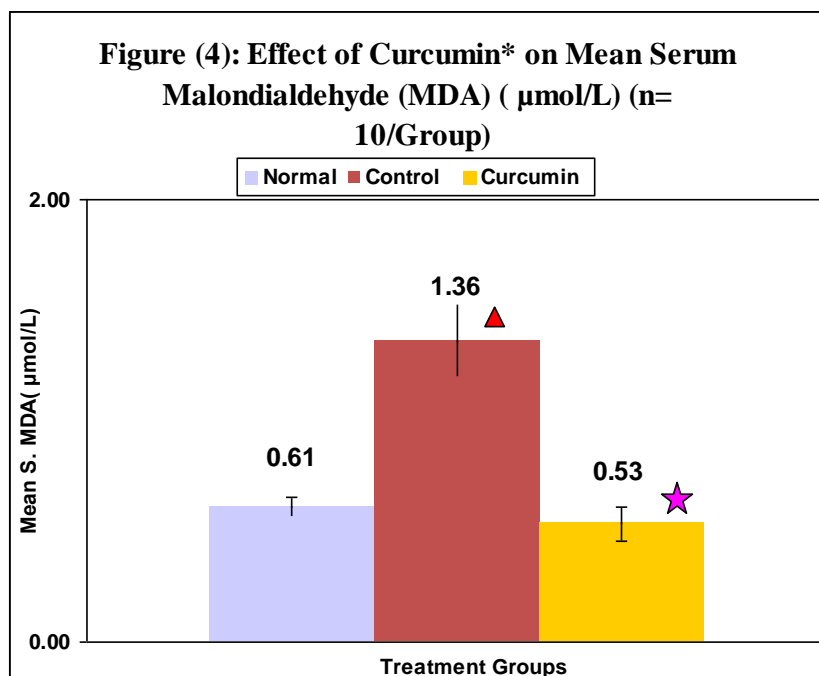
★ = Significant difference ( $p = 4.1 \times 10^{-08}$ ) when compared to corresponding value in control group.

✦ = Significant difference ( $p = 0.0029$ ) when compared to corresponding value in normal group.





\* = Curcumin (50mg/kg) I.P.

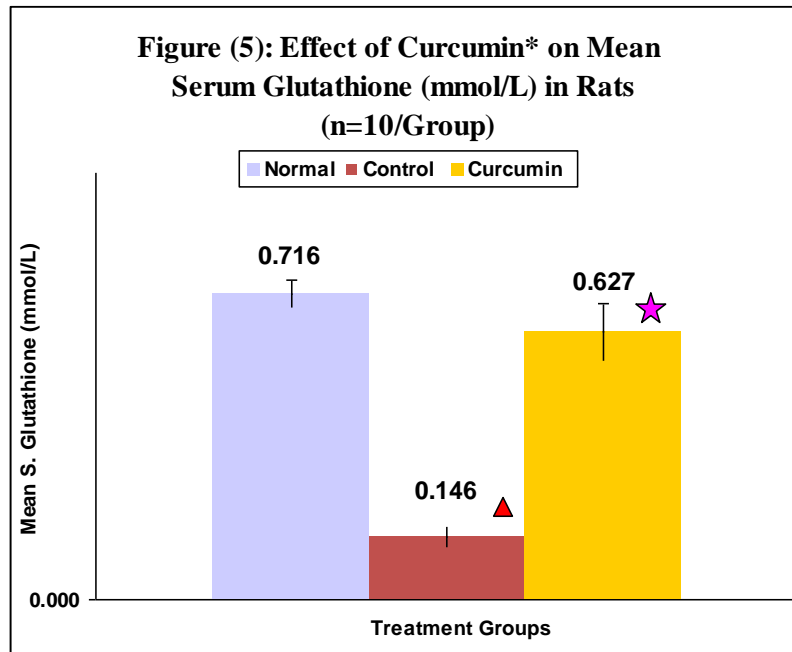


\* = Curcumin (50mg/kg) I.P.

▲ = Significant difference (P =0.000339) when compared to corresponding value in normal group.

★ = Significant difference (p =0.00025) when compared to corresponding value in control group.



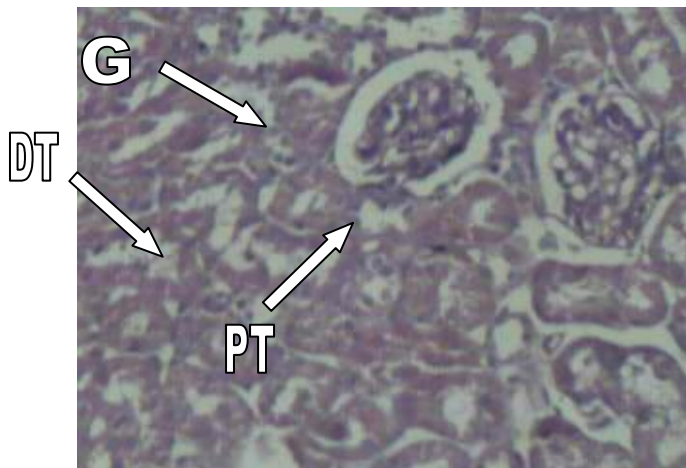


\* = Curcumin (50mg/kg) I.P.



=Significant difference ( $p = 5.314 \times 10^{-11}$ ) when compared to corresponding value in normal group

=Significant difference ( $p = 2.61 \times 10^{-6}$ ) when compared to corresponding value in control group.



**Figure (6): Normal appearance of glomeruli, proximal and distal convoluted tubules of rat's kidney.**

( Magnification: 200 X, staining; haematoxyline and eosin)

G: glomerulus , PT : proximal tubule , DT : distal tubule



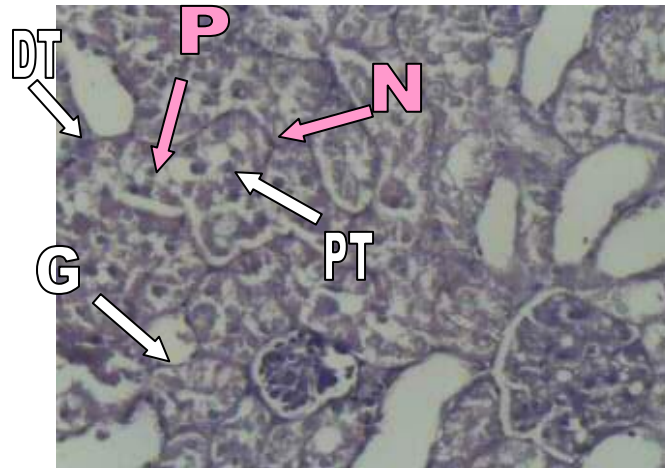


Figure (7): Nephrotoxic effect of cisplatin on rat's kidney  
Magnification: 200 X, staining; haematoxylline and eosin.

G: glomerulus , PT : proximal tubule , DT : distal tubule  
N: Necrosis, P: Pyknosis

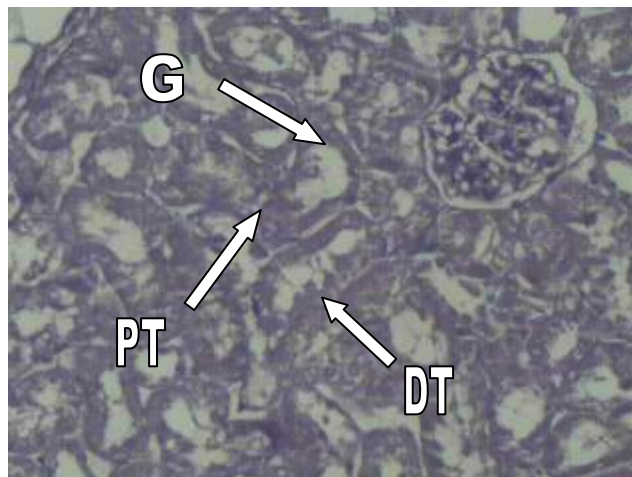


Figure (8): Protective Effect of Curcumin on Cisplatin-Induced Nephrotoxicity in Rats

Magnification: 200 X, staining; haematoxylline and eosin

(Very mild degenerative changes of the epithelial cells of the proximal tubules with accumulation of glycoprotein inside the renal tubules. The glomeruli look like normal)

G: glomerulus , PT : proximal tubule , DT : distal tubule

## References

1. Todd RC, Lippard SJ. Inhibition of transcription by platinum antitumor compounds. *Metallomics* 2009; 1(4): 280–291.
2. Boulikas T. Molecular mechanisms of cisplatin and its liposomally encapsulated form, Lipoplatin™. Lipoplatin™ as a chemotherapy and antiangiogenesis drug. *Cancer Therapy* 2007; 5: 351-376.
3. Howland RD, Mycek MJ. Anticancer Drugs: In Lippincott's Illustrated Reviews: Pharmacology, 3<sup>rd</sup> ed. A Wolters Kluwer company, Philadelphia 2006:479-480.
4. Hanigan MH, Devarajan P. Cisplatin nephrotoxicity: molecular mechanisms. *Cancer Therapy* 2003; 1: 47–61.
5. Brunton LL, Parker KL, Blumenthal DK, Buxton IL. Goodman & Gilman's Manual of Pharmacology and Therapeutics. 11<sup>th</sup> ed. McGraw-Hill. New York 2008. 189; 858-868.
6. Filipski KK, Mathijssen RH, Mikkelsen TS, Schinkel AH, Sparreboom A. Contribution of organic cation transporter 2 (OCT2) to cisplatin-induced nephrotoxicity. *Clin Pharmacol Ther*; 2009; 86(4): 396–402.
7. Kim YH, Choi BK, Kim KH, Kang SW, Kwon BS. Combination therapy with cisplatin and anti-4-1BB synergistic anti-cancer effects and amelioration of cisplatin-induced nephrotoxicity. *Cancer Research*. 2008; 68(18): 7264– 7269.
8. Joy J. Nair CK. Amelioration of cisplatin induced nephrotoxicity in Swiss albino mice by *Rubia cordifolia* extract. *Journal of Cancer Research and Therapeutics*; 2008; 4(3):111-115.
9. Taguchi T, Nazneen A, Abid MR and Razzaque MS. Cisplatin- associated-nephrotoxicity and pathological events. *Contrib Nephrol* ; 2005; 148: 107-121.
10. Kintzel PE. Anticancer drug-induced kidney disorders. *Drug Safety: an international journal of medical toxicology and drug experience*; 2001; 24(1): 19–38.
11. Kolev TM. , Velcheva EA., Stamboliyska BA. , Spiteller M. DFT and experimental studies of the structure and vibrational spectra of curcumin. *International Journal of Quantum Chemistry* ; 2005; 102(6): 1069 – 1079.
12. Priyadarsini KI., Maity DK., Naik GH., Kumar MS., Unnikrishnan M.K., Satav JG. and Mohan H.. Role of phenolic O-H and methylene hydrogen on the free radical reactions and antioxidant activity of curcumin. *Free Radical Biology and Medicine*; 2003; 35(5): 475–484.
13. Hartojo W., Silvers AL., Thomas DG. , Seder CW., Lin L., Rao H.,Wang Z.,

- Greenson JK. , Giordano TJ. and Orringer MB. Curcumin promotes apoptosis, increases chemosensitivity, and inhibits nuclear factor  $\kappa$ B in esophageal adenocarcinoma · Transl Oncol ; 2010; 3(2) : 99–108.
14. Yallapu MM., Maher DM., Sundran V., Bell MC., Jaggi M and Chauhan SC. Curcumin induces chemo/radio-sensitization in ovarian cancer cells and curcumin nanoparticles inhibit ovarian cancer cell growth. J Ovarian Res. 2010; 3 (11).
  15. Wilken R., Veena MS., Wang MB., and Srivatsan ES. Curcumin: A review of anti-cancer properties and therapeutic activity in head and neck squamous cell carcinoma. Mol Cancer. 2011; 10 (12).
  16. Henry RJ. Clinical chemistry, principles and tecnics ,2<sup>nd</sup> edition, Harper and Raw 1974: 525.
  17. Fawcett JK, Scott JE. Determination of urea in blood or serum. J.Clin. Path. 1960; 13: 156-159.
  18. koller A. Total serum protein. Kaplan A. *et al* . clinical chemistry. The C.V. Mosby Co. St Louis. Toronto. Princeton 1984 : 418; 1316- 1324.
  19. Gendler S, Kaplan A. *et al* .clinical chemistry. The C.V. Mosby Co. St Louis. Toronto. Princeton. 1984: 425; 1268-1273.
  20. Godin DV., Wahaieb SA. and Garent ME. Antioxidant enzyme alteration in experimental and clinical diabetes . Mol Cell Biochem 1988 ; 84 : 223-231.
  21. Karatas F., Kara H., Servi S., Tug T., Erulas FA. and Koca M. Investigation of antioxidant vitamins (A, E, C) and lipid peroxidation levels in rats injected N-(1, 3-Benzothiazol-2-yl)- N-(4, 5-dihydro-1H-imidazol-2-yl) amine. Molecules 2005: 10: 922- 928.
  22. Bauer JD, Ackermann PG, Toro G. Clinical lab methods. The C.V. mosby company Saint Louis 1978; 813-817.
  23. Daneil WW. Biostatistics: A foundation for analysis in the health sciences. 3rd ed. John Wiley and Sons. New York health 1983; 89- 92; 102-103.
  24. Rao M, Rao MN. Protective effects of selenomethionine against cisplatin-induced renal toxicity in mice and rats. J.Pharm.Pharmacol. 1998 ; 50:687-691.
  25. Nolin TD, Himmelfarb J. Drug –induced kidney disease. In: DiPiro JT., Talbert RL. , Yee GC. , Matzke GR., Wells BG., Posey LM. Pharmacotherapy a pathophysiologic approach 7<sup>th</sup> ed. McGraw-Hill Companies. New York. 2008. 796-799.
  26. Gamal el-Din AM, Al-Bekairi AM. Carvedilol, a beta adrenoceptor blocker with antioxidative potential, attenuates Cisplatin-induced Nephrotoxicity in Rats. Journal of Applied Scienes Research 2006; 2(6): 331-335.
  27. Kim M., Yang HN. Kim H., Jo S., Cho WY. and Kim HK. IL-10

- mediates rosiglitazone-induced kidney protection in cisplatin nephrotoxicity. *Journal of the Korean Academy of Medical Sciences*; 2010; 25(4): 557–563.
28. Yao X, Panichpisal K, Kurtzman N, Nugent K. Cisplatin nephrotoxicity . *Am J Med Sci* 2007; 334(2): 115-124.
29. Arda-Pirincci P, Bilgin-Sokmen B, Yanardag R, Bolkent S. Effects of zinc on intestinal injury and some serum Parameters in ethanol-administered rats. *Biosci. Biotechnol. Biochem*; 2009; 73(2): 260-267.
30. Menon VP. and Sudheer AR. Antioxidant and anti- inflammatory properties of curcumin . *Advances in Experimental medicine and Biology*; 2007; 595: 105-125.
31. Shukla PK., Khanna VK. , Ali MM., Khan MY., Srimal RC. Anti-ischemic effect of curcumin in rat brain . *Neurochemical Research*; 2008; 33(6): 1036-1043.
32. Kohli K. , Ali J., Ansari M. J., Raheman Z. Curcumin: A natural antiinflammatory agent. *Indian J. Pharmacol.*; 2005; 37(3): 141-147.
33. Sikora E., Scapagnini G and Barbagallo M. Curcumin, inflammation, ageing and age-related diseases. *Immunity and Ageing*; 2010; 7: (1).
34. Babu E., Gopalakrishnan V. K., Sriganth INP., Gopalakrishnan R. and Sakthisekaran D. Cisplatin induced nephrotoxicity and the modulating effect of glutathione ester. *Molecular and Cellular Biochemistry*; 1995; 144(1): 7-11.
35. Brunton LL., Parker KL., Blumenthal DK., and Buxton IL. Goodman & Gilman's Manual of Pharmacology and Therapeutics. 11<sup>th</sup> ed. McGraw-Hill. New York. 2008: 189; 858- 868.
36. Jiang M., Wei Q., Pabla N., Dong G., Wang C., Yang T., Smith SB. And Dong Z. Effects of hydroxyl radical scavenging on cisplatin -induced p53 activation, tubular cell apoptosis and nephrotoxicity. *Biochemical Pharmacology*; 2007; 73(9):1499-1510.
37. Ali BH., Al-Moundhri M., Eldin MT., Nemmar A., Al- Siyabi S. and Annamalai K. Amelioration of cisplatin-induced nephrotoxicity in rats by tetramethylpyrazin , a major Constituent of the Chinese Herb *Ligusticum wallichii*. *Experimental biology and medicine*; 2008; 233 (7): 891-896.