

Genetic study of relationship between resistance of *Enterobacter aerogenes* to some antibacterial agents and plasmid containing

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لبعض المضادات البكتيرية *Enterobacter aerogenes* دراسة وراثية للعلاقة بين مقاومة بكتريا ومحتواها البلازميدي

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الخلاصة :

هدفت الدراسة الى تحديد العلاقة بين مقاومة بكتريا *Enterobacter aerogenes* الوراثة لبعض المضادات الحيوية ومحتواها البلازميدي. اذ تم جمع (70) عينة ادرار من اشخاص مصابين بالتهاب المجاري البولية حسب التشخيص السريري وتم الحصول على (15) عزلة تعود لبكتريا *Enterobacter aerogenes*. تم اجراء فحص المقاومة لمضادات البكتيريا بأستخدام (11) مضاداً بكتيرياً وكانت نسبة المقاومة لمضاد النايتروفورانتين (100%) ، الترياميثوبريم (90%) ، اميكاسين (35%) ، نورفلوكساسين (45%) ، سيفوتاكزيم (50%) ، سفترياكزون (38%) ، كلورامفينيكول (64%) ، دوكسوسايكلين (80%) ، جنتاميسين (58%) ، سبروفلوكساسين (53%) ، اميبينيم (10%) . اشارت البيانات الى ازدياد نسبة اصابة المجاري البولية بهذه البكتريا . اظهرت البكتريا مقاومة كاملة لمضاد النايتروفورانتين ومقاومة شبه كاملة لمضاد الترياميثوبريم مما يؤدي الى عدم استخدامها مستقبلاً . ارتبط مقاومة هذه البكتريا المختلفة لمضادات الحياة وراثياً بزيادة الحزم البلازميدية ، حيث كلما كان عدد الحزم البلازميدية أكثر كانت البكتريا أكثر مقاومة لمضادات الحياة المستخدمة في الدراسة .

Abstract

The present study aimed to determine the relationship between the genetic resistance of *Enterobacter aerogenes* to some antibacterial agents, and its plasmid containing. For this purpose (70) samples of urine were collected from patients with clinically diagnosed urinary tract infection, there were (15) isolate *Enterobacter aerogenes*. The Antibiotic susceptibility test was used through using (11) antibiotics and resistance bacteria *Enterobacter aerogenes* was nitrofurantion (100%) , trimethoprim (90 %) , amikacin (35 %) , norfloxacin (45%) , cefotaxime (50 %) , ceftriaxon (38 %) , chloramphenecol (64 %) , imipenem (10 %) , doxocyclin (80 %) , gentamycin (58 %) and ciprofloxacin (53 %) . The data indicated the increase infection of urinary tract infection that caused by *Enterobacter aerogenes* . There are two antibiotics nitrofurantion and trimethoprim must not use to treatment against *Enterobacter aerogenes* in the future because the bacteria gave high resistance these antibiotics. The resistance of *Enterobacter aerogenes* to antibiotics used in this study linked with increasing bounds of plasmid.

1. Introduction

Urinary tract infection is among the most common nosocomial and community acquired infections. Information on prevailing levels of antimicrobial Resistance among common pathogens associated with urinary tract infection is useful in making an appropriate choice of empiric therapy^[1]. Resistance to antibiotic treatment in patients with urinary tract infection is a representative example of the increasing problem of antimicrobial resistance^[2]. The bacteria *Enterobacter aerogenes* is an opportunistic pathogen and its one type very important cause urinary tract infection and nosocomial infection in addition to wound and blood stream infections because it has multidrug resistance to antibiotics^[3,4,5,6,7]. *Enterobacter aerogenes* is an important pathogen in hospital acquired infection. It generally exhibits resistance to a variety of broad-spectrum antimicrobial agents, including beta-lactamase^[8]. The existence of a prevalent resistant clone of *Enterobacter aerogenes* has been reported^[9]. *Enterobacter aerogenes*; a nosocomial pathogen, is frequently exhibiting multidrug resistance mechanisms associated with a change in membrane permeability^[10].

2-Materials and methods

2 – 1 : Specimens

The study includes collecting (70) urine samples from some hospitals in Baghdad Governorate. Then the isolates were cultured on MacConkeys and blood agar plates as well as used analytic profile index twenty enterobacteriaceae (API20E) to identify the bacterial species.

2 – 2 : Antibacterial susceptibility testing

Antibiograms were tested according to (Bauer et al.(1966)^[11]) as follows:

1.Preparation of bacterial supernatant by use of normal saline and compare turbidity of bacterial supernatant with standard turbidity (MacFarland) that refers to about (1.5×10^8) cell/ml.

2.The cotton swabs were used to spread part of bacterial supernatant on the plates Muller-Hinton agar. The antibacterial discs were put on the isolate cultured in Muller-Hinton agar (five discs in one plate) by using sterile forces.

3.The plates Muller-Hinton agar were incubated in the incubator at (37°C) for (24) hours. The inhibition zones were measured. The results were expressed as susceptible or resistant according to the (NCCLS,2007)^[12].

2 – 3 : DNA plasmid extraction

Used boiling method to obtain DNA plasmid according to (Holmes and Ougly.1981)^[13] as follows:

1.Transport (1.5 ml) from bacterial supernatant to a microfuge tube (all one isolate alone) and separated by using microcentrifuge in speed (5000 rpm/minute) for (5) minutes.

2.Add (350µl) from solution Sucrose Tris-HCl EDTA (STET) and (25µl) from solution lysozyme (10mg/ml) to the deposit and mixed solution by using vortex for (3 seconds).

3.The solution was put in a water bath (100°C) for (40 seconds) and separated solution by using microcentrifuge in speed (13000 rpm/minute) for (10) minutes.

4.Remove the viscous pellet and add (40µl) from solution potassium acetate and (420µl) from isopropyl alcohol, this material mixed and saved in (-20°C) for (1-2) hours.

5. separate solution by using microcentrifuge in speed (13000 rpm / minute) for (15) minute and add (50µl) Tris-Hcl EDTA and became ready for the electrophoresis.

2 – 4 : DNA plasmid electrophoresis in gel agarose

The gel electrophoresis used in detection of plasmid DNA according to (Maniatis *et al* 1982) ^[14] as the following:

1. Preparation gel agarose in concentration (0.7%) by using Tris-Hcl boric acid EDTA (TBE). The gel agarose heated to the boiling degree and cooling in (45-50 °C) and add (10µl) ethidium bromide in concentration (0.5µg/ml).

2. The comb fixation in the slab to create wells that contain the sample and add gel agarose carefully and abandon for (30 minute) to solidify.

3. Remove comb from gel agarose carefully and fixate the slab in electrophoresis instrument and add Tris-Hcl boric acid EDTA to cover surface of the gel agarose.

4. Put (10µl) from the sample that will be tested in eppendorf tube and add (5µl) loading buffer and mixed carefully.

5. The samples were put in wells and pass the electricity (5 volt / cm²) for (1-2) hour until the pigment arrive to other side to the gel agarose.

6. The agarose test by using ultraviolet illuminator in wave length (360) nanometer.

3- Results and Discussion

We received and examined (70) urine specimens during the study period cultured the specimens on the blood and MacConkey agar plates. Then after incubation period late lactose fermented will be taken, after that identification the isolates by analytic profile index enterobacteriaceae (API20E). The results were (15) isolates of *Enterobacter aerogenes*. Then antibiotics susceptibility test was used by using (11) antibiotics disks. The results clarified that the resistance nitrofurantoin (100%), trimethoprim (90%), amikacin (35%), norfloxacin (45%), cefotaxime (50%), ceftriaxone (38%), chloramphenicol (64%), imipenem (10%), doxycycline (80%), gentamycin (58%) and ciprofloxacin (53%), after that choice (4) isolates resistance to antibiotics in differential levels E₅, E₁, E₁₁ and E₉ who appeared 10, 8, 6 and 4 respectively of antibiotic resistance. Extraction and gel electrophoresis of plasmid DNA were used for these four isolates and the results show that these isolates contained number of DNA plasmid bands differed from isolate to another (shape 1). The isolate became more resistant when it contained more plasmid bands.

Alaa Salim Hamzah

Table (1) Resistance the *Enterobacter aerogenes* to the Antibiotics

Bacteria	Antibiotics	%
<i>Enterobacter aerogenes</i>	nitrofurantoin	(100%)
	trimethoprim	(90 %)
	amikacin	(35 %)
	norfloxacin	(45 %)
	cefotaxime	(50 %)
	ceftriaxone	(38 %)

	chloramphenicol	(64 %)
	doxocyclin	(80 %)
	gentamycin	(58 %)
	ciprofloxacin	(53 %)
	imipenem	(10 %)

containinig to the
Plasmid :Shape (1)



isolates (E₁ , E₅ , E₉ , E₁₁)

Urinary tract infections constitute one type of problems of the most frequently encountered conditions in clinical medical practice requiring antimicrobial therapeutic intervention . The research was done to known relationship between the presence of extended spectrum beta lactamase encoded plasmid (ESBL) and the drug resistance of *Enterobacter aerogenes* . This bacteria can show resistance to Gentamycin , Amikacin and Ciprofloxacin as well as a resistance to betalactam drug ^[15] . *Enterobacter aerogenes* have other mechanisms to resistance antibiotics like modification of outer membrane and change metabolic pathway may be resistant chloramphenicol , Trimethoprim and betalactam antibiotics ^[16,17] . Other mechanism efflux pump may appear multidrug resistance to antibiotics ^[18,19] . The isolates *Enterobacter*

aerogenes in this study appear differential resistance to antibiotics used in this study and increase resistance with increasing number of bounds plasmid to this bacteria .

5- Conclusion

This study indicated the spread infection of urinary tract infection that was caused by *Enterobacter aerogenes* . This bacteria has 4 differential levels to resistance common antibiotics that used in treatment . more resistance was nitrofurantion (100%) and trimethoprim (90%) , these antibiotics can not be used in treatment of UTI caused by *Enterobacter aerogenes* in the future . The increase resistance *Enterobacter aerogenes* to antibiotics in this study linked with increasing plasmid bounds .

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