

***Leuconostoc mesenteroides* cause Nosocomial UTI  
At a tertiary care center in North India**

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**Abstract**

Eight strains of gram-positive cocci highly resistant to vancomycin (MICs of 512 - 1,024 µg/ml) were isolated from urine sample. These organisms were identified as *Leuconostoc mesenteroides*. They were tested to determine susceptibility to 12 antimicrobial agents by agar dilution method. Out of 8 isolates of *L. mesenteroides*, all of these isolates (100%) were resistant to vancomycin and 6 (75%) were resistant to teicoplanin. Seven (87.5%) isolates were sensitive to clindamycin, 6 (75.0%) isolates were sensitive to chloramphenicol, erythromycin, gentamicin and streptomycin, 5 (62.5%) isolates were sensitive to ampicillin, penicillin G and tetracycline, while 4 (50%) isolates were sensitive to kanamycin and Trimethoprim. We are reporting nosocomial UTI caused by these organisms. The results provide evidence for the possibility of nosocomial transmission of this unusual opportunistic, vancomycin-resistant pathogen.

## Introduction

*Leuconostoc* species are gram-positive catalase-negative coccoid or coccobacillary (Facklam and Elliott, 1995). In 1985, Buu-Hoi *et al.* ([1985](#)) reported the first cases of *Leuconostoc* infection in humans. Since then, *Leuconostoc* spp. have been implicated in a variety of infections (Handwerger *et al.*, 1990; Ferrer *et al.*, 1995; Jimenez-Mejias *et al.*, 1997; Cappelli *et al.*, 1999; Albanese *et al.*, 2006), particularly in patients being treated with vancomycin and in immunocompromised patients (Albanese *et al.*, 2006). *Leuconostoc* species are often misidentified as lactobacilli, streptococci, pediococci or enterococci as all these genera share several biochemical properties ([MacGowan et al.](#), 1989; [Winston et al.](#), 2004).

The *Leuconostoc* are naturally highly resistant to vancomycin with MIC>256 µg/ml but could be successfully treated with penicillin with MIC ranging from 0.25 -1.0 unit/ml (Buu-Hoi *et al.*, 1985; Kulwichit *et al.*, 2007).

In the last decade *Leuconostoc* species have been reported with increasing frequency as human pathogens, causing bacteremias (Barreau C, Wagener, 1990), meningitis (Friedland *et al.*, 1990), breast abscess (Barry *et al.*, 1993), abdominal abscess (Montejo *et al.*, 2000), peritonitis (Templin *et al.*, 2001; Helali *et al.*, 2005). Occasionally it has been isolated in cases of catheter associated infections, sepsis, pneumonia, osteomyelitis and hepatic dysfunction (Jofré *et al.*, 2006). Recently, these organisms have also been implicated in causing a small outbreak of nosocomial urinary tract infection UTI. The aims of the present study were to characterize the epidemiologic features of the infection with *L. mesenteroides*.

## **Methods**

### **Sample collection and identification**

A total of 558 urine samples (midstream and catheter) were recovered for a period of 5 months from January to June 2008. The samples were submitted for culture to the Department of Medical Microbiology at Postgraduate Institute of Medical Education & Research (PGIMER), Chandigarh, India, from admitted patients for suspected UTI, and cultured on cysteine lactose electrolyte deficient medium (CLED) (Difco Laboratories, Detroit, USA) during the period. Suspected *Leuconostoc* colonies were streaked onto brain heart infusion (BHI) agar (Difco Laboratories, Detroit, USA) supplemented with 6 µg/ml vancomycin and incubated overnight at 37°C. Only the initial isolate from each patient was tested.

### **Biochemical characteristics**

Tentative identification of bacterial colonies belonging to *L. mesenteroides* was performed by culture characteristics, Gram stain, catalase test, ability to hydrolysis of esculin using bile esculin azide (BEA) broth (Difco Laboratories, Detroit, USA), ability to utilize different carbon sources and production of gas from glucose, ability to grow in BHI agar containing 6.5% NaCl and growth in BHI broth at incubation temperature of 10°C as per conventional identification scheme (Facklam *et al.*, 1989). Vancomycin susceptibility was tested by E-test (AB Biodisk, Solna, Sweden). Definitive identification up to species level was made with the BD-BBL Crystal Identification System, (Gram-Positive ID Kit-Sparks, Maryland, USA).

## Antimicrobial susceptibility testing

*L. mesenteroides* strains were tested for resistance to ampicillin, penicillin G, erythromycin, chloramphenicol, clindamycin, gentamicin, kanamycin, streptomycin, tetracycline, trimethoprim, vancomycin and teicoplanin according to the current guidelines of (NCCLS, 2000). Bacterial suspensions were adjusted to a 0.5 McFarland turbidity standard. A 10.0 µl aliquot of each suspension was spotted onto the agar surface to achieve a final inoculum of approximately  $10^5$  CFU per spot. It was allowed to absorb into the agar for 10 min. The results were read after incubation at 37°C for 18-24 h and 48 h. Plates were read against a dark, nonreflecting background. Strains were considered antibiotic resistant if growth was positive or weak or if more than one colony was observed.

## Results

### Biochemical identification.

Eight *L. mesenteroides* isolates were recovered from 8 patients (aged 2 months to 65 years) were gram-positive cocci occurring in pairs or short chains, all of them are highly resistant to vancomycin (MICs 512 - 1,024 µg/ml). The biochemical analysis by the BD-BBL Crystal Identification System are shown in Table 1.

**Table 1. Characteristics for identification of *L. mesenteroides* by BD BBL CRYSTAL™ system.**

Test	Substrate	<i>L. mesenteroides</i>
Morphology	Cocci	+
	Coccobacilli	+
	Rod	+
Arrangement	Chains	+
	Pairs	+
	Clusters	-
Gas from	Trehalose	+
	Lactose	V
	Galactose	-
	Mannitol	-
	Maltose	+
	Arabinose	+
	Ralbose	-
	Raffinose	+
	Esculin	+
	Arginine	-
Growth	PYRase	-
	at 10°C	+
	at 45°C	-
	in 6.5% NaCl	+
	Methyl red	-
	Voges-Proskauer	V
Resistant to	vancomycin	512 - 1,024 µg/ml

#### Antimicrobial susceptibility testing

The susceptibility testing results of 8 *L. mesenteroides* strains against 12 antimicrobial agents are shown in (Table 2).

Table 2. Susceptibility of 8 strains of *Leuconostoc mesenteroides* to 12 antimicrobial agents.

Antibiotics	No of resistance / 8	(100%)
Ampicillin	3	(37.5)
Chloramphenicol	2	(25.0)
Clindamycin	1	(12.5)
Erythromycin	2	(25.0)
Gentamicin	2	(25.0)
Kanamycin	4	(50.0)
Penicillin G	3	(37.5)
Streptomycin	2	(25.0)
Teicoplanin	6	(75.0)
Tetracycline	3	(37.5)
Trimethoprim	4	(50.0)
Vancomycin	8	(100)

### Clinical details

The clinical details for 8 patients (5 females and 3 males) are shown in (Table 3). The age ranged from 2 months to 65 yr. All females were admitted in the obstetrics and gynaecological unit. Three of them suffered from catheter related nosocomial UTI and responded to removal of catheter plus antibiotics. In two females there were no symptoms related to UTI and therefore the organisms were considered as contaminants. Among the three male patients who had nosocomial UTI, one had malignancy of urinary bladder and another had stricture urethra. The third was a two-month old child with exstrophy bladder. All patients responded to antibiotics. None of the patients developed bacteremia.

**Table 3. Clinical Profile of Patients With Significant Bacteriuria Due to *Leuconostoc mesenteroides***

<i>Patient s</i>	<i>Age / Gender</i>	<i>Ward</i>	<i>Clinical diagnosis</i>	<i>Catheter/Urinary Instrumentation</i>	<i>Type of UTI</i>	<i>Treatment</i>
<b>1</b>	<b>25y/F</b>	<b>Obstetrics &amp; Gynecology</b>	<b>Preterm labor</b>	<b>Yes</b>	<b>Nosocomial UTI</b>	<b>Amoxycillin</b>
<b>2</b>	<b>43y/F</b>	<b>=</b>	<b>Surgery for Fibroid uterus with endometriosis</b>	<b>Yes</b>	<b>=</b>	<b>Ciprofloxacin &amp; gentamicin &amp; removal of catheter</b>
<b>3.</b>	<b>25y/F</b>	<b>=</b>	<b>Preterm labor</b>	<b>No</b>	<b>Asymptomatic</b>	<b>-</b>
<b>4</b>	<b>55y/F</b>	<b>=</b>	<b>Surgery for Ovarian tumor</b>	<b>Yes</b>	<b>Nosocomial UTI</b>	<b>Gentamicin &amp; removal of catheter</b>
<b>5</b>	<b>27y/F</b>	<b>=</b>	<b>Preterm labor</b>	<b>No</b>	<b>Asymptomatic</b>	<b>-</b>
<b>6</b>	<b>65y/M</b>	<b>Surgery</b>	<b>Surgery for Urinary bladder</b>	<b>Yes</b>	<b>Nosocomial UTI</b>	<b>Ciprofloxacin</b>
<b>7</b>	<b>60y/M</b>	<b>Urology</b>	<b>Stricture urethra</b>	<b>Yes</b>	<b>=</b>	<b>Augmentin &amp; amikacin</b>
<b>8</b>	<b>02m/ M</b>	<b>Pediatric surgery</b>	<b>Exstrophy bladder</b>	<b>No</b>	<b>=</b>	<b>Penicillin &amp; Gentamicin</b>

F, female; M, male; y, year; m, month

## Discussion

*L. mesenteroides* is one of several uncommon, gram-positive, intrinsically vancomycin-resistant bacteria, that can cause serious human infections (Facklam and Elliott, 1995; Ferrer *et al.*, 1995; Moellering, 1995). *Leuconostoc* species are difficult to detect with routine methods (Facklam and Elliott, 1995) and can easily be misidentified as *Lactobacillus*, alpha-hemolytic streptococci, *Pediococcus*, *Enterococcus*, or *Lactococcus* (Facklam and Elliott, 1995). It must be distinguished from other mentioned species as there share almost similar biochemical properties (Facklam *et al.*, 1989; Isenberg *et al.*, 1988; Riebel and Washington, 1990; Barreau and Wagener, 1990). A total of 558 urine samples were cultured on (CLED) medium. The *Leuconostoc* were incidentally found. These species we observed were more coccobacillary than coccal in shape. All the strains were high resistant to vancomycin (MIC>1024µg/ml). In 1984, Shlaes and his co workers suggested that gram-positive bacteria should be routinely tested for vancomycin susceptibility. Vancomycin-resistant gram-positive cocci merit close examination, since both conventional tests and the API 20 Strep System can lead to misidentification of *Lactobacillus* spp. and of strains belonging to the genus *Leuconostoc* as viridans streptococci (Thornsberry *et al.*, 1984). In our experience, *Leuconostoc* spp. can be suspected from the vancomycin resistance and the gas production. All the strains are catalase-negative, PYR negative, could grow in presence of 6.5% NaCl and at 10°C. All produced acid from sucrose, maltose, raffinose, arabinose, and trehalose. In the present study, 6 out of 8 were lactose-positive and all were Mannitol-negative. None of the strains deaminated arginine. In the BD-BBL Crystal Identification System, none of the *Leuconostoc* strains were positive for the PRYase test. The strains were identified as *L. mesenteroides* based on conventional scheme of Facklam *et al.*, (1989).



Knowledge of the susceptibility of *L. mesenteroides* should help physicians treat infections caused by these strains. Many of the antimicrobial agents that we tested did not have uniform activities against the *L. mesenteroides* tested, so it appears that proper identification will help in the formulation of optimal antimicrobial regimens. In the present study, the results of the antibacterial activities of the various antibiotics showed that all *L. mesenteroides* isolates were resistant to vancomycin, while 6/8 of *L. mesenteroides* isolates were resistant to teicoplanin; our study is similar to the study of (de la Maza *et al.*, 1989; Albanese *et al.*, 2006). The present study's results showed that, out of 8 isolates of *L. mesenteroides*, 7 were susceptible to clindamycin, 6 *L. mesenteroides* strains were susceptible to chloramphenicol, erythromycin, gentamicin and streptomycin. While 5 isolates were susceptible to ampicillin, penicillin G and tetracycline, and 4 *L. mesenteroides* isolates were susceptible to kanamycin and Trimethoprim. Our results are a slightly similar to the observation of Buu-Hoi *et al.*, (1985) who recognized that Penicillin G and ampicillin were more active. While all strains were susceptible to tetracycline and chloramphenicol. Erythromycin and clindamycin were highly active against all strains. Trimethoprim is not active. All strains were susceptible to gentamicin. While, both of streptomycin and kanamycin were less active (Buu-Hoi *et al.*, 1985). Several reports have found that those with the best activities against all the *L. mesenteroides* strains tested were clindamycin, chloramphenicol, gentamicin, erythromycin, penicillin and tetracycline (Swenson *et al.* 1990; Bou *et al.*, 2008). As *L. mesenteroides* are intrinsically resistant to vancomycin, infections occur more frequently in patients being treated for underlying diseases with vancomycin therapy (Bauer *et al.*, 1966). None of the patients in the present study had received vancomycin. Therefore, though uncommonly isolated (8/558-<0.014%) organisms can cause nosocomial UTI and have the potential for causing an outbreak. the

study of Bou *et al.*, (2008) have demonstrated that out of 42 patients infected with *L. mesenteroides*, 9 of the patients died, and 3 of the deaths (7.1%) were directly related to the *Leuconostoc* infection. In addition to these cases, a vancomycin resistant viridans group streptococcus reported by Shlaes *et al.*, 1984 may actually have been a *Leuconostoc* sp. (Isenberg *et al.*, 1988). This report is further evidence that, although rarely pathogenic, leuconostocs may cause severe infection in humans. In summary, though uncommonly isolated these organisms can cause nosocomial UTI and have potential for causing outbreak. These are

likely to be misidentified as enterococci in routine microbiology laboratory. **Conclusions**  
The *Leuconostoc* species has been considered as a potential pathogen especially in the immunocompromised host. Its clinical significance in other patients may be questionable. Further work need to be done to determine if the *Leuconostoc* species are part of the skin flora and thus give rise to contaminated urine culture results when the skin is not adequately cleaned prior to urine sampling. It would be also necessary to determine the conditions in which these isolates would be of clinical significant.

## References

- Albanese A, Spanu T, Sali M, Novezgno F, D'Inzeo T, Santagelo R, Mangiola A, Anile D. and Fadda G. (2006). Molecular identification of *Leuconostoc mesenteroides* as a cause of brain abscess in an immunocompromised patient. *J Clin Microbiol.* 44:3044-3045.
- Barreau C. and Wagener G. (1990). Characterisation of *Leuconostoc lactis* strains from human sources. *J Clin Microbiol.* 28:1728-1733.
- Barry H, Clancy MT, Brady A. and O'Higgins N. (1993). Isolation of a *Leuconostoc* species from a retroareolar breast abscess. *J Infect.* 27:208-210.
- Bauer AW, Kirby WM, Sherris JC. and Turck M. (1966). Antibiotic susceptibility testing by a standardized single disk method. *Am J Clin Pathol.* 45:493-496.

- Bou G., Saleta JL., Nieto JAS., Tomás M., Valdezate S., Sousa D., Lueiro F., Villanueva R., Pereira MJ. and Llinares P. (2008). Nosocomial Outbreaks Caused by *Leuconostoc mesenteroides* subsp. *Mesenteroides*. *Emerg Infect Dis.* 14:968-971.
- Buu-Hoi, A., C. Branger, and J. F. Acar. (1985). Vancomycin-resistant streptococci or *Leuconostoc* sp. *Antimicrob Agents Chemother.* 28:458-460.
- Cappelli EA, Barros RR, Camello TCF, Teixeira LM. and Merquior VLC. (1999). *Leuconostoc pseudomesenteroides* as a cause of nosocomial urinary tract infections. *J Clin Microbiol.* 37:4124-4126.
- de la Maza L, Ruoff KL. and Ferraro MJ. (1989). In vitro activities of daptomycin and other antimicrobial agents against vancomycin-resistant gram-positive bacteria. *Antimicrob Agents Chemother.* 33: 1383-1384.
- Facklam R. and Elliott JA. (1995). Identification, classification, and clinical relevance of catalase-negative, gram-positive cocci, excluding streptococci and enterococci. *Clin Microbiol Rev.* 8: 479-495.
- Facklam R, Hollis D. and Collins MD. (1989). Identification of gram positive coccal and cocobacillary Vancomycin-resistant bacteria. *J Clin Microbiol.* 27: 724-730.
- Ferrer S, G deMiguel, P Domingo, R Pericas. and G. Prats. (1995). Pulmonary infection due to *Leuconostoc* species in a patient with AIDS. *Clin Infect. Dis.* 21: 225-226.
- Friedland IR, Snipelisky M. and Khoosal M. (1990). Meningitis in a neonate caused by *Leuconostoc* sp. *J Clin Microbiol.* 28: 2125-2126.
- Handwerger S, Pucci MJ, Volk KJ, Liu J. and Lee MS. (1994). Vancomycin-resistant *Leuconostoc mesenteroides* and *Lactobacillus casei* synthesize cytoplasmic peptidoglycan precursors that terminate in lactate. *J Bacteriol.* 176: 260-264.
- Helali A, McAlear D. and Osoba A. (2005). *Leuconostoc* bacteremia in a child with short-gut syndrome. *Saudi Med J.* 26: 311-313.
- Isenberg HD, Vellozzi EM, Shapiro J. and Rubin LG. (1988). Clinical laboratory challenges in the recognition of *Leuconostoc* spp. *J Clin Microbiol.* 26: 479-483.
- Jimenez-Mejias ME, Becerril B, Gomez-Cia T, Del Nozal M. and Palomino-Nicas J. (1997). Bacteriemia caused by *Leuconostoc cremoris* in a patient with severe burn injuries. *Eur J Clin Microbiol Infect Dis.* 16: 533-535.

- , Sakurada ZA, Ulloa FMT, Hormázabal OJC, Godoy MV, Fernández OJ, Jofré ML Gutiérrez MM, Monteverde OMP, Castillo GM. and Canales PA. (2006). *Leuconostoc* infections in patients with short gut syndrome, parenteral nutrition and continuous enteral feeding. *Rev Chilena Infectol.* 23: 340-345.
- Kulwichit W, Nilgate S, Chatsuwan T, Krajiw S, Unhasuta C. and Chongthaleong A. (2007). Accuracies of *Leuconostoc* phenotypic identification: a comparison of API systems and conventional phenotypic assays. *BMC Infect Dis.* 7: 69.
- , Marshall RJ. and Reeves DS. (1989). Evaluation of API 20 STREP system for MacGowan AP identifying *Listeria* species. *J Clin Pathol.* 42: 548-550.
- Moellering RCJr. (1995). *Enterococcus* species, *Streptococcus bovis* and *Leuconostoc* species, p. 1826-1835. In G. L. Mandell, J. E. Bennet, and R. Dolin (ed.), *Principles and practice of infectious diseases*, 4th<sup>ed</sup>. Churchill Livingstone, New York, N.Y.
- Montejo M, Grande C, Valdivieso A, Testillano M, Minguillan J, Aguirrebengoa K. and Ortiz de Urbina J. (2000). Abdominal abscess due to *Leuconostoc* species in a liver transplant recipient. *J Infect.* 41: 197-198.
- National Committee for Clinical Laboratory Standards. (2000). Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically, (5<sup>th</sup> ed). Approved standard M7-A5. National Committee for Clinical Laboratory Standards, Wayne, Pa, USA.
- Riebel WJ. and Washington JA. (1990). Clinical and microbiologic characteristics of pediococci. *J Clin Microbiol.* 28: 1348-1355.
- Shlaes DM, Marino J. and Jacobs MR. (1984). Infection caused by vancomycin-resistant *Streptococcus sanguis* II. *Antimicrob Agents Chemother.* 25: 527-528.
- Swenson JM, Facklam RR. and Thornsberry C. (1990). Antimicrobial susceptibility of vancomycin-resistant *Leuconostoc*, *Pediococcus*, and *Lactobacillus* species. *Antimicrob Agents Chemother.* 34: 543-549.
- Templin KS, Crook T, Riley T<sup>3rd</sup>, Whitener C. and Aber RC. (2001). Spontaneous bacterial peritonitis and bacteremia due to *Leuconostoc* species in a patients with end-stage liver disease: a case report. *J Infect.* 43: 155-157.
- Thornsberry C, Baker CN. and Facklam RR. (1974). Antibiotic susceptibility of *Streptococcus bovis* and other group B streptococci causing endocarditis. *Antimicrob Agents Chemother.* 5: 228-233.
- , Pang S, Haller BL, Wong M, Chambers HF<sup>3rd</sup>. and Perdreau-Remington F. Winston LG (2004). API 20 strep identification system may incorrectly speciate enterococci with low level resistance to vancomycin. *Diagn Microbiol Infect Dis.* 48: 287-288.