

Effect of Methanolic Extract of *Myrtus Communis* on Some Pathogenic Bacteria

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ABSTRACT

In this study the antibacterial activity of the methanolic extract of *Myrtus communis* was investigated against *Escherichia coli*, *Klebsiella pneumonia*, *Pseudomonas aerogenosa* and *Staphylococcus aureus* by agar disc diffusion. Serial concentrations of the extract were tested (1, 2, 3, 4 and 5mg/ml). Most of the extract concentrations showed a relatively high antibacterial activity against all the tested bacteria. The concentration 5mg/ml of plant extracted has the maximum antibacterial activity against all bacterial isolates. Antimicrobial activity of *Myrtus communis* extract was compared with a number of antibiotics like: Ampicilin (AM10), Tetracycline (TE30), Erythromycin (E15), Carbncilin (PY100) and Gentamycin (CN10) using antibiogram test. Antibacterial activity of latest concentration (5mg/ml) of *Myrtus communis* was better than that to Gentamicin against *Klebsiella pneumoniae*, *Escherichia coli* and *Pseudomonas aerogenosa*.

Key words: Biological Effect, *Myrtus communis*, Pathogenic Bacteria

تأثير المستخلص الميثانولي لنبات الآس على بعض أنواع البكتريا الممرضة

الخلاصة

تم في هذه الدراسة التحري عن الفعالية التضادية للمستخلص الميثانولي لنبات الآس ضد بكتريا *Escherichia coli*, *Klebsiella pneumonia*, *Pseudomonas aerogenosa* and *Staphylococcus aureus* باستخدام طريقة الانتشار. سلسلة من التراكيز للمستخلص تم اختبارها

(١، ٢، ٣، ٤ و ٥ ملغم/مل). معظم التراكيز المحضرة من المستخلص اظهرت فعالية تضادية للأنواع البكتيرية قيد الدراسة. من بين التراكيز المحضرة من المستخلص الميثانولي كان التركيز الأخير ٥ ملغم/مل هو الأكثر فعالية حيث أعطى أكبر منطقة تثبيط. وقد تم مقارنة الفعالية التثبيطية للمستخلص الميثانولي بعدد من المضادات الحيوية (الامبيسيلين ١٠ مايكروغرام، تتراسايكلين ٣٠ مايكروغرام، الأثرثرومايسين ١٥ مايكروغرام، كاربينيسيلين ١٠٠ مايكروغرام و جنتاميسين ١٠ مايكروغرام) المعروفة بقدرتها على تثبيط البكتيريا قيد الدراسة. أعطى التركيز الأخير (٥ ملغم/مل) من المستخلص الميثانولي لنبات الأس فعالية تضادية أعلى من المضاد الحيوي *Klebsiella pneumoniae*, *Escherichia coli* and *Pseudomonas aeruginosa*.

INTRODUCTION

Traditional systems of medicine continue to be widely practised on many accounts. Population rise, inadequate supply of drugs, prohibitive cost of treatments, side effects of several allopathic drugs and development of resistance to currently used drugs for infectious diseases have led to increased emphasis on the use of plant materials as a source of medicines for a wide variety of human ailments [1]. Global estimates indicate that 80% of about 4 billion population cannot afford the products of the Western Pharmaceutical Industry and have to rely upon the use of traditional medicines which are mainly derived from plant material; this fact is well documented in the inventory of medicinal plants, listing over 20,000 species [2]. In spite of the overwhelming influences and our dependence on modern medicine and tremendous advances in synthetic drugs, a large segment of the world population still likes drugs from plants [3]. In many of the developing countries the use of plant drugs is increasing because modern life saving drugs are beyond the reach of three quarters of the third world's population although many such countries spend 40-50% of their total wealth 4 on drugs and health care [4]. As a part of the strategy to reduce the financial burden on developing countries, it is obvious that an increased use of plant drugs will be followed in the future [5].

Myrtus communis (Myrtaceae) is an evergreen sclerophyll shrub or small tree, the leaves emit an aromatic and refreshing smell somewhat reminiscent of myrrh or eucalyptus; the taste is very intensive, quite unpleasant and strongly bitter. The plant grows abundantly in the North Western to Eastern Mediterranean [6]. Myrtle is a spice finding no wide application because of its bitterness, despite the pleasant odor. Its culinary importance is limited to the region of origin. Foods flavoured with the smoke of myrtle are common in rural areas of Italy or Sardinia [7]. The most important constituents of myrtle oil are myrtenol, myrtenol acetate, limonene, linalool, α -pinene, 1,8-cineole, α -caryophyllene in addition to p-cymene, geraniol, nerol and the phenylpropanoid, methyleugenol [8]. However, there is considerable variability in the composition of oil from different locations [9]. In folk medicine, the fruit of this plant is used in the treatment of many types of infectious disease, including diarrhea and bloody diarrhea, and the leaves are used as antiseptic and anti-inflammatory agent, and as a mouthwash, for the treatment of candidiasis [10, 11]. So in this study we investigated the antibacterial properties of methanolic extract of *myrtus communis* against some pathogenic bacteria.

MATERIALS AND METHOD

Plant extraction

Leaves of *Myrtus communis* (Myriaceae) were collected, washed thoroughly in tap water and dried at dark room temperature for one week. The dried leaves were ground using an electric blender, 25g of the ground material (Myriaceae) placed in a conical flask and 100 ml of methanol was added to the flask and put on a rotary shaker at 180 rpm for 24 h. The crude extracts were obtained by filtration through (Whatman No.1) filter paper. The filtrate was concentrated and dried at 60 °C in oven. Extract was then prepared to 1, 2, 3, 4 and 5mg/ml by using sterilized distilled water [12].

Microorganisms

The test microorganisms, *Escherichia coli*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa* and *Staphylococcus aureus* were obtained from Central Health Laboratory in Baghdad. The organisms were inoculated onto Nutrient Broth (Hi-media) and incubated at 37 °C for overnight.

Disc diffusion assay

The disc diffusion method of Iennette [13] was used with some modification to determine the rate of growth inhibition of bacteria by the examined plant extracts. The isolated suspensions broth were adjusted to an optical density of 0.5 McFarland, and then streaked with a sterilized cotton swab on plates containing Muller Hinton Agar (Hi-media). Sterilized Filter paper discs (6 mm diameter) which impregnated with a different concentrations of crude extract (1, 2, 3, 4 and 5mg/ml) were placed on the inoculated agar surfaces and incubated for 24 hours. Antibiotic standard discs of erythromycin (E15), Ampicillin (AM10), Tetracycline (TE30), Carbapenem (PY100) and Gentamycin (CN10) supplied from (Difeco) were tested against the bacterial isolates and with same above conditions. Inhibition zone diameter measurement after incubation period of 24hr.

RESULT AND DISCUSSION

The presented results show that Gram-positive bacteria *Staphylococcus aureus* were more resistance than Gram-negative bacteria *E. coli*, *Klebsiella pneumoniae* and *Pseudomonas aeruginosa* Table(1).

Table(1): The mean of inhibition zone of the methanolic extract of Myriaceae against *Klebsiella pneumoniae*, *Escherichia coli*, *Pseudomonas aeruginosa* and *Staphylococcus aureus*.

Bacteria isolate	Inhibition zone (mm) at concentration (mg/ml)*				
	1	2	3	4	5
<i>Klebsiella pneumoniae</i>	—	—	10	18	24
<i>Escherichia coli</i>	—	—	—	20	24
<i>Pseudomonas aeruginosa</i>	—	8	12	18	22

Staphylococcus aureus	—	—	6	10	16
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* Myriaceae extract

The antibacterial activities of the extract of the plant varied in relation to the test organisms. The most active was 4 and 5 mg/ml concentration that inhibited completely the growth of all tested bacteria Figure (1). Abdullah et al. [14] reported that Gram-negative bacteria are generally more resistant compared to the Gram-positive ones. Also, Shan et al. [15] reported that Gram-positive bacteria (*L. monocytogenes*, *Staphylococcus aureus* and *B. cereus*) were generally more sensitive to the tested extracts than Gram-negative (*Escherichia coli* and *Salmonella anatum*). *Pseudomonas aeruginosa* was the most sensitive, while *E. coli* was the most resistant.

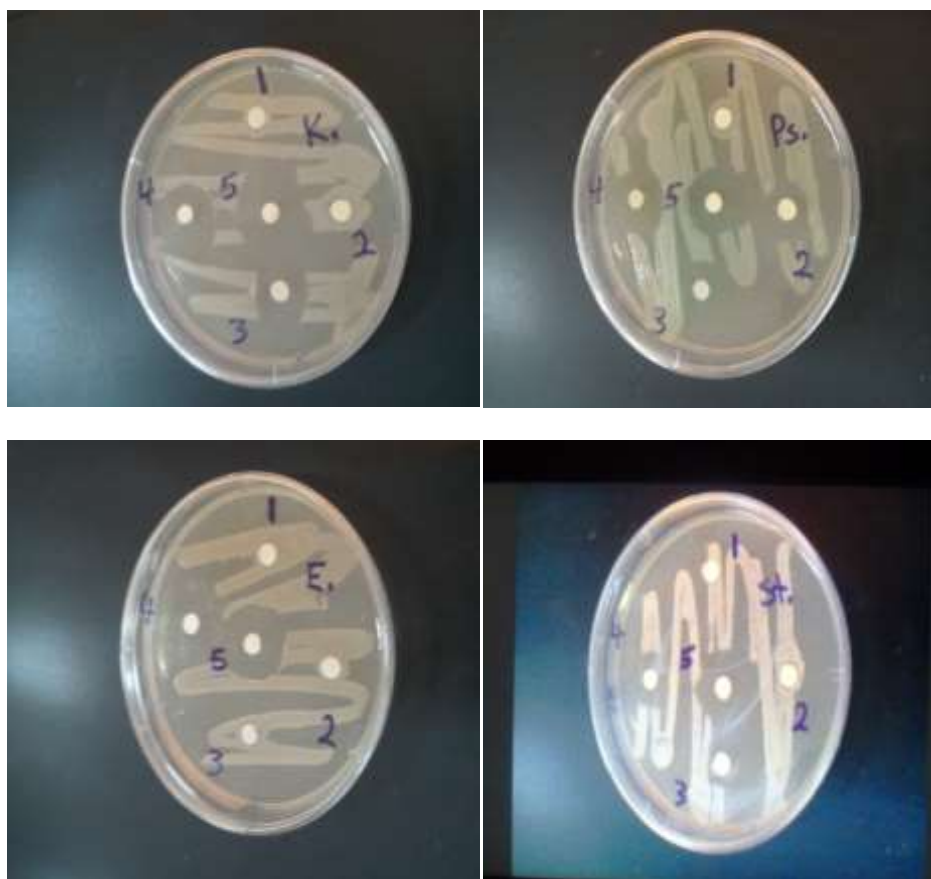


Figure (1): The antibacterial activity of methanolic extract of Myriaceae against *Klebsiella pneumoniae*, *Escherichia coli*, *Pseudomonas aeruginosa* and *Staphylococcus aureus*.

A possible explanation for these observations may lie in the significant differences in the outer layers of Gram-negative and Gram-positive bacteria. Gram-negative bacteria possess an outer membrane and a unique periplasmic space not

found in Gram-positive bacteria [16]. The resistance of Gram-negative bacteria towards antibacterial substances is related to the hydrophilic surface of their outer membrane which is rich in lipopolysaccharide molecules, presenting a barrier to the penetration of numerous antibiotic molecules. It is also associated with the enzymes in the periplasmic space, which are capable of breaking down the molecules introduced from outside [17]. Gram-positive bacteria do not possess this type of outer membrane and cell wall structure. Antibacterial substances can easily destroy the bacterial cell wall and cytoplasmic membrane and result in a leakage of the cytoplasm and its coagulation [18].

Antibacterial activity of methanolic extract of Myriaceae was compared with number of antibiotics that known for their ability included, Ampicilin (AM10), Tetracycline (TE30), Erythromycin (E15), Carbncilin (PY100) and Gentamycin (CN10) using antibiogram test Table (2).

Table (2): The inhibition zone of standard discs against some pathogenic bacteria.

bacteria strain	Zone of inhibition(mm)				
	Ampicilin	Tetracylin	Carbncilin	Gentamicin	Erythromycin
Klebsiella pneumonia	R	R	R	6	R
Escherichia coli	R	10	R	12	R
Pseudomonas aerogenosa	R	R	R	10	R
Staphylococcus aureus	6	4	R	20	R

Antimicrobial activity of latest concentration of of methanolic extract of Myriaceae was better than that to gentamicin against *Klebsiella pneumoniae*, *Escherichia coli*, *Pseudomonas aerogenosa* and *Staphylococcus aureus*. The resistance of pathogenic bacteria to Carbncilin, Ampicilin and Erythromycin was noticed Figure (2). The frequent use of antibiotics stimulates the emergence of new strains of pathogenic bacteria, show resistance to these antibiotics and these findings in our study and in other studies.

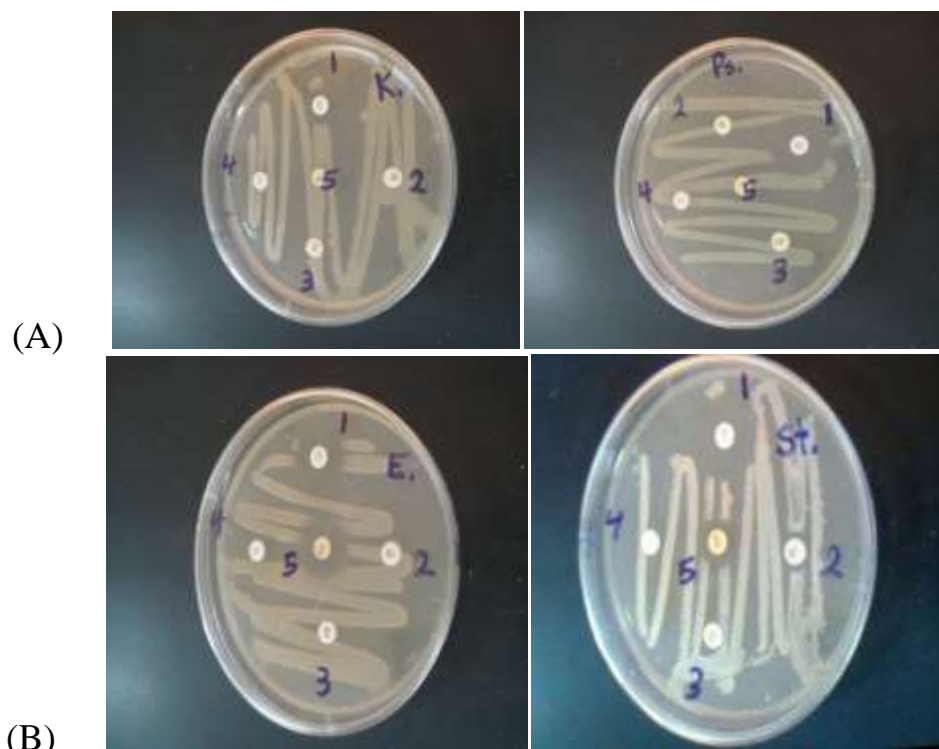


Figure (2): The antimicrobial activity of some antibiotics
(A) against *Klebsiella pneumoniae*, *Escherichia coli*,
(B) *Pseudomonas aeruginosa* and *Staphylococcus aureus* .

In this study, the extract was significantly active against Gram-positive bacteria and Gram-negative bacteria. Some studies claim that the phenolic compounds present in spices and herbs might also play a major role in their antimicrobial effects [19]. There has been no large scale, systematic investigation of the relationship between bacterial inhibition and the total phenolic content of spices and herbs. Previous studies [20] showed that a highly positive linear relationship exists between antioxidant activity and total phenolic content in some spices and herbs.

According to the findings of this study, most of the extract concentrations (1, 2, 3, 4 and 5mg/ml) showed relatively high antibacterial activity against all the tested bacteria.

Finally we can conclude from this study that the active compound of leaf extract is a potential source of natural antibacterial agents.

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