Study of some biochemical marker in abortion women infected with Toxoplasmosis

Widad hashim yahay Al-Mahana College of Education for Girls , Kufa University Email – Wadad Hashim yahya@ gmail. com

Abstract

The current study consisted of 60 medical cases infected with Toxoplasmosis as well as 30 cases of uninfected women, all who attended Al-Sadr Medical City and Al-Hakeem Hospital in the holy city of Najaf from December to August 2014. The study was designed for the purpose of determining the levels of "Bilirubin, Albumin, and Creatine" in women who have the "*Toxoplasma gondi*" parasite in Najaf by using the latex test. The current study shows that there is a significant difference (p <0.05) in the concentration of bilirubin and albumin, the value of bilirubin was (0.53*mg/dl) for those who have parasite compared with the control group which reached (0.99mg/dl) while the value of albumin (0.98*mg/dl) compared with the control group that reached (6.88mg/dl). However, the study found there to be a significant increase (p<0,05) in the concentration of creatine for those who have toxoplasmosis when its value reached (4.6mg/dl) compared with the control group which reached a value of (1.2*mg/dl).

Introduction

Toxoplasma gondii is the most frequent protozoan causing opportunistic infection in immunocompromised individuals (1). Several researches have been carried out on the immune response in patients with Chronic renal failure (CRF) and proved there was impairment of cell-mediated immunity (2), also these patients have significant lower immune response to vaccines such as tetanus vaccine than healthy subjects (3).CRF patients are under risk from a variety of infections (4).

Acute T. gondii infections are asymptomatic and self -limiting, especially among healthy immunocompetent hosts. However the infection may cause severe complications in pregnant women and immunocompromised patients (5,6), such as HIV/AIDS patients (7), cancer patients (8), end stage renal disease undergoing hemodialysis patients (9) and those having organ transplantation (10). In the latter setting, the disease can result from T. gondii transmission within the allograft from a seropositive donor into a seronegative recipient (11).

T. gondii has complex life cycle consisting of three stages. Tachyzoite, develops during the acute stage of infection, invades and replicates within the cell. Bradyzoite (pseudocysts) develops during latent infection and presents in tissue cysts (12). The first and second stages represent the asexual development of the life cycle which occurs in the intermediate hosts of parasite including man. The

third stage (oocysts) represents the sexual development of life cycle which occurs in intestinal tissue of cat only (13,14).

Strains of *T.gondii* exhibit virulence differences, studies with mice have shown that infections with the different clonal lineages of T. gondii result in very different outcomes Type I strains are highly virulent; whereas type II and type III are relatively nonvirulent (15,16). All three genotypes can cause human infections, however the genotype II predominates in human toxoplasmosis (17)Genotypes I and II occur more frequently in AIDS and congenitally infected patients than in animals, whereas the genotype III predominates in animals (17). However in the case of severe ocular disease in man, strains of the genotype I and recombinants of genotype I and III are more common than the genotype II (18). These findings suggest that type I strains may be more likely to cause sever disease in humans (19). Aim of the present study to defermine the level of some biochemical parameter in women that infection with toxoplasmosis.

Material and methods

Latex agglutination test

The kit is provided from Linear Chemicals-Spain, where the principle of the test is based on antigen – antibody reaction directly. The sensitiivity of the test is 10 IU/ml (20). The kit included:

1-Toxoplasmosis latex reagent: Suspension of polystyrene latex particles coated with *T. gondii*

soluble antigen in buffer containing bovine serum albumin < 0.1 % sodium azide.

- 2- Positive control: Diluted human serum contains rabbit IgG anti *Toxoplasma* containing < 0.1 % sodium azide.
- 3- Negative control: No reactive diluted human serum contains < 0.1 sodium azide.

Test procedure

it was done according to a procedure described by (20) which is included:

Qualitative test: 50µl of serum was mixed gently with one drop of *Toxoplasma* latex reagent, for 5min. The macroscopic visible of agglutination indicated the positive reaction and smooth suspension with no visible agglutination indicated negative reaction.

Estimation of Serum Albumin

This test was performed by using albumin liquid reagent (Biomaghreb, Maghreb).

The principle

Albumin, in a buffered solution, reacts with of bromocresol green to form a red-color complex.

Reagents

1-Reagent 1 Bromocresol green 0.14g/l

Succinate buffer 75mml/l

Brij 7ml/l

2-Reagent 2 (standard) Bovine Albumin

50g/1

The procedure:

1. All the reagent was left at 25C before used.

- 2. From standard reagent, 10µl was mixed with 2ml of reagent 1.
- 3. From each blood sample 10µl also mixed with 2ml of the same reagent.
- 4. After 5 minutes the optical density of each sample and the standard reagent were read against the blank at the wavelength of 628nm.

Estimation of Serum creatinine

This test was performed by using creatinine liquid reagent (Syrbio diagnostic reagents laboratories, Syria). According the following:

The procedure:

- 1. Mixing 100µl of the standard reagent with 1ml of working reagent.
- 2. Then 100µl of each sample was mixed with 1ml of working reagent.
- 3. After 30 second, the optical density (O.D1) of each sample was read at

500nm by using spectrophotometer.

Second reading was obtained (O.D2) at exactly,
 Iminute after the first
 reading,

Statistical analysis

The stastical analysis were conducted by using (Graph ped prism)

Results

The statistical analysis of the current study showed a highly significant decrease (P < 0.05) in serum bilirubin concentration (0.53^* mg/dl) of patients with T. gondii infection compared to the control group (0.99mg/dl); as seen in Figure (1 The statistical analysis of the current study showed a highly significant decrease (P < 0.05) in serum albumin concentration (2.98*mg/dl) of patients with T. gondii infection compared to the control group (6.88mg/dl); as seen in Figure (1).) . The statistical analysis of the current study showed a significant increase (P < 0.05) in serum creatinine concentration (4.8 mg/dl) of patients with T. gondii infection compared to the control group

Discussion

The current study has revealed that the serum albumin and bilirubin significantly decrease in *T. gondii* infection patients compared to control

group. While the level of creatinine was significantly increase in patients infected with T. gondii in compared with control group. The decrease of albumin level in the serum of patients

(1.2*mg/dl); as seen in Figure (1)

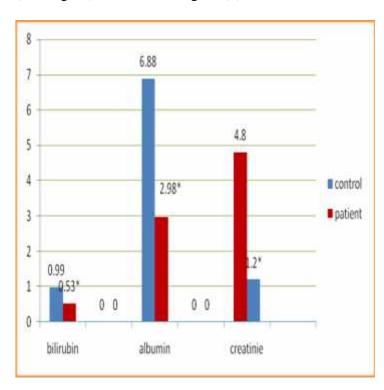


Figure (1) Showed bilirubin , Creatiinie and
 Albumin in patient compared with control group

may be due to increasing in it's the catabolism or due to decrease in the albumin synthesis. The increase in creatinine concentrations in infected group may be explained as *Toxoplasma* parasite causes glomerular lesions and urinary abnormalities which lead to decrease in glomerular filtration rate, is typically detected by an elevated serum creatinine level in the urine (21). Toxoplasmosis causes extensive and progressive damage to the liver remarkable proliferations of organisms such

damage in the liver metabolism (22) and toxoplasmosis Infects liver cells and leads to inflated cells as provided and expansion in central necrosis of hepatic veins and damage in various parts of the Liver (22; 23). Changes of protein fractions AST, AIT varied according to the qualitative difference in the intensity of inflammation by strains of *Toxoplasma*.

Bilirubin showed decrease below the normal value in all studied samples especially ALT, AST, total protein and globulin indicated that the treatment improve the immune system and slow rate of hepatocytes metabolism either by an increase the anabolism and decrease catabolism.

References

- [1] **Ferreira**, M. and Borges, A.S. (2002). Someaspects of protozoan infection immunocompromised patients: A review. Mem. Inst. Vol. 97(4): 443-457.
- [2] Langhoff, E. and Ladefoyed, J. (1988). *In vitro* immune function in patients with minor, moderate and severe kidney impairment. AMPIS, 96: 655-659.

 [3] Sotoodeh, J.; Raoofi, A.; Sarikhani, M. and
- Madani, A. (2009). Evaluation of anti-tetanus immunity in haemodialysis patients. Am. J. Immunol., 5: 108-112.
- [4] Assarehzadegan, M.A.; Shakerinejad, G.; Noroozkohnejad, R.; Amini, A. and Rahim S.A.
- (2009). Prevalence of hepatitis C and B infection and HC V Rezaee, genotypes among hemodialysis patients in Khuzestan province, southwest Iran. Saudi J. Kidney Dis. Transpl., 20: 681-684.

- [5] Holland, G.N.; O'Connors, G.R.; Belfort, J.R. and Remington, J.S.(1996). Toxoplasmosis. p. In Peprose, J.S., Holland,G.N. and Wilhelmus, K.R. (eds), Ocular infection and immunity.Mosby Yearbook, St. Louis.P. 1183-1223.
- **[6] Espinoza**, L. A. 2005. Toxoplasmosis. Caribbean AIDS education and training center. HIV primary care guide. Chapter11, section 6:1-4.
- [7] Lindström, I.; Kaddu-Mulindwa, D.H.; Kironde, F.; Lindh, J.(2006).Prevalence of latent and re-activated *Toxoplasma gondii* parasites in HIV-patients from Uganda Oswaldo Cruz, 97: 443-457.
- [8] Rai, S.K.; Upadhyay, M.P.; Shrestha, H.G. (2003). *Toxoplasma* infection in selected patients in Kathmandu, Nepal. Med. Coll. J. 5:89-91.
- [9]Yazar, S, Demirtars, Fyaicin, S, yaman, O., Tokgoz, B., utas C. & Sahin, I.(2003) Anti-Toxoplasma gondii antibodies in haemodialysis patients with chronic renal failner – yorisei med J 44, 288-292.
- [10]Sukthana, Y.; Chintana, T.; Damrongkitchaiporn, S. and Lekkla, A.(2001). Serological study of *Toxoplasma gondii* in kidney recipients.J. Med. Assoc. Thai. 84: 1137-1141.
- [11] Assi, M.A; Rosenblatt, J.E. and Marshall, W.F. (2007). Donortransmitted toxoplasmosis in liver transplant recipients: a case report and literature review. Transpl Infect Dis. 9:132-136.
- [12] Wilson, M.; McAuley, J. (1999). *Toxoplasma*: Manual of clinical microbiology: 1374-1382.
- [13] Zeibig, E. A. (1997). Protozoa. In: Clinical Parasitology: A practical approach. Zeibig, E. A.

- (ed.) W. B. Saunders Company, Philadelphia,: 125Pp.
- [14] **Dubey,** J.P. (2006).Comparative infectivity of oocysts and bradyzoite of *Toxoplasma gondii* for intermediate (mice) and definitive (cat) hosts. Vet. Prasatol.140:69-75.
- [15] Susuki, Y., Conley, F.K. and Remington, J. S. Impartance of endogenors IFN- gamma for prevetion of Toxoplasmic encephalitis in mice J Immuno . 1989 . 143: 2045- 2050.
- [16] Khan, I.A.; Schwartzman, J.D.; Matsuura, T. and Kasper, L.H. (1997). A dichotomous role for nitric oxide during acute *Toxoplasma gondii* infection in mice. Proc. Nat. Acad. Sci., 94:13955-13960
- [17] Howe, D.K. and Sibley, L.D. (1995). *Toxoplasma gondii* comprises three clonal lineages: correlation of parasite genotype with human disease. J. Infect. Dis. 172: 1561-1566.
- [18] Grigg, M.E.; Ganatra, J.; Boothroyd, J.C. and Margolis, T.P. (2001). Unusual abundance of. atypical strains associated with human ocular toxoplasmosis. J. Infect. Dis. 184:633-639.
- [19] Sibley, L.D.; Mordue, D.G.; Su,C.; Robben, P M. a nd Howe, D.K. (2002). Genetic approaches to studying virulence and pathogenesis in *T. gondii*. Phil. Trans. R. Soc. Lond. 357: 81 88.
- [20] Roller, A.; Bartlett, A. and Bidwell, D.E. (1987). Enzyme immunoassay with special reference ELISA technique. J. Clin. Path. 31:507-520.
- [21] Gharadaghi, Y., Shojaee, S., Khaki, A., Fathiazad, F., Khaki, A., Ghdamkheir, E. and

- Rouhaninia, M. **2012**. Antiprotozoal effect of *Allium cepa* on acute renal failure caused by
- Toxoplasma gondii. Afr. J. Pharm. Pharmacol., 6(10),:771-777.
- [22] Hokeleck, M.; Kiling, M.; Erturt, M. and Yyar, Y. (2001). Production of *Toxoplasma gondii* in human amnion cell cultures. Turt. Parasite. Derg., 25(4):323-325.
- [23] Khan, A.; Su, C.; German, M.; Storch, G.A.; Clifford, D.B; and David Sibley, L. (2005). Genotyping of *Toxoplasma gondii* Strains from Immunocompromised Patients Reveals High Prevalence of Type I Strains J. Clin. Microbiol. 43(12): 5881–5887.

•

()

/

Email - Wadad Hashim yahya@ gmail. com

30

60

. 2014

Toxoplasma gondii

(P<0.05) Latex test

(0.53 mg/dl) T. gondii

(2.98* mg/dl) (0.99mg/dl)

> (P<0.05) . (6.88 mg/dl)

> > (4.6 mg/dl)

. (1.2*mg/dl)