

### **Iraqi Journal of Veterinary Sciences**



www.vetmedmosul.com

# Molecular and pathological identification of ovine pulmonary adenomatosis at Mosul city abattoirs

### O.Z. Jiad<sup>1</sup><sup>©</sup> and A.M. Al-Saidya<sup>2</sup><sup>©</sup>

<sup>1</sup>Veterinarian, Private Sector, <sup>2</sup>Department of Pathology and Poultry Diseases, Faculty of Veterinary, Mosul University, Mosul, Iraq

Article information	Abstract
Article history: Received 21 May, 2024 Accepted 09 September, 2024 Published online 01 January, 2025	Ovine Pulmonary adenomatosis OPA is a frequent condition that affects small ruminants, characterized by inflammation of the lung parenchyma, and causes polyp-like hyperplasia of the epithelium lining the alveoli and interstitial tissue like bronchi. It is frequently brought on by the Jaagsiekte virus. Samples from sheep animals at Mosul's
<i>Keywords</i> : Lung Pathology PCR Sheep	slaughterhouse and butcher shops were collected. The RT-PCR results consider the first molecular identification of the JSRV virus in Mosul city. The Macroscopic examination revealed gray-colored tumor nodules scattered across the surfaces of the lung lobes. The tumor tissue had clear and distinct boundaries from the adjacent pink pneumatic lung.
Correspondence: A.M. Al-Saidya al2011saidya@uomosul.edu.iq	Histological analysis showed nodular tumor areas (proliferative pulmonary nodules) were composed of carcinomatous cells supplementary with an adequate amount of extracellular matrix, characteristic columnar and cuboidal metaplasia, as well as hyperplasia of pulmonary cells, especially the pneumocytes (type II), an epithelial cell (secretory types) in the pulmonary alveoli, Non-ciliated Clara cells as well as the epithelial lining of the terminal bronchioles. Finally, our study sheds light and provides data on the importance of OPA in Mosul abattoir through the PCR diagnosis of the causative agent, and the

macroscopic and histological pathological changes.

DOI: <u>10.33899/ijvs.2024.148936.3622</u>, ©Authors, 2025, College of Veterinary Medicine, University of Mosul. This is an open access article under the CC BY 4.0 license (<u>http://creativecommons.org/licenses/by/4.0/</u>).

#### Introduction

Ovine pulmonary lesions" are a lung disease that can affect sheep. These diseases can lead to serious health problems, and they can even be fatal. Ovine pulmonary lesions can be challenging to diagnose and can be mistaken for other types of diseases (1,2). Jaagsiekte ovine Retrovirus (JSRV) is a  $\beta$  - retrovirus etiology of ovine pulmonary adenomatosis (OPA) or ovine pulmonary adenocarcinoma, which is a contagious pulmonary and cancerous disease of sheep (3). The virus is present in respiratory discharges and mainly replicates inside the pulmonary alveolar cells (4,5). Jaagsiekte disease spreads primarily through inhaling infected respiratory secretions via airborne transmission and can also spread through milk

and colostrum. Ovine pulmonary adenomatosis is confined to the lungs and, in rare instances, the lymph nodes are affected. They appear as hard, white to gray and welldefined nodules of varying sizes, ranging from small nodules to large solid areas encompassing one or more pulmonary lobes (6). The conducting airways contain a significant amount of white, foamy fluid. The diseases are characterized histologically by papillary proliferation of columnar type II pneumocytes and associated cells in the bronchioles (7,8). The current study aimed to demonstrate the pathological and molecular detection of ovine pulmonary adenomatosis in Mosul, Iraq.

#### Materials and methods

#### **Ethical approval**

The Ethical Committee for Animal Experimentation of the Veterinary Faculty of the University of Mosul approved the experimental work U.M.VET.2024.015.

#### **Collection of a lung sample**

A total of 11 lung samples were collected from previous work from sheep suspected of OPA, where 25 (16%) Lung samples had been revealed pulmonary adenomatosis lesions at Mosul's slaughterhouse and butcher shops were collected during weekly inspections.

#### Gross and histopathological evaluation

The collected specimens were examined obviously to record any unusual lesions in the lung that appear by the naked eve or by palpation and observation noting any changes in the lung's consistency, size, and color. Two parts of the samples were taken from the lesion areas. The first part was placed in a buffer solution of formalin at a concentration of 10% for 48 hours or more for fixation. Histopathological sections were carried out through routine paraffin embedding technique, firstly dehydrated with ascending ethanol concentration, clearance with xylol and embedded with paraffin wax (9,10) then sectioned about 5-6 µm in thickness and finally, stain with routine hematoxylin and eosin stain (11). For the second part of the diagnosis, suspected samples were placed in small plastic containers, and tests using polymerase chain reaction (PCR) technology to diagnosis the virus that causes OPA cases.

## Molecular detection of ovine pulmonary adenomatosis virus

RNA was extracted from suspected lung tissue samples using (Gena Bioscience, Germany kit) accurately per the manufacturer's instructions. Detection of the JSRV virus using a conventional RT-PCR kit was provided by Bioron GmbH, Germany. One RT-PCR Master Mix is designed for all applications requiring Reverse RNA transcription into cDNA and subsequent PCR amplification. Only primers and sample RNA were added to the Master mix. We used Forward/ 5' Reverse primer TGGGAGCTCTTTGGCAAAAGCC-3 5'-TGATATTTCTGTGAAGCAGTGCC-3, respectively (12,13). The primers were manufactured by Macrogen Corporation, Korea. According to the manufacturer's instructions, the subsequent optimal reaction was at 55 °C for 30 min reverse transcription. RNA denaturation, and inactivation of reverse transcriptase at 95 °C for 3 min. Then, 40 cycles PCR, denaturation at 92 for 10 s, annealing at 62 °C for 10 s, extension at 72 °C for 20 s, and extension at 72 C° for 7 min. The RT-PCR technique was achieved on the MiniAmp PlusTM Thermocycler PCR, USA. Electrophoresis (Cleaver Scientific, England) was prepared on 1.5% agarose gel holding gel-safe stain to separate charged DNA fragments according to size. RT-PCR bands were 176 bp JSRV cDNA.

#### Results

#### **RT-PCR** for Ovine Pulmonary adenomatosis virus

All 11 lungs tissue showing gross and microscopic characteristic features for ovine pulmonary adenomatosis were positive for the JSRV virus and located in lanes 2-12 (Figure 1).

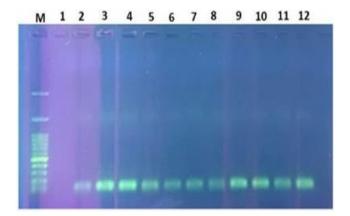


Figure 1: Positive JSRV virus is located in lanes 2-12 (176 bp), whereas negative control is in lane 1. Lane M: 100 bp DNA ladder (Addbio company, Korea).

## Pathological characteristic of ovine pulmonary adenomatosis

The Macroscopic examination of lung samples suspected of being infected with OPA revealed graycolored tumor nodules scattered across the surfaces of the lung lobes. The tumor tissue had clear and distinct boundaries from the adjacent pink pneumatic lung. Additionally, pneumonia lesions showing congestion and consolidation were observed. The Histological analysis of a lung suspected with OPA, showed nodular tumor areas (proliferative pulmonary nodules) were composed of carcinomatous cells supplementary with an adequate amount of extracellular matrix, characterized by hyperplasia of pulmonary cells, especially the pneumocytes (type II), an epithelial cell (secretory types) in the pulmonary alveoli, Non-ciliated Clara cells as well as the epithelial lining of the terminal bronchioles. These tumors cuboidal or columnar types of cells exchange the typical alveolar cells and form glandular acinar, papillary, and mixed architectural arrangements. Some cancer cells were observed shedding and sloughing inside the lumen of alveoli, along with thickening of alveolar walls due to infiltration of inflammatory cells and deposition of collagen fibers (Figures 2-7).



Figure 2: Macroscopic images of Ovine lungs with OPA suspected revealed signs of pneumonia like congestion and consolidation as well as the presence of gray nodules with clear and distinct boundaries between the tumor tissue and the adjacent airy pink lung.

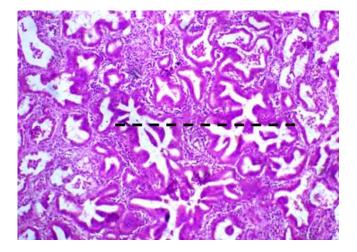


Figure 3: Histological section of ovine lung showing nodular tumor areas (proliferative pulmonary nodules) were composed of carcinomatous cells (H&E stain, 40x).

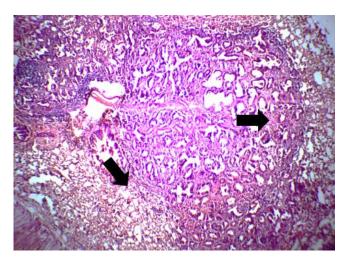


Figure 4: Histological section of ovine lung showing nodular tumor areas (proliferative pulmonary nodules) were composed of carcinomatous cells (H&E stain, 40x).

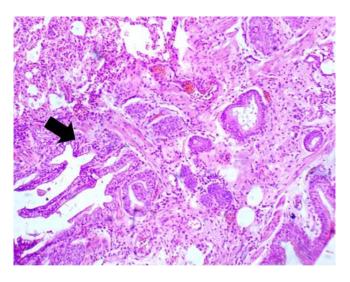


Figure 5: Histological section of ovine lung showing hyperplasia of epithelial cells lining the alveoli and bronchi, forming a glandular acinar, papillary and mixed architectural arrangements, with Some cancer cells were observed shedding and sloughing inside the lumen of alveoli, along with thickening of alveolar walls (H&E stain, 100x).

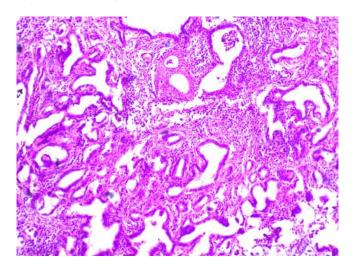


Figure 6: Histological section of ovine lung showing a glandular acinar, papillary and mixed architectural arrangements, with Some cancer cells, along with thickening of alveolar walls (H&E stain, 100x).

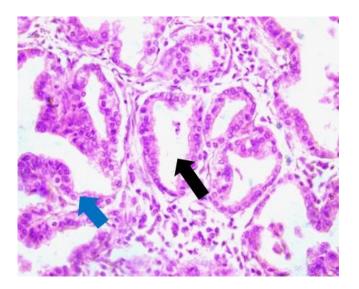


Figure 7: Histological section of ovine lung showing hyperplasia of epithelial cells lining the alveoli and bronchi, forming a glandular acinar, papillary and mixed architectural arrangements, as well as cuboidal and columnar metaplasia of the squamous epithelium lining the alveoli.

#### Discussion

Ovine pulmonary adenomatosis (OPA) has been documented in various countries globally, such as Iraq (13,14). Also known as Jaagsiekte, it's a contagious lung tumor in sheep caused by Jaagsiekte sheep retrovirus. JSRV can infect and transform the alveolar and bronchiolar secretory epithelial cells, causing the formation of tumors that may occupy significant portions of the lung. Furthermore, tumor growth frequently disrupts normal respiration leading to increased fluid production in the lung. This study is dedicated to detecting and identifying pathological, gross, and microscopic characterization and the definitive diagnosis of the viral causative factor of OPA using the PCR reaction in sheep at Mosul city abattoirs.

The results indicated that ovine lungs with characteristic macro and micro lesions were positive for the JSRV virus, this agreed with different previous researchers (15,16). PCR analysis can detect the JSRV virus from blood, lung samples, mediastinal lymph nodes and lung fluid (17,18). Our RT-PCR results consider the first molecular identification of the JSRV virus in Mosul city (19,20). Different studies categorized the Pathological findings of OPA into Classic and Atypical gross and microscopic features (21). The Classical type of the suspected OPA lung samples revealed a gray-colored tumor nodule scattered across the surfaces of the lung lobes. The tumor tissue had clear and distinct boundaries from the adjacent pink, pneumatic lung (22).

The Word Typical form of OPA, is described by multiple nodules in the lung lobes especially the diaphragmatic lobe, as well as large volumes of frothy fluid inside the conducting airways (23,24). Microscopically, The Classical OPA included hyperplasia of pulmonary cells, especially the pneumocytes (type II), an epithelial cell (secretory types) in the pulmonary alveoli, Non-ciliated Clara cells as well as the epithelial lining of the terminal bronchioles (25,26). The OPA virus (JSRV) replicates inside these types of pulmonary cells, which express a particular type of cellular receptor called Hyal-2 considered the primary receptor for JSRV (27,28). JSRV viral envelope glycoprotein, surface protein attached to the Hyal-2 receptor, leading to enter the virus by the endocytic process. The main multiplying cancerous cell type, signifying as 60-80% of cell residents, were epithelial cells in alveoli type II (Pneumocytes) with papillary or acinar like structures noted. as well as the proliferative Clara cells as 15- 20% (29,30). All lung samples with OPA lesions also show multiple degrees of inflammatory lesions, like inflammation of bronchitis, bronchiolitis, interstitial pneumonitis, and inflammatory cell reaction as a pulmonary tissue defense mechanism (31,32). One of the significant causes described by OPA as a malignant type of tumor is metastasis. Unusually, most metastases of the primary tumor are microscopically found in the mediastinal lymph nodes but in only 10% of the ovine Pulmonary adenocarcinoma (OPC) cases (33,34). R-PCR and immunohistochemical studies for JSRV capsid protein markers were used to identify the metastasis of OPC in other tissues and organs (35,36).

#### Conclusions

The current study concluded that JSRV was identified molecularly using R-PCR. This was further confirmed by the fact that a significant percentage of the Pulmonary samples displayed the typical macroscopic lookand all the typical histological abnormalities associated with the OPA characteristic lesions.

#### Acknowledgment

We appreciate each person who presented the possibility of completing our research, which is for the most part conducted by the Veterinary Medicine College, Pathology and Poultry Diseases Department at Mosul University.

#### **Conflict of interest**

The investigator announces there is no conflict of interest.

#### References

- Shivasharanappa N, Dheeraj Reddy BN, Apoorva KN, Rashmi L, Suresh KP, Gulati BR, Patil SS. Ovine pulmonary adenocarcinoma (OPA) in sheep: An Update on epidemiology, pathogenesis and diagnosis. J Exp Biol Agric Sci. 2023;11(6):997–1009. DOI: 10.18006/2023.11(6).997.1009
- Azizi S, Tajbakhsh E, Fathi F. Ovine pulmonary adenocarcinoma in slaughtered sheep: A pathological and polymerase chain reaction study. J S Afr Vet Assoc. 2014;85(1):1-5. DOI: <u>10.4102/jsava.v85i1.932</u>
- Mustafa ES, Al-Jameel WH, Al-Mahmood SS. Immunohistochemical detection of P53 and MDM2 and its correlation with histological grading system in ovine pulmonary adenocarcinoma. Iraqi J Vet Sci. 2021;35(4):687-92. DOI: <u>10.33899/ijvs.2021.127779.1527</u>
- 4. Lee AM, Wolfe A, Cassidy JP, Moriarty J, O'Neill R, Fahy C, Connaghan E, Cousens C, Dagleish MP, McElroy MC, Messam LL. An approach to diagnosis of Jaagsiekte sheep retrovirus infection in sheep based on assessment of agreement between macroscopic examination, histopathologic examination and reverse-transcriptase polymerase chain reaction. Small Rumin Res. 2019;181:29-33. DOI: 10.1016/j.smallrumres.2019.10.006
- Mansour KA, Al-Husseiny SH, Kshash QH, Jassim A. Clinicalhistopathological and molecular study of ovine pulmonary adenocarcinoma in Awassi sheep in Al-Qadisiyah Province, Iraq. Vet World. 2019;12(3):454. DOI: <u>10.14202/vetworld.2019.454-458</u>
- Quintas H, Pires I, Garcês A, Prada J, Silva F, Alegria N. The diagnostic challenges of ovine pulmonary adenocarcinoma. Ruminants. 2021;1(1):58-71. DOI: 10.3390/ruminants1010005
- Ortín A, De las Heras M, Borobia M, Ramo MA, Ortega M, de Arcaute MR. Ovine pulmonary adenocarcinoma: A transmissible lung cancer of sheep, difficult to control. Small Rumin Res. 2019;176:37-41. DOI: <u>10.1016/j.smallrumres.2019.05.014</u>
- Mishra S, Kumar P, Dar JA, George N, Singh V, Singh R. Differential immunohistochemical expression of JSRV capsid antigen and tumour biomarkers in classical and atypical OPA: A comparative study. Biol Rhythm Res. 2021;52(6):946-56. DOI: 10.1080/09291016.2019.1610857
- Al-Mahmood SS, Farhan AM, Daoud ZS, Hamed OS. Pathological study of liver lesions in cattle slaughtered at Kirkuk province abattoir. Iraqi J Vet Sci. 2017;31(1):7-16. DOI: <u>10.33899/ijvs.2017.126713</u>
- Al-Sabawy HB, Rahawy AM, Al-Mahmood SS. Standard techniques for formalin-fixed paraffin-embedded tissue: A pathologist's perspective. Iraqi J Vet Sci. 2021;35(1):127-135. DOI: <u>10.33899/ijvs.2021.131918.2023</u>
- Al-Mahmood SS. Improving light microscopic detection of collagen by trichrome stain modification. Iraqi J Vet Sci. 2020;34(2):473-481. DOI: <u>10.33899/ijvs.2020.126176.1256</u>
- Lee AM, Wolfe A, Cassidy JP, McV. Messam LL, Moriarty JP, O'Neill R, Fahy C, Connaghan E, Cousens C, Dagleish MP, McElroy MC. First confirmation by PCR of Jaagsiekte sheep retrovirus in Ireland and prevalence of ovine pulmonary adenocarcinoma in adult sheep at slaughter. Irish Vet J. 2017;70:1-2. DOI: <u>10.1186/s13620-017-0111-z</u>
- Al-Ajeli RR, Al-Qadhi AS, Al-Mahmood SS, Alkattan LM. Pathological study of neoplasms surgically excised from animals attended the veterinary teaching hospital. Iraqi J Vet Sci. 2021;35(1):9-14. DOI: <u>10.33899/ijvs.2019.126188.1260</u>
- 14. Shi W, Jia S, Guan X, Yao X, Pan R, Huang X, Ma Y, Wei J, Xu Y. A survey of Jaagsiekte sheep retrovirus (JSRV) infection in sheep in the three northeastern provinces of China. Arch Virol. 2021;166:831-40. DOI: <u>10.1007/s00705-020-04919-6</u>
- Sonawane GG, Tripathi BN, Kumar R, Kumar J. Diagnosis and prevalence of ovine pulmonary adenocarcinoma in lung tissues of naturally infected farm sheep. Vet World. 2016;9(4):365. DOI: <u>10.14202/vetworld.2016.365-370</u>
- Toma C, Bâlteanu VA, Tripon S, Trifa A, Rema A, Amorim I, Pop RM, Popa R, Catoi C, Taulescu M. Exogenous Jaagsiekte Sheep Retrovirus type 2 (exJSRV2) related to ovine pulmonary

adenocarcinoma (OPA) in Romania: Prevalence, anatomical forms, pathological description, immunophenotyping and virus identification. BMC Vet Re. 2020;16:1-5. DOI: 10.1186/s12917-020-02521-1

- Heras M, Gonzalez L, Sharp JM. Pathology of ovine pulmonary adenocarcinoma. In: Fan H, editor. Jaagsiekte sheep retrovirus and lung cancer. Germany: Springer Berlin, Heidelberg; 2003. 25-54 pp. DOI: <u>10.1007/978-3-642-55638-8</u> 2
- Jassim A, Al-Husseiny SH, Mansour KA, Kshash QH. First molecular diagnosis of ovine pulmonary adenocarcinoma in Awassi sheep in Iraq. Al-Qadisiyah J Vet Med Sci. 2017;16(1):112-7. DOI: <u>10.29079/vol16iss1art46</u>
- Abd Abass F, Khudhair YI. Clinical, molecular, and pathological investigations of ovine pulmonary adenocarcinoma in the middle of Iraq. Open Vet J. 2022;12(2):264-72. DOI: 10.5455/OVJ.2022.v12.i2.15
- Devi VR, Manasa BB, Samatha V, Mahesh M, Srikanth KV, Rao TY, Satheesh K, Kishore KN. Pathology of natural cases of ovine pulmonary adenocarcinoma (Jaagsiekte) in goats. Braz J Vet Pathol. 2016;9(3):108–112. [available at]
- Gebeyehu DT. A review on sheep pulmonary adenocarcinoma. J Adv Allergy Immunol Dis. 2017;2(1):1. DOI: <u>10.25177/JAAID.2.1.3</u>
- Ortega J, Corpa JM, Castillo D, Murphy BG. Pathological spectrum of ovine pulmonary adenocarcinoma in small ruminants: A focus on the mixed form. Animals. 2023;13(18):2828. DOI: <u>10.3390/ani13182828</u>
- Al-Husseiny S, Jassim A, Mansour KA, Kshash QH. Phylogenetic analysis of Jaagsiekte sheep retrovirus (JSRV) in Iraqi Awassi sheep. Iraqi J Vet Sci. 2020;34(2):351-5. DOI: 10.33899/ijvs.2019.126172.1255
- Belalmi NH, Sid N, Bennoune O, Ouhida S, Heras ML, Leroux C. Evidence of Jaagsiekte sheep retrovirus-induced pulmonary adenocarcinoma in Ouled Djellal breed sheep in Algeria. Vet Res Forum. 2020;11(1):93–95. DOI: <u>10.30466/vrf.2019.107160.2549</u>
- Cousens C, Alleaume C, Bijsmans E, Martineau HM, Finlayson J, Dagleish MP, Griffiths DJ. Jaagsiekte sheep retrovirus infection of lung slice cultures. Retrovirol. 2015;12:1-6. DOI: <u>10.1186/s12977-015-0157-5</u>.
- Griffiths DJ, Martineau HM, Cousens C. Pathology and pathogenesis of ovine pulmonary adenocarcinoma. J Comp Pathol. 2010;142(4):260-83. DOI: <u>10.1016/j.jcpa.2009.12.013</u>
- Miller AD. Hyaluronidase 2 and its intriguing role as a cell-entry receptor for oncogenic sheep retroviruses. Semin Cancer Biol. 2008;18(4):296–301. DOI: <u>10.1016/j.semcancer.2008.03.010</u>
- Al-Sabaawy HB, Mustafa ES, Rahawi AM, Jaber MT, Al-Hamdany EK, Al-Hmdany SM. Histopathological study of sheep lung roaming in dump zones. Iraqi J Vet Sci. 2022;36(I):151-60. DOI: 10.33899/ijvs.2022.135830.2527
- Griffiths D, Cousens C. Dissecting the pathogenesis of ovine pulmonary adenocarcinoma with RNA-Seq. Impact. 2017;(7):95-97. DOI: <u>10.21820/23987073.2017.7.95</u>
- Karakurt E, Beytut E, Dağ S, Nuhoğlu H, Yildiz A, Kurtbaş E. Immunohistochemical detection of TNF-α and IFN-γ expressions in the lungs of sheep with pulmonary adenocarcinomas. Acta Vet Eur. 2022;48(3). DOI: <u>10.5152/actavet.2022.21124</u>
- 31. Devi VR, Yadav EJ, Rao TS, Satheesh K, Suresh P, Manasa BB. Nucleotide sequencing and phylogenetic analysis using PCR amplicons of U3 gene of Jaagsiekte sheep retrovirus (JSRV) detected in natural cases of ovine pulmonary adenocarcinoma in India. Open J Vet Med. 2014;4(11):267. DOI: <u>10.4236/ojvm.2014.411032</u>
- 32. Singh R, Singh S, Singh R, Varshney R, Dhama K, Kumari S, Singh KP, Dar JA, Kashyap G, Kamdi B, Kumar P. Patho-Epidemiological study of Jaagsiekte sheep retrovirus infection in the sheep and goats population, India. Biol Rhythm Res. 2020;51(8):1182-96. DOI: 10.1080/09291016.2018.1559422
- Al-Mahmood SS, Khalil KW, Edreesi AR. Histopathology and immunohistochemistry of tumors in animals attending veterinary teaching hospital. Iraqi J Vet Sci. 2022;36(2):309-14. DOI: 10.33899/ijvs.2021.130114.1733
- Dutkowska A, Szmyd B, Kaszkowiak M, Domańska-Senderowska D, Pastuszak-Lewandoska D, Brzeziańska-Lasota E, Kordiak J, Antczak

A. Expression of inflammatory interleukins and selected miRNAs in non-small cell lung cancer. Sci Rep. 2021;11(1):5092. DOI: 10.1038/s41598-021-84408-1

- Karakurt E, Coşkun N, Beytut E, Keleş ÖF, Dağ S, Yılmaz V, Nuhoğlu H, Yıldız A, Kurtbaş E. Evaluation of the relationship between inflammatory reaction and interleukins in ovine pulmonary adenocarcinomas. Vet Res Forum. 2023;14(1):1–6. DOI: 10.30466/vrf.2022.542311.3279
- Al-Jameel WH, Al-Sabaawy HB, Abed FM, Al-Mahmood SS. Immunohistochemical expression of proliferation markers in canine osteosarcoma. Iraqi J Vet Sci. 2022;36(4):1097-1102. DOI: 10.33899/ijvs.2022.133138.2177

# الكشف الجزيئي والمرضي للورم الغداني الرئوي في الأغنام في مجازر مدينة الموصل

أسامة زياد جياد و أحمد محمد على السيدية

لطبيب بيطري، قطاع خاص، أفرع الأمراض وأمراض الدواجن، كلية الطب البيطري، جامعة الموصل، الموصل، العراق

#### الخلاصة

الورم الغداني الرئوي في الأغنام هو من الحالات المرضية المتكررة والتي تصيب المجترات الصغيرة والذي يتميز بالتهاب النسيج الرئوى مسبباً فرط تنسج للخلايا الطلائية المبطنة للحويصلات والهوائية والقصيبات الهوائية والمسبب هو فايروس الريترو. تم جمع عينات من الأغنام في مسالخ الموصل ومحلات الجزارة. يعدّ التشخيص تعتبر نتائج اختبار تفاعل البوليميراز المتسلسل أول تحديد جزيئي لُفيروس الريترو في مدينة الموصل. اظهر الفحص العياني وجود عقيدات ورمية رمادية اللون منتشرة على أسطح فصوص الرئة وكان للنسيج الورمي حدود واضحة ومتميزة عن الرئة الطبيعية المجاورة. نسيجياً لوحظ أن مناطق الورم العقيدي (العقيدات الرئوية التكاثرية) تتكون من خلايا سرطانية التي كانت تعانى من حؤول عمودي ومكعبى مميز بالإضافة إلى تضخم الخلايا الرئوية وخاصة الخلايا الرئوية (النوع الثاني)، والخلايا الظهارية (أنواع إفرازية) في الحويصلات الرئوية وخلايا كلارا غير الهدبية وكذلك البطانة الظهارية للقصيبات الهوائية. أخيرًا سلطت الدراسة الحالية الضوء على أهمية مرض الورم الغداني الرئوي في الأغنام في مدينة الموصل ومن خلال التشخيص الجزيئي للعامل المسبب للمرض عن طريق تفاعل البوليميراز المتسلسل، وكذلك دراسة التغيرات المرضية العيانية والنسيجية.