DIVA MOLECULAR DETECTION OF SUSPECTED CASE OF NEWCASTLE AND ENCEPHALOMYLITIS DISEASE IN LAYERS

Amjed. H. Ulaiwi Assist.Prof.

Coll. Vet. Med., University of Baghdad E-mail: amjed.h@covm.uobaghdad.edu.iq

ABSTRACT

The aim of the study investigates the molecular diagnosis between (the Avian encephalomyelitis virus and Newcastle disease virus and differentiating the infected from vaccinated animals (DIVA) strategies. The 1st detection by RT-PCR for(PMV-1) as primary detection then, the samples were prepared as in FTA card Whatman[®] and sent to (AniCon Labs) Germany to detect by one step RT-PCR (DIVA technique) between NDV and AEV genotype. The result of the molecular investigation as primary detection revealed three samples were positive with Ct values (34.0, 26.6, and 35.8) respectively. Then, in the detection of AEV the result showed all samples were negative. finally, by using (DIVA) with two primers to detect general (PMV1-9) by (M-gene) all samples were positive at Ct values (20.8, 20.1, and 25.2) respectively, and then path type by (F gene) to differentiate infected from vaccinated samples also all samples were positive with Ct values (23.8, 23.3 and 23.1) respectively. the study concluded that the samples can differentiate between AEV and NDV by using DIVA strategies to find pathotypes and differentiating between infected and vaccinal isolate based on certain amino acids in the primers.

Key words: M gene, F gene, one-step RT-PCR, meso-velogenic ND.

مجلة المعلوم الزراعية المعراقية -892: 390:(3):42 عليوي عليوي

الكشف الجزيئي بتقنية DIVA عن الحالات المشتبه فيها لمرض نيوكاسل والتهاب الدماغ والنخاع في الدجاج البياض أمجد حسين عليوي

. أستاذ مساعد

كلية الطب البيطري - جامعة بغداد

المستخلص

تهدف الدراسة إلى التحقق عن طريق التشخيص الجزيئي بين مرضي التهاب الدماغ والنخاع الطيري الفايروسي للقاروسي للتفريق بين العزلات اللقاحية والممرضة (DIVA). اجري التشخيص الاولي بواسطة RT-PCR نيوكاسل NDV الفايروس نيوكاسل النوع الأول (PMV-1)، ثم تم تحضير العينات كما هو الحال في بطاقات PTA Whatman® Card لفايروس نيوكاسل النوع الأول (AniCon Labs)، ثم تم تحضير العينات كما هو الحال في بطاقات OIVA) المانيا، من خلال استخدام تقنية NDV و NDV المانيا، من خلال استخدام تقنية الميني، كانت المنابع عن ثلاث عينات فايروسات NDV و NDV كانت (34 و 26, 6و 38, 8) على التوالي. بعد ذلك، من خلال الكشف عن AEV كانت إيجابية بواسطة مع قيم Ct كانت سلبية. باستخدام تقنية (DIVA) مع اثنين من البادئات للكشف العام لفيروس مرض أظهرت النتائج أن جميع العينات كانت سلبية. باستخدام تقنية (DIVA) مع اثنين من البادئات للكشف العام لفيروس مرض نيوكاسل ولجميع الأنواع المصلية (PMV1-9) للجين كالتمييز بين العينات المصيبة واللقاحية وكانت النتائج 20) على التوالي، كما استخدم النمط الوراثي بواسطة جين F للتمييز بين العينات المصيبة واللقاحية وكانت النتائج NDV و 20، استخدام استراتيجيات 8 و 25, 3 و 20, 1 و 32, 3 و 100، استخدام استراتيجيات الكان المصيبة واللقاحية على بعض الأمينية في البادئات.

الكلمات المفتاحية: جين M، جين F، خطوة وإحدة لتفاعل البوليمر المتسلسل، عترة متوسطة -ضارى لنيوكاسل

Received:13/4/2022, Accepted:14/8/2022

INTRODUCTION

(ND) Newcastle disease and Avian encephalomyelitis disease (AE) revealed similar signs as well as with other nervous diseases in poultry clinically (18). In poultry, production (ND) is a serious viral disease with major economic losses and worldwide distribution (2,6). The isolates of (PMV) are categorized into nine serotypes (APMV-1 to APMV-9), especially (APMV-1) (31). The molecular detection of Newcastle disease virus (NDV) depends on six main proteins especially matrix protein (M) for general detection as (APMV) and (F protein) for velogenic NDV, with major RNA genome negative-sense, and single-stranded nonsegmented (27,28,29). Experimentally, the virulence of NDV is classified according to pathogenicity: into (velogenic, mesogenic, lentogenic, and asymptomatic) forms, the velogenic (vvNDV) **NDV** produce neurological signs with hemorrhagic lesions and/ or respiratory signs (12). While(AE) is one of the important viral diseases many avian young species with an economic impact on poultry production (3,7,30). The disease has neurological signs especially, tremors and transmitted ataxia is vertically and horizontally, and also caused 40-60% morbidity and a 25% mortality rate (21,22). The AE virus belongs to the (family Picornaviridae, genus *Tremovirus*) single-stranded RNA with a small genome positive sense (19,20). Molecular classification of virus contains four structural proteins (VP-1, VP-2, VP-3, and VP4) with seven nonstructural proteins (15). Targeting of AEV detection by an (RT-PCR) method using primers depends mainly on the (VP2 gene) (32). DIVA strategies are the goal for differentiation between the diseases and eradication (23).

MATERIALS AND METHODS

Sampling: Brown table egg layer (70 days old), the number (50 000) chickens, the flock exhibits a sudden onset of nervous manifestation with mortality rate elevated reach to 5% of the flock, the preparation of samples as follows.

1-Six samples (Three Brain and three Trachea) detected in Iraq RT-PCR for(PMV-1)

- **2-** the samples were prepared as in FTA card Whatman[®] FTA Cards for isolation, purification, and storage of genetics material for diagnostic and research applications
- **3-** The samples were sent to (AniCon Labs) in Germany sponsored by (Boehringer Ingelheim) a financier and supporter of scientific research, to detect by one step RT-qPCR (DIVA technique) the presence of NDV and AEV and to genotype the existing virus.

Primers: Avian Encephalomyelitis Virus (**AEV**) :Method: Species-specific RT-PCR (Kylt[®] Avian Encephalomyelitis) (16).

Parameter: General NDV (aPMV-1) (**M-Gene**) Name of the oligonucleotide: PROBE: HEX APMV-1 (HEX / BHQ) Sequence (5, to 3,): GGGACRGCHTGCTATCC (Method: Serotype-specific RT-PCR) (Kylt® Paramyxovirus 1).

NDV type 1 (aPMV-1) -1- Parameter: DIVA - detection of velogenic strains (Fprotein) Name of the oligonucleotide: PRIMER: APMV-1 F Sequence (5, to 3,): AGTGATGTGCTCGGACCTTC (F- Gene) oligonucleotide: PRIMER: Name of the (APMV-1 R Sequence 5, to 3,): CCTGAGGAGAGGCATTTGCTA Method: Pathotype-specific Real-Time RT-PCR (Kylt[®] aPMV-1 pathotype). The protocol of primers occurs according to GoTaq® 1-Step RT-PCR System: Based on certain amino acids in the matrix protein gene as general Paramyxovirus-1 (PMV-1). (F protein) the sequenced strain could be categorized to the velogenic (highly virulent) PMV-1 strains, meaning the strain carries amino acids typical for velogenic strains. The PMV-1 was classified on the pathogenicity site especially (F2/F1 cleavage). The (F protein) was based on (360 bp) represented (APMV-1).

RESULTS AND DISCUSSION

The result of clinical signs and Post mortem findings showed the flock exhibits a sudden onset of nervous manifestation (head trimming, star gazing, incoordination, imbalance, and unilateral paralysis) with fever, mortality rate elevated reach to (5%) of the flock. The Post mortem changes include some with petechial congested carcass, hemorrhages, congested liver, pale spleen, congested viscera and proventriculus, necrotizing and engorged payer's patches, transparent intestine, with bile stained content and engorged cecal tonsils. The result of clinical signs and post-mortem findings agree with kommers et. al. showed the neurological NDV strains that were isolated in the flocks accompanied by nervous signs and visceral changed paralysis of wings or legs was (unilateral or bilateral), head or muscular tremors, torticollis with congestion of body organs and mortality rate reach to 50%. Also, the mesogenic strain of PMV-1 caused low mortality in chickens at age 4-week-old with nervous signs such as paralysis, torticollis and tremors of head (1). The post-mortem lesion (4) the outbreak of 2-week-old chickens with observed enlargement APMV-3 and congestion of the spleen and liver with focal necrosis in the pancreas. Also, the infection with APMV-5 showed splenomegaly with discoloration of the liver and hemorrhages in the proventriculus and small intestine (10). As well as the infection in doves with APMV-7 observed congestion and enlargement in the spleen and liver (11). Also, these signs and lesions could cause by Avian encephalomyelitis (epidemic tremor) AEV affecting the nervous system with the same rate of morbidity and mortality (14). The similarity between AEV and APMV in neurological symptoms as uni or bilateral paralysis, ataxia, and head or neck tremors (16,17). The molecular results revealed the 1st detection of PMV-1 in Iraq in six samples (3 Brain and 3 Trachea) revealed three samples were positive (2 Brain and. Trachea) with Ct values of (34.0, 26.6 and 35.8) respectively.

Table 1. Samples detected of APMV-1 by RT-PCR

NDV Site ID	Sample ID	Channel Result	Ct	EndPt
A9	Trachea 1	NEG	0	9
A10	Brain 1	NEG	0	6
A11	Trachea 2	POS	34.0	19
A12	Brain 2	POS	26.6	37
A13	Trachea 3	NEG	0	9
A14	Brain 3	POS	35.8	16

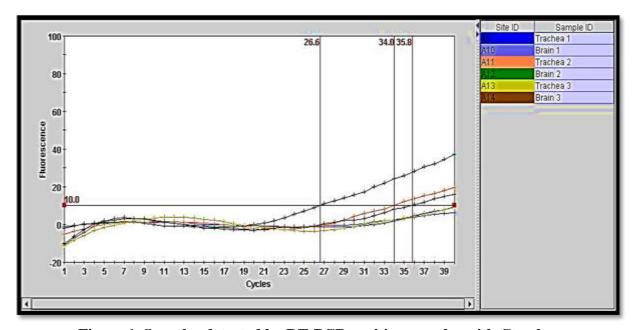


Figure 1. Samples detected by RT-PCR positive samples with Ct values

The molecular identification of PMV-1 was the best and most accurate method that agrees with sameera *et. al.* molecular detection of NDV was the sensitive method by using RT-PCR. Also, recently quicker and more sensitive (RT-PCR) for detection and genotyping of PMV-1 (24,25). As well as the

using PCR assays specific for mesogenic/velogenic PMV-1 with Ct values (23.4, 29.0, and 30.8) respectively in chicken had high analytical sensitivity (13). The results to AEV showed not detected by RT-PCR all samples were negative.

Table 2. Species-specific RT-PCR (Kylt® Avian Encephalomyelitis)

Sample No.	Sample	Result
	Description	
A1804251.001	FTA-card	not
	(spot 1)	detectable
A1804251.002	FTA-card	not
	(spot 2)	detectable
A1804251.003	FTA-card	not
	(spot 3)	detectable

Finally, the samples used one-step RT-PCR (DIVA) to differentiate infected from vaccinated animals. The result showed in 1st step of detection of general (PMV1-9) by

matrix protein gene (M-gene) all samples were positive at Ct values (20.8, 20.1, and 25.2) respectively.

Table 3. Samples detection for (NDV, aPMV) RT-PCR (Kylt® PMV)

Sample No.	Sample Description	CT	Result
A1804251.001	FTA-card (spot 1)	20,8	positive
A1804251.002	FTA-card (spot 2)	20,1	positive
A1804251.003	FTA-card (spot 3)	25,2	positive

Then, the result in 2nd step of detection pathotype of APMV-1 to differentiate infected (meso-velogenic strain) from vaccinated samples (lentogenic strain). The result

reveled all samples were infected (mesovelogenic) strain with Ct values (23.8, 23.3 and 23.1) respectively.

Table 4. Samples detection for serotype-1 (NDV, PMV-1) - DIVA - detection of velogenic strains, Pathotype-specific RT-PCR (Kylt® aPMV-1 pathotype).

Sample No.	Sample Description	CT lentogenic	CT velo-/meso genic	Result
A1804251.001	FTA-card (spot 1)	-	23,8	velogenic strain positive
A1804251.002	FTA-card (spot 2)	-	23,3	velogenic strain positive
A1804251.003	FTA-card (spot 2)	-	23,1	velogenic strain positive

The PMV-1 was detected on (360 bp) and for the pathogenicity site especially (F2/F1 cleavage). The (F protein) was based on represented (APMV-1). The results of one-step RT-PCR (DIVA) to differentiate infected (meso/velogenic) strains from vaccinated (lentogenic) strain for detection of general PMV and pathotype strains correspond to the following studies that showed the detection of NDV by (RT-PCR) assays is now extensively applied because this assay is less tired as well as faster and accurate through discovering important two proteins matrix protein (M), fusion protein (F) (8,9). Also The identity of the reference of general PMV can be detected by the rRT-PCR protocol by using specific primer and probes for the M gene (26). Then the molecule reaction by PCR specifically (DIVA) technique differentiating between lentogenic (vaccinal) and mesogenic/ velogenic (infected) the detection of approximately 10 and 20 copies of strains had high analytical sensitivity (13). As well as the Iraqi isolates of NDV can be diagnosed by using specific primers designed through using the hemagglutinin-neuraminidase (HN) gene based on discovering the site of pathogenicity by using one-step RT-PCR (5).

CONCLUSION

The study was concluding that the samples were isolated and detected by RT-PCR through 1st detection of general PMV by using the M gene and using DIVA strategies to find pathotypes and differentiating between infected and vaccinal isolate based on certain amino acids in the (F protein) the sequenced strain could be categorized as the velogenic (highly virulent) PMV-1 strains, meaning the strain carries amino acids typical for velogenic strains.

REFERENCES

- 1. Abolnik, C. 2017. History of Newcastle disease in South Africa. Onderstepoort J. Vet. Res. 84(4):1–7.
- 2. Ahmed, A.I. and S.M. Odisho.2018. Isolation identification and pathotyping of newcastle disease viruses from naturally infected chickens in Iraqi Kurdistan region. Iraqi J. Agri. Sci. 49(1):132-14. DOI: https://doi.org/10.36103/ijas.v49i1.216
- 3. Ali, Z., W. S. Taohid, M. M. Mohammad, B. Akramul, A. Al Momen, A.K. Shamsul, A.B. Zafar and M. Giasuddin. 2021. First report on the seroprevalence of avian encephalomyelitis virus antibody in Sonali (cross-bred) chickens in Bogura, Bangl.Vet. 8(1): 78–83.
- 4. Alphin, R.L., C.D. Ciaverelli, D.P. Hougentogler, K.J. Johnson, M.K. Rankin and E.R. Benson. 2010. Post outbreak disinfection of mobile equipment. Avian Dis. 54(1):772–776
- 5. Al-Shammari, A.A., A. Huda and A. M. Murtadha.2014. Molecular diagnosis of Newcastle disease Iraqi Virulent strain virus HN gene by specific primers design.2014. Kufa J. Vet. Med. Sci.5(2):196-203
- 6. Ayoub, M.A., W. K. Elfeil, D. El Boraey, H. Hammam and M. A. Nossair. 2019. Evaluation of some vaccination programs in protection of experimentally challenged broiler chicken against newcastle disease virus. Am. J. Anim. Vet. Sci. 14(3): 197-206
- 7. Barros, M.E., B. P. Rocha, F.A. Souza, F. S. Mendonça and J. Evencio-Neto. 2019. Severe outbreak of avian encephalomyelitis in laying hens in northeastern brazil. Braz J Poult. Sci. 21 (02):1–4.
- 8. Beenish, Z., I. Q. Javed, Z. Amir, A. Asim, A. Raheela, S. Haleema, A. Qurat. S. Razia, I. Irfan and A. Sobia. 2020. Detection and molecular characterization of virulent Newcastle disease virus in ducks (*Anas platyrhynchos domesticus*). Pakistan J. Zool.52(1):1-4.
- 9. Chen, H.T., J. Zhang, D. H. Sun, J. L. Zhang, X. P. Cai, X. T. Liu, Y. Z. Ding, L. N. Ma, S. H. Yang, L. Jin and Y. Liu. 2019. Development and application of multiplex PCR method for simultaneous detection of seven viruses in ducksVet. Res. Commun., 15(103): 1-10.

- 10. Chen, J.P. and C.H. Wang. 2002. Clinical epidemiologic and experimental evidence for the transmission of Newcastle disease virus through eggs. Avian Dis. 46(2):461–465
- 11. Cucco, M., I. Pellegrino and G. Malacarne. 2010. Immune challenge affects female condition and egg size in the grey partridge. J. Exp. Zool. A. Ecol. Genet. Physiol. 313(9):597–604.
- 12. Dimitrov, K.M., C. Abolnik, C. L. Afonso, E. Albina, J. Bahl, M. Berg, F.X. Briand, I. H. Brown, K. S. Choi and I. Chvala. 2019. Updated unified phylogenetic classification system and revised nomenclature for Newcastle disease virus. Infect. Genet. Evol. 74 (103917): 1-15.
- 13. Farkas, T., E. Sze'kely, S. Bela'k and I. Kiss. 2009. Real-Time PCR-based pathotyping of newcastle disease virus by use of TaqMan minor groove binder probes. 2009. J. clini. microbiol. 47(7): 2114–2123
- 14. Fatima, K.E, Y. A. Almofti, K. A. Abdelrahman, M. A. Nouri and E. A. Eltilib. 2021. Structural analysis of avian encephalomyelitis virus polyprotein for development of multi epitopes vaccine using immunoinformatics approach. J. Pure. Appl. Microbiol.15(1):262-278
- 15. Freitas, E. S. and A. Back. 2015. New occurance of avian encephalomyelitis in broiler is this an emerging disease. Braz. J. Poult. Sci.17(3):399–404
- 16. Hameed, S.S., Amjed H.U and Hamad S.M.2022. Diagnosis of E. coli isolated from arthritis in chicken by vitk and molecular methods. Iraqi J. Agri. Sci.53 (1),141-146. DOI: https://doi.org/10.36103/ijas.v53i1.1518
 17. Hauck, R., C. G. Senties-Cue and Y. Wan. 2017. Evolution of avian encephalomyelitis virus during embryoadaptation. Vet. Microbiol. 204(12):1-7.
- 18. Hesham, A., K. Wael, A. Ahmed, T. Laila, G. Elsayed, M. Emad, A. Ahmad and T. Shaimaa. 2022. Efficacy of the newcastle disease virus genotype VII.1.1-matched vaccines in commercial broilers. Vaccines.10 (1): 1-17
- 19. Hussein, S. I., A.F. Khalaf, Y. Ahmed, B. Ahmed and A. Iyad. 2020. Determination of inhibition activity of α -amylase enzyme, antioxidant activity, antibacterial activity and

phenolic compounds by using some medical plants. Iraqi J. Agri. Sci. 51(1):411-421.

DOI: https://doi.org/10.36103/ijas.v51i1.940

- 20. Knowles, N. J., T. Hovi, T. Hyypia, A. M. King and L. M. Lindberg. 2012. Family picornaviridae. In: King, A. M. Q., M. J. Adams, E. B. Carstens and E. J. Lefkowitz. editors. Virus taxonomy: classification and nomenclature of viruses: nineth report of the international committee on taxonomy of viruses. London: Elsevier. J. Gen. Virol.94(9): 2029–2035
- 21. Kommers, G.D., D.J. King, B.S. Seal and C.C. Brown. 2001. Virulence of pigeon-origin Newcastle disease virus isolates for domestic chickens. Avian Dis. 45(4):906–921
- 22. Lau, S. K. P., P. C. Woo, C.C. Yip, K. S. Li, R. Y. Fan, R. Bai, Y. Huang, K. H. Chan and K. Y. Yuen. 2014. Chickens host diverse picornaviruses originated from potential interspecies transmission with recombination. J. Gen. Virol. 95(9):1929-1944
- 23. Magdalena, M., H. M. Bernd, G. Christian, R. O. Angela and C. M.Thomas. 2020. A novel recombinant newcastle disease virus vectored DIVA vaccine against peste des petits ruminants in goats. Vaccines.8(1): 205-211.
- 24. Mohammad, A.H., and A.S. Al-Hassani.2022. Effect of different levels of turmeric root powder to diet to some traits of broiler exposed to heat stress. Iraqi J. Agri. Sci. 53:(4), 950-957.
- DOI:https://doi.org/10.3610 3/ ijas.v53i4.1607 25. Qiu, X., Y. Yu, S. Yu, Y. Zhan, N. Wei, C. Song, Y. Sun, L. Tan and C. Ding. 2014. Development of strand-specific Real-Time RT-PCR to distinguish viral RNAs during Newcastle disease virus infection. Sci. World J.Article ID 934851 01
- 26. Rahman, M.M., L. R. Barman, E. H. Chowdhury and M. R. Islam.2016. Detection of Newcastle disease virus of poultry by real

- time reverse transcription polymerase chain reaction. Bangl. Vet.33(1):16 22
- 27. Romer-Oberdorfer, A., O. Werner, J. Veits, T. Mebatsion and T. C. Mettenleiter. 2003. Contribution of the length of the HN protein and the sequence of the F protein cleavage site to Newcastle disease virus pathogenicity. J. Gen. Virol. 84 (11):3121-3129
- 28. Sameera, A., A.M. Muhammad, M. Khushi, Y. T. Muhammad, R. Masood, R. A. ul-Rahman and Z. S. Muhammad.2016. Genetic characterization and phylogeny of pigeon paramyxovirus isolate (PPMV-1) from Pakistan. Springer Plus. 5(1): 1295-1312.
- 29. Shihab, I.M.2017. Effect of different levels of turmeric supplementation with diet on humoral immune response to newcastle and infectious bursal disease virus and histopathological changes to some internal organs of broiler chickens. Iraqi J. Agri. Sci. 48: (Special Issue): 041-044.

DOI:https://doi.org/10.3610/ijas.v48iSpecial.256.

- 30. Suarez, D. L. 2020. Newcastle Disease, Other Avian Paramyxoviruses, and Avian Metapneumovirus Infections. pp. 111–166. In: Diseases of Poultry, 14th ed. (Swayne, D. E., J. R. Glisson, L. R. McDougald, L.K. Nolan, D. L. Suarez and V. Nair. Wiley-Blackwell, Ames
- 31. Sultan, H.A. S. Talaat, W. K. Elfeil, K. Selim, M. A. Kutkat, S.A., Amer and K. S. Choi. 2020. Protective efficacy of the Newcastle disease virus genotype VII matched vaccine in commercial layers. Poult. Sci. 99(3):1275-1286
- 32. Xie, Z., M. I. Khan, T. Girshick and Z. Xie. 2005. Reverse transcriptase-polymerase chain reaction to detect avian encephalomyelitis virus. Avian Dis. 49(2):227-30.