

The inhibitory effect of the aqueous extraction of *Calotropis procera* against some pathogenic fungi

Saba Hassan Alwan¹ and Hussein Ali Salim²

Diyala Education Directorate, Ministry of Education, Iraq¹

Directorate of Diyala Agriculture, Ministry of Agriculture, Iraq²

E-mail: sab1071981a@gmail.com¹

E-mail: h_salim11111@yahoo.com²

Abstract

A laboratory experiment was conducted in the Diyala Agriculture Directorate's plant protection laboratory in November 2018 to investigate the effects of an aqueous extract of the Ashar plant, *Calotropis procera*, at three different concentrations (10, 20, and 30%) against the pathogenic fungi (*Rhizoctonia solani*, *Aspergillus ochraceus*, and *Penicillium* sp.) in vitro. Findings from the tests showed that the aqueous extract of *C. procera* had the highest percentage of inhibition against *R. solani* (86.67%). On the other hand, *Penicillium* sp. and *A. ochraceus* had lower percentages of inhibition, at 27.59 and 13.61%, respectively, at the same concentration (30%). The concentration (20%) reached 59.11, 26.44, and 9.68%, respectively, while the concentration (10%) reached 35.11, 24.37, and 6.82%, respectively.

Keywords: *Calotropis procera*, *Rhizoctonia solani*, *Aspergillus ochraceus*, *Penicillium* sp

Introduction

Plant diseases cause significant damage during plant growth and development, leading to a reduction in yield and quality of produce [25]. *Rhizoctonia solani* is an indigenous pathogen found in soil that affects numerous plant families and attacks plants at all phases of growth, resulting in damping off and seed rot either before or after germination [11,29]. One of the most common food-contaminating fungi in nature, *Aspergillus ochraceus*, is a filamentous fungus that is known to produce mycotoxins such as ochratoxin and citrinin [9]. One of the prevalent fungi found in a variety of environments, including soil, air, and vegetation, is *penicillium*. It produces mycotoxins [7] and phytopathogens that cause crop rot [20,26]. It is common practice to inhibit phytopathogenic fungi by using synthetic fungicides, and in order to minimise environmental pollution and damage to plants

caused by the synthetic fungicides, it is therefore important to reduce the use of synthetic chemicals. As a result, eco-friendly methods must be used to improve crop health and yield, and the most effective replacement for these hazardous chemicals is botanical extracts [24,22]. *Calotropis procera* is a soft, woody dendritic plant that can grow up to 6 meters tall. It has broad, oval leaves that are green in color and covered in white cotton bristles. Inside, it has purple flowers, hollow green fruits, and brown seeds with silky hair on top [28]. The plant belongs to the class of Dicotyledons, the order Gentianales, the family Asclepiadaceae, and the genus *Calotropis* [15]. The plant, also known as the huge milkweed, glazes, or apple of Sodom, Burmukh, and Karnaka, is also known as Ashar or Ashr in Arabic. It contains glycosides as well as quasi-alkaloids such as

calotropin, calactin, clotropagin, and gigantol, which are considered toxic to living organisms [4]. This plant was used as a treatment in popular medicine, but it is toxic by scientific standards, useful by industrial standards, and it can be used medically by treating it with a chemical or enzyme system to become a therapeutic substance [3]. This study was conducted to test the effectiveness of the aqueous extract of the Ashar plant *Calotropis procera* against the growth of *Aspergillus ochraceus*, *Penicillium* sp., and *Rhizoctonia solani* in vitro.

Materials and Methods

A lab experiment was conducted at the Plant Protection Laboratory of the Directorate of Diyala Agriculture in 2018. *Calotropis procera* leaves were collected from a single tree in Baqubah district, Diyala province. The fungi (*Aspergillus ochraceus*, *Rhizoctonia solani*, and *Penicillium* sp.) were obtained from the College of Agriculture, University of Diyala.

Preparation of the aqueous extract of *Calotropis procera* leaves

After washing the *Calotropis procera* leaves with water to get rid of dust and suspended impurities, the process involved weighing 20 g of leaves, adding them to 100 ml of sterile distilled water in a beaker for 24 hours, filtering them through a double-layered muslin cloth, and storing them in the refrigerator until use [10].

Poisoned food test technique

The potato dextrose agar medium (PDA) used in this process was prepared according to [5]. Extract concentrations (10, 20, and 30%) were prepared by adding 10 ml of extract to 90 ml of PDA to get 10%, 20 ml of extract to 80 ml of PDA to get 20%, and 30 ml of extract to 70 ml of PDA to get 30%. The control treatment included only PDA medium. After the medium became solid, agar discs (6 mm) of *A.*

ochraceus, *R. solani*, and *Penicillium* sp. from the seven-day-old culture were transferred to petri plates and incubated at 25 ± 2 °C for 7 days, according to the poison food technique [18,16]. The tests were performed in three replications, and the diameter of fungus growth was calculated. The percent inhibition of fungal growth was calculated according to [19].

$$\text{Inhibition (\%)} = (1 - T/C) \times 100$$

Where, C = Colony growth of control, T = Colony growth of treatment

Statistical analysis

The data were analyzed by one-way analysis of variance (ANOVA) by using the completely randomized design (CRD) [6].

Results and discussion

The results in Table 1 and Figure 1 showed that *Calotropis procera* concentrations led to a significant increase in the inhibition percentage of *Rhizoctonia solani*, *Aspergillus ochraceus*, and *Penicillium* sp., where a concentration of 30% was significantly superior in the inhibition percentage of mycelial growth, which reached (86.67, 13.61, and 27.59%), followed by concentrations of 20% (59.11, 9.68, and 26.44%) and 10% (35.11, 6.82, and 24.37%), respectively, compared to control (0.0%).

The aqueous and alcoholic extracts from the Ashar plant are highly effective and inhibit the growth of fungal and bacterial microbes. Their effectiveness may be attributed to the fact that they contain calotropain protein [13]. In a previous study [2], *Aspergillus* conidia were treated with different concentrations of *Calotropis procera* extract (5, 10, and 20%), which led to a significant decrease in the percentage of conidia survival by increasing the extract concentration. [8] stated that the tested concentrations of Ashar leaves were effective in inhibiting the growth of the fungi

Alternaria sp. and *Fusarium* sp. [17] reported that the alcoholic aqueous extracts of Ashar flowers inhibited the growth of the fungi *Sclerotinia* sp. and *Geotrichum candidum*, as well as three genera of bacterial plant pathogens, *Erwini* sp., *Xanthomonas* sp., and *Pseudomonas* sp. This clearly shows the effectiveness of the *Calotropis procera* extract as a strong anti-microbial because it contains the protein calotropain [3]. This is consistent with the study [12], in which *Calotropis procera* leaf extract inhibited the germination of the stony bodies of *Macrophomina phaseolina*, *Helminthosporium* sp., and *Alternaria radicina*. [27] stated that the extract of *Calotropis procera* inhibited the growth of the following fungi, *Fusarium oxysporum*, *Helminthosporium spiciferum*, *Aspergillus flavus*, and *Curvularia lunata*. This is because the aqueous extract of *Calotropis procera* kills microbes and stops the growth of the fungus *Penicillium* sp. It also has a lot of terpenes and

alkaloids that have complex biological effects [14]. The variation in the inhibitory activity of the extract on the growth of fungi is due to the nature of the contrast in the active substances and components and their quantities, which affect the inhibitory ability and the nature of the fungus [1]. This is consistent with [21], where it was found that some plant extracts had stimulating effects for some fungi, some of them had an inhibitory effect, while others had no effect. Among the plant extracts that inhibited these fungi (*Rhizoctonia solani*, *Aspergillus ochraceus*, and *Penicillium* sp.) are fenugreek seed extracts [23].

Conclusion

Calotropis procera extract can inhibit a lot of different types of fungi, including *Rhizoctonia solani*, *Aspergillus ochraceus*, and *Penicillium* sp.; whenever the extract concentration increased, it led to an increase in the rate of fungal inhibition

Table 1. Effect of different concentrations of *Calotropis procera* extract on inhibition percentages of *Rhizoctonia solani*, *Aspergillus ochraceus*, and *Penicillium* sp. in vitro

<i>Calotropis procera</i> concentrations	<i>Rhizoctonia solani</i>	<i>Aspergillus ochraceus</i>	<i>Penicillium</i> sp.
Control	0.00	0.00	0.00 d
10%	35.11	6.82	24.37 c
20%	59.11	9.68	26.44 b
30%	86.67	13.61	27.59 a
CD (0.05)	0.01	0.01	0.01

CD = Critical difference



Figure 1. Effect of different concentrations of *Calotropis procera* extract on inhibition percentages of *Rhizoctonia solani*, *Aspergillus ochraceus*, and *Penicillium* sp. in vitro

References

- [1] Al- Rawi, H. and HL Chakravary . 1988. Medicinal plants of Iraq. 2nd edition. Ministry of Agriculture and Irrigation, State Board for Agricultural and Water Resources Research, National Herbarium of Iraq.
- [2] Ali, Hani Al-Siddiq Hamza Muhammad. 2005. Evaluation of the genetic toxicity of the dairy material of the Ashar plant by using the nutritional deficiency test in the fungus *Aspergillus* Turis / King Abdul Aziz University Teachers College, Jeddah Governorate .
- [3] Amer, Wafa Mahrous. 2001. Al-Ashar plant a disease and a medicine. Assiut Journal of Environmental Studies - Twentieth Issue:93-99 .
- [4] Arish, Fatima Suleiman. 2009. Evaluation of the genetic toxicity of polar gradient extracts of the leaves and fruits of the brumpet

plant on the mitotic division of the cells of the growing apices of the roots of the onion plant / Master Thesis / Sebha University, Faculty of Science .

[5]Collee, JG., AG, Fraser, BP, Marmion.and, A., Simmons. (1996). Practical Medical Microbiology. Mackie and Macarthey pearson professional limited. 14th ed

[6]Fisher RA, Yates, 1968 Statistical method for research workers. Oliver and boyd ltd. Edinburgh and London, 10. <http://www.ccari.res.in/wasp/index.php>.

[7]Frisvad JC, Smedsgaard J and Larsen T O 2004. Mycotoxins, drugs and other extrolites produced by species in *Penicillium* subgenus. *Studies in Mycology* 49: 201-241.

[8]Ganie, SA Pant, VR Ghani, MY Lone, AH Anjum, Q. and Razvi, SM 2013. In vitro evaluation of plant extracts against *Alternaria brassicae* (Berk .) Sacc. Causing leaf spot of mustard and *Fusarium oxysporum* f. sp. *lycopersici* causing wilt of tomato. *Scientific Research and Essays*, 8 (37): 1808-1811.

[9]Ghibaudo G and Peano A 2010. Chronic monolateral otomycosis in a dog caused by *Aspergillus ochraceus*. *Veterinary Dermatology* 21 (5): 522-6. Retrieved from doi: 10.1111/j.1365- 3164.2010.00884.x.

[10]Harborne, J. B. (1984). phytochemical methods. A. Guide to modern techniques of plants, analysis London. New York, chapman & Hall. ed. nd 2

[11]Jabr, KS, Farhan, DA and Rashid AH 2008. Evaluating the efficacy of some biological control agents and Beltanol against fungi *Rhizoctonia solani*, *Fusarium oxysporum* which causes seed rot and damping off of watermelon. *Iraqi Agricultural Science Journal* 39(2): 78-68.

[12]Jalal, AO and Ghaffar, A. 1992. Antifungal properties of *Ocimum sanctum* L. National Symposium on the Status of Plant

Pathology in Pakistan. Univ. Of Karachi., pp. 283-287.

[13]Kareem, So. Akpan,I. and Ojo,Op. 2008. Antimicrobial Activities *Calotropis procera* on Selected Pathogenic Microorganisms African Journal of Biomedical Research, 11: 105--110.

[14]Kori, P. and Alawa, P. 2014. Antimicrobial activity and phytochemical analysis of *Calotropis gigantea* root, latex extracts. *IOSR Journal of Pharmacy*, 4 (6): 07-11.

[15]Kumar, P.S.; E. Suresh and S. Kalavathy.2013. Review on a potential herb *Calotropis gigantea* (L.) R.BR. *Scholars Academic Journal of Pharmacy* .2(2):135-143 .

[16]Misra,S . B. and SN., Dixit. (1976) fungicidal spectrum of the leaf extract of *Allium sativum*. *India phytopathology*, 29: 448-449

[17]Muhammad, Nawara Ali, Najia Muhammad Jadallah, Ghazaleh Ibrahim Fadil and Asma Muhammad al-Mabrouk .2018. The effect of ginger and ten extracts on the growth of some plant pathogens. *The Libyan Journal of Plant Protection*, Issue No) .8)):97-107 .

[18]Nene VL and Thaplyal PN 1987. Fungicides in Plant Disease Control. India: Oxford & IBH Publ. Co. Pvt. Limited 507 p.

[19]Pautiz , TC, T., Zhou and L., Rankin. 1991. Selection of Rhizosphere bacteria for biological control of *Pythium aphanidermatum* on hydroponically grown cucumber. *Biological control*, 2: 222-23

[20]Pitt JI and Hocking AD 2009. Fungi and Food Spoilage. Springer, New York.

[21]Qasem, JR and HA Abu- Blan. 2008. Fungicidal activity of some common weed extracts against different plant pathogenic fungi. *Journal of Phytopathology*, 144 (3): 157-161.

[22]Salim H, Alsaady M, Al-zuhairi A, Kassoub F. 2023. Evaluation of the fungicidal

activity of the aqueous extracts of some medicinal plants against *Fusarium* spp. *Revis Bionatura*; 8 (2) 84. <http://dx.doi.org/10.21931/RB/2023.08.02.84>.

[23]Salim H. A., Jabar A. K., Alwan S. H., Abdulrazaq S. M., Hussein H. H., Gasam H. S., Zedan D. A. and Rosoki B. O. 2021. Efficacy of Fenugreek Seeds Extract (*Trigonella foenumgraecum*) against *Penicillium* sp, *Rhizoctonia solani* and *Aspergillus ochraceus* in vitro, *Indian Journal of Ecology* (2021) 48 Special Issue (15): 165-168.

[24]Salim H.A., Salman I.S., Jasim B.N., 2016. IPM Approach for the management of wilt disease caused by *Fusarium oxysporum* f. sp. *lycopersici* on tomato (*Lycopersicon esculentum*). *Journal of Experimental Biology and Agricultural Sciences* 4: 742–747.

[25]Salim H.A., Simon S. and Lal A.A. 2017. Integrated diseases management (IDM)

against tomato (*Lycopersicon esculentum* L.) *Fusarium* wilt. *J Environ Agric Sci* 11: 29-34 .

[26]Samson RA, Houbraken J and Thrane U 2010. Food and indoor fungi. CBS KNAW Biodiversity Center, Utrecht.

[27]Singh, HNP Prasad, MM and Sinha, KK 1993. Efficacy of leaf extracts of some medicinal plants against disease development in banana. *Letters in Applied Microbiology*, 17 (6): 269-271.

[28]Suleiman, M. H.1995. A series of studies in Flora of the Kingdom of Saudi Arabia, a study of vegetation on the road to Mecca up to Kilo14 In Jeddah. Page 10. Umm Al Qura University .

[29]Thornton RC, Gronhof CA, Forrest R and Lamotte R 2004. A one step immune chromatographic lateral flow device specific to *Rhizoctonia solani* and certain related species and its use to detect and quantify *R. solani* in soil. *Phytopathology* 94: 280- 288.