Impact of seed soaking by Salicylic Acid, Biozyme application, and covering on Okra (Abelmoschus esculentus L.) germination and yield.

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Abstract:

The study carried out during (April to October 2020) at two different locations in Sulaimani governorate, the first location was the Agricultural nursery in Tasluja (back to nature), and the second at the vegetable farmer in Dukan, the study determines the response of an okra variety and the effect of salicylic acid on the growth and germination of seeds and foliar application with biozyme as well as the use of temporary vegetation covering on growth, yield and seed quality. Then the two locations were compared in quantitative and qualitative appearances of the local okra production, and their interactions on yield and its growth characteristics and the quantitative and qualitative of the local okra crop. This study was designed as a factorial according to a randomized complete block design (R.C.B.D.) with three replicates and (54 experimental units). An experiment was conducted to find out the effect of the vital properties of local okra seeds obtained directly in Sulaymaniyah during the season from (2020). The seeds were soaked for 4 hours in three different concentrations (0 ppm, 75 ppm and 150 ppm)) of salicylic acid with distilled water to achieve a higher germination, Biozyme foliar application at three levels (0 ml.l⁻¹, 0.75 ml.l⁻¹ and 1.5 ml.l⁻¹), and covering on okra yield quality in green shade-net house. The results indicated that salicylic acid treatment led to significant effect on seed germination and yield.

The interaction among locations, covering, salicylic acid and biozyme foliar application, significantly (P<0.05) effected positively on pod weight (4.829 g), Yield Per Hectare (9.491 ton/ hectare), number of seeds per pod (78.467) and seed weight per pod (68.000 g). Keywords: Okra, salicylic acid, biozyme, covering, seed quality

Introduction:

Okra (Abelmoschus esculentus L. Moench.) is a vegetable crop that belongs to the Malvaceae family (1). Okra has a deep root system, smooth, striped seeds that range in color from dark green to dark brown, and is a valuable vegetable crop grown in tropical and subtropical regions around the world (2, 3, 4). Gumbo originated in Ethiopia (5), and by the 12th century, it had spread across North Africa, the Mediterranean, Arabia, and India (6). The majority of suitable land has been cultivated, and expanding into new areas to increase okra production as a staple food in most countries is both feasible and desirable. It's a common crop in India because it's easy to grow, and it's highly adaptable to different climates and humidity levels. Okra is extremely vulnerable to cold temperatures, resulting in thin plants with poor ability to produce leaves and flowers, as well as

irregular pods that are reduced in quantity and quality. According to (7) global food production would need to increase by 38 percent by 2025 and 57 percent by 2050. Because of its medicinal nature, dietary fibers, and distinct seed protein in both lysine and tryptophan amino acids, okra has been dubbed "the ideal villager's vegetable" in Africa accounts for more than 75% of okra produced , but the average productivity in region is very low (2.5 t/ha) compared to East (6.2 t/ha) and North Africa (8.8 t/ha) (8).

Germination is a crucial phase in the growth of plants because it prepares the seedling for installation, adaptation to the climate, and productivity (9). Plant bio stimulants such as plant and algae extracts generally fall within these categories i.e. hormoneone of containing products, micronutrients based products, amino acid-containing products, and

humic acid-containing products (10, 11), Plant bio-stimulants are used to treat crops in a commercial setting in view of their ability to increase growth rates. increase stress tolerance, increase the photosynthetic rate (12, 13) ,one endogenous plant hormone, salicylic acid (SA), is essential to plant growth and growth, has demonstrated many key plant functions and can alter the plants' physiological conduct (14), However, SA has been shown to influence a wide range of plant processes, including seed germination and fruit yield (15, 16). Exogenous salicylic acid or indole acetic acid (IAA) has an impact on endogenous levels of soaked grains, as well as germination and maize development, at various concentrations. The study showed that pre-soaking maize seeds absorbed less seed soaked in IAA than SA in various solutions like SA on their intrinsic content in soaking seeds (17). There has been substantial speculation that phenolic in general act as regulators of plant growth (18, 19). Salicylic acid is a plant growth regulator that greatly enhances a plant's bioproductivity. It has a beneficial impact on a variety of vegetable and ornamental plants (20, 12).

Shading is an easy, non-chemical, and lowcost alternative to plastic polyethylene for growing and developing plants in hot weather (21). Under cover, the plant's response to sunlight can be more effective. Since shading has a major impact on plant textures and the percentage of chlorophyll in the leaves, a combination of shading and plant management is needed to improve crop productivity and quality. As a result, the sun It reaches the Shade net house at a lower rate than in the open field, resulting in a more favorable climate for the crop inside the tunnel. Furthermore, under the green and white strips shade net house, growth parameters such as plant height, number of branches, leaf area, diameter significantly and stem were increased, studied by (22). The aim of this study was to improve germination and the quality of the okra seeds and the quality and quantity of the yields.

Materials and Methods:

The experimental period was conducted during (March to October 2020) at two different locations in Sulaimani governorate, Kurdistan Region-Iraq. The first location, in the agricultural field in Tasluja, by 20 km (longitute:45,2474053. from center with latitute:355901740). the second location. Dukan Region of Sulaymaniyah/Iraq by 70.77 from center with (longitude: km 44. 985734.latitude 35.887146) during season 2020. The climate of the study area is classified as semi-arid region, hot and dry in summer and cold in winter (23).

Experimental Design and Treatments

The experiments in both were arranged in a randomized complete block design with three replications. The ground is divided into two longitudinal halves, half of which are covered by a shad net (tunnels) to Knowing the comparison of germination of seeds, plants growth, yield and seed yield between cover plants and uncover plants, the other half was left clear of the shade net cover. Shading was performed on May 12, 2020 and the tunnel cover was removed on June 30 after transplantation, and then the plant feeding programs were randomly arranged with application at different biozyme foliar concentrations (0, 0.75 ml. l^{-1} and 1.5 ml. l^{-1}) done three times, the first Biozyme application was done 3 weeks after transplanting, and repeated every 15 days after the last application during the period, were plants spray early in the morning or late in the evening, to avoid burns of leaves of plants. In this study we will use three factors and their interactions between two location Sulaymaniyah (S) and Dukan (D) as follows:

Factor 1: Soak okra seeds with different concentrations of salicylic acid (0, 75 ppm and 150 ppm) (for 4 hr.)

Factor 2: Biozyme (foliar application) with 3 concentration (0, 0.75 ml. l^{-1} and 1.5 ml. l^{-1}) and spraying the seedlings two weeks after transplanting, 3 times between them 15days.

Factor 3: Covering (covering and without covering), and then the interaction between two locations Sulaymaniyah (S) and Dukan (D) covering (C) and non-covering (NC) ,

salicylic acid (S) and biozyme (B) treatments was taken.

Germination process:

Germination process was performed according to (24), cultivation of okra seeds in bags began after they were treated with salicylic acid at different concentrations (0, 75, 150) ppm, with the use of distilled water on April 6, 2020, then the seeds germinated three days after sowing at a temperature of 12-22 °c, time was calculated after 10 days between the initial and maximum emergence.

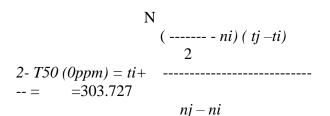
After a perfectly random design the experiment with three replicates was done. Visible radical seeds were regarded as germinated. Dates for germination counts have been recorded daily after the seedling assessment technique provided in the manual of the Association of official seed analysis (25).

At the end of the experiment the final percentage of germination was calculated. Mean germination time (MGT) was derived by the (26).

 $1-MGT (0 ppm) = \frac{\sum Dn}{\sum n} = 6.227$

Were n is the number of germinated seeds on day D, the number of days counted from the started of germination.

The time required for germination T50 was determined based on the modified formula by (27).



Were *N* is the final number of emerged seeds and *ni*, *nj* are the cumulative number of seeds germinated by adjacent seed count at times *ti* and *tj* respectively, when

ni < N/2 < nj.

Germination index (GI) was calculated according to the following formula given by

Association of official seed analysis (28).

	N° of germinated seeds	
N° of germinated	seeds	
3- GI(0ppm) =		_+
+		
	Days of first count	

Days of first count =14.346

On the 7-day following planting, germination energy (GE) was recorded. It is the % of germinating seeds seven days after planting in comparison with the total seeds.

Characteristic Of Quantitative Yield Data Collection

Data collection began 25 days after transplanting the seedlings for a period of 16 weeks. The investigations included data related plant growth and seed yield of local variety okra plant, and a comparison between the two locations of Tasluja and Dukan. Using both salicylic acid and Biozyme, cover plants and without covering in both locations. The cultivar was studied in addition to salicylic acid for seeds by soaking them for 4 hours with distilled water and spraying the seedlings after 3 weeks with Biosyme as a foliar spray for treated and untreated plants season 2020. Fresh okra fruits were harvested manually from the immature fruit of the okra plant in the early morning, fruits were harvested three times a week, leaving a stalk of 2-3 cm attached to the pods, and the following parameters were measured:

Pod Weight (g)

The average weight of the pods was obtained as shown below:

Average pod weight (g) = yield of exp. unit (g)/Number of pods of the exp. unit

Yield Per Hectare (ton / hectare)

The total yield was measured by calculating the plant yield for each experimental unit, then the crop was converted in hectares to tons.

Seed Character

Okra fruit contains many seeds, so twenty pods for each experimental unit were taken without harvest until the end of the season before shattering stage occurs, the seeds of okra may spread up to 2-3 meters.

Number of Seed Per Pod

The number of seeds per pods from twenty pods in each experimental unit was randomly counted and the mean was measured.

Weight of 1000 seed (g)

The average seed weight was obtained from one hundred seeds and then multiplied by ten in each experimental unit for covering and without cover and then the mean was measured.

Statistical Analysis

Statistical analysis of data and comparison of arithmetic averages for evaluated by use oneway analysis of variance (ANOVA). Transactions according were separated by the Duncan test at a level P-value of 0.05 (29) Duncan test is a new multiple range test procedures of the Statistical Analysis System (30).

Results and Discussion:

Germination was observed in the seeds treated with (75 ppm and 150 ppm) and the lowest germination percentage was observed in control.

Factors	0 ppm	75 ppm	150 ppm
MGT	6.226 a	7.069 a	6.414 a
T50	303.727 b	189.084 c	421.643 a
GI	14.348 a	14.673 a	14.558 a
GE (7 days)	223.667 c	296.000 a	260.000 b

Table 1: Effect of salicylic acid on seed okra germination

Different letter in same row means significant differences (P<0.05).

The germination effect of priming the seeds was positive. In table 1, the results showed no significant difference (p > 0.05) in MGT & GI among different levels of the hormonal priming when the seed socked concentrations (0, 75, 150) ppm of salicylic acid for four hours. (31) reported that the mean germination time and germination index was increased in okra seed treating with salicylic acid solution, this is may be due to that the treatment of seeds enhances antioxidants' enzymes such as ascorbate and glutathione in seeds and reduces lipid peroxidation during germination. and increasing germination percentages may be the result of dormancy in fresh seeds after treatment with SA, these findings agree with (32) for the average time required for the maximum germination T50 of a seed lot and can have the same time units used for germination calculation. The highest germination percentage at time to reach 50% germination (T50) observed was in concentration (150 ppm) of SA treatments for four hours caused gradual increases was 421.643 compared with the other concentrations which showed lower values. Seed soaking at concentration (75 & 150) ppm increased germination energy (GE 7 days) compared with control. The reason for this rise may be increasing cell division within the seedling roots and shooting apex that caused plant growth to increase and treatments with salicylic acid maintain cytokinine and IAA in plant tissues, which increase cell division within the apical meristem. These findings also consistent with the results were mentioned by (33, 34).

Pod Weight (g)

The results data reveled in table (2) that the tetra interaction among locations, covering, salicylic acid and biozyme foliar application, significantly (P<0.05) effected on pod weight (g) in both locations, the highest value (4.829)

g) was obtained from okra plant that noncovering in Dukan location when the seeds treated with salicylic acid at (75 ppm) and treated with biozyme at (0.75 ml.l-1) concentration (DNC*S1*B0), while in okra plant from Sulaymaniyah with non-covering when the seeds treated with (75 ppm) and spray okra plants with biozyme at (0.75 ml.l-1) concentration (SNC*S1*B1) recorded the lowest pod weight (3.848 g).

Results of tri-interaction among locations, covering and salicylic acid, show in table (2) indicated that in okra plant from Dukan location treated with non-covering and the seeds treated with 75 ppm of salicylic acid (DNC*S1) was the most significant effective treatment which gave pod weight of (4.699 g). Results obtained Okra plant from Sulaymaniyah with non-covering and the seeds treated at (0 ppm) concentration (SNC*S0) which gave the lowest pod weight (4.029 g).

Concerning the tri interaction among locations, covering and biozyme foliar application results, indicated that effect was significant on pod weight (g), the highest value (4.551 g) was noticed from the Okra plant in Dukan location with non-covering and spray okra plants with biozyme at (1.5 ml.l-1) concentration (DNC*B2), while Okra plant in Sulaymaniyah with covering and plants treated with biozyme foliar application at (0.75 ml.l-1) concentration (SNC*B1) gave the lowest pod weight, which was (665.778 g).

The interaction between locations and covering results, displayed that there were significant (P<0.05) effects on pod weight, the maximum value (4.479 g) recorded in Okra plant from Dukan location with non-covering (DNC), whereas okra plant from Sulaymaniyah with covering (SC) obtained the lowest pod weight (4.052 g).

The results in the same table showed that there were no significant (P>0.05) differences in pod weight when the seeds are treated with salicylic acid at different concentrations. The results in table (2) illustrated that the pod

weight of okra plants spraying with biozyme non-affect significantly (P>0.05).

In both locations, showed that there were no significant (P>0.05) differences in pod weight between covering and non-covering.

Results in the same table demonstrated that, Dukan location gave the best value of pod weight compared to Sulaymaniyah location.

Yield (ton / hectare)

Results in table (3) indicated that the tetra interaction among locations, covering. salicylic acid and biozyme foliar application significantly affected yield per hectare. The highest value (9.491 ton/ hectare) was obtained in Okra plant from Dukan location with non-covering and seeds treated at (75 ppm) of salicylic acid and biozyme foliar application at (0 ml.l-1) concentration (DNC*S1*B0), whereas in Okra plant from Sulaymaniyah with covering and seeds treated with (0 ppm) salicylic acid and biozyme foliar application at (1.5 ml.l-1) concentration (SC*S0*B2) gave the lowest value (2.832 ton/ hectare).

Tabulated data declare that in both locations, the tri interaction among locations, covering and salicylic acid significantly affected yield per hectare, the highest significant value (8.814 ton/ hectare) recorded in Okra plant from Dukan with non-covering when seeds treated with salicylic acid at (75 ppm) concentration (DNC*S1), whereas the Okra plant from Sulaymaniyah with covering and salicylic acid at (75 ppm) concentration (SC*S1) gave the lowest significant yield per hectare, which was (3.930 ton/ hectare).

Concerning the tri-interaction among location, covering and biozyme, the effect was significant (P<0.05) in increasing of yield per hectare. The highest value (8.574 ton/ hectare) was noticed in Okra plant from Dukan location with non-covering and spray okra plants with biozyme foliar application at (1.5 ml.l-1) concentration (DNC*B2), nevertheless the lowest value was obtained in Okra plant from Sulaymaniyah, covering when the okra plants treated with biozyme at (1.5 ml.l-1) concentration (SC*B2) which was (3.468 ton/hectare).

For the interaction between locations and covering, the results shows that the Okra plant from Dukan with non-covering (DNC) gave the highest significant value (8.425 ton/hectare), whereas the Okra plant from Sulaymaniyah and covering (SC) gave the lowest on yield per hectare was (4.016 ton/hectare).

Results in table (3) indicated that there was no significant (P>0.05) effect on yield per hectare when the seeds treated with salicylic acid at different concentration However, biozyme foliar application for okra plants had no significant (P>0.05) effect on yield per hectare.

The recorded data in table (3) demonstrated that there was significance (P < 0.05) effect of covering and non-covering for both locations on yield per hectare, okra plants grown under non-covering gave the highest value (7.048 ton/ hectare) compared with covering.

Treatments						Average of				
	Type of	Salicyli		Biozyme	Salicylic acid		Average of		verage of	
location coverin g		c acid	B0	B1	B2	effect	covering	COV	ering	
	Coverin	S 0	4.365 a-f	4.637 a-d	4.702 ab	4.568 ab	- 4.445 a			
		S 1	4.547 a-f	4.411 a-f	4.138 a-f	4.365 a-c			4.249 a	
	g	S2	4.258 a-f	4.481 a-f	4.468 a-f	4.402 a-c		+J a		
Dukan	Avera Bioz	0	4.390 a	4.510 a	4.436 a			Coverin g		
Dukan	Non	S 0	4.455 a-f	4.285 a-f	4.471 a-f	4.404 a-c	4.479 a			
	coverin	S 1	4.580 a-f	4.829 a	4.688 a-c	4.699 a				
	g	S2	4.116 a-f	4.398 a-f	4.492 a-f	4.335 a-c				
	Average of Biozyme		4.384 a	4.504 a	4.551 a	Average of Dukan effect		4.462 a		
	Coverin g	S 0	4.207 a-f	3.960 c-f	3.936 d-f	4.034 c	- 4.052 b			
		S 1	4.408 a-f	3.939 d-f	3.894 f	4.080 c				
		S2	4.250 a-f	3.976 b-f	3.900 ef	4.042 c		4.032 0 Non		
Sulaiman		Average of Biozyme		3.958 bc	3.910 c			coverin	4.292 a	
i	Non	S 0	4.147 a-f	3.965 c-f	3.974 b-f	4.029 c	4.105 b	g		
	coverin	S 1	4.287 a-f	3.848 f	3.953 d-f	4.029 c	4.105.0			
	g	S2	4.279 a-f	3.863 f	4.627 а-е	4.256 bc				
	Average of Biozyme		4.238 a-c 3.892 c 4.185 a-c		Average of Sulai	mani effect	4.0	78 b		
Total Av	Total Average of Biozyme			B0		B1	B2			
	effect		4.325 a			4.216 a	4.270 a			
Total Av	erage of Sa	licvlic	SO		S 1	S2				
	acid effect		4.259 a			4.293 a	4.259 a			

Table (2): Impact of Seed Soaking by salicylic acid, Biozyme Foliar Application and Covering Okra plant on the Pod Weight (g)

The averages with identical letters (same cells color) are not significantly different (P>0.05) between them

The results in the same table showed that Okra plant from Dukan location gave the best value (6.975 ton/ hectare) compared with the data Okra plant from Sulaymaniyah location on yield per hectare. Biozyme increases the effectiveness of photosynthesis because the symbiotic coexistence between the plant and biozyme helps to absorb nutrients, which increases the growth of the plant (35). This was led to an increase in the weight of pod and plant yield, which is a reflection on the increase of the total yield. Biozyme increased pod weight and yield per hectare when the plants at triple interaction with location, covering and biozyme at high concentration through the formation of more epidermis cells and cell enlargement. The present study supports the findings of (36), this application of Biozyme increases fruit number per plant in tomato by assuring the rapid absorption of plant nutrients by improving the fruit environment and activating the development of larger and quality fruit (37).

The higher concentration of Biozyme of over control gave higher yield, whose yield depends on production of high number of fruits, responded more to biozyme application in the open field, because dependent variable exhibits a highly significant and positive relationship (38, 39, 40) could be caused by an auxin presence (NAA) that encourages the cell proliferation and cell extension and leads to an increase in fruit sink strength. On the other hand, the increasing influence of biozyme on the availability of nutrients and the role of potassium in the plant nutrition can be linked to this increase in plant growth. For the noncovering factors that significantly enhanced vield per hectare compared with covering. This may be due to the fact that okra is one of plants requiring high the temperatures. covering reduced sunlight, delaying flowering and fruit production even after vegetation was removed. significant differences were observed in the number of pods per plant after treating plants with biozyme. The effect of the growing regulators incorporated in the foliar fertilize, this may be due to that biozyme observed rise in okra pod weight results were expected since biozyme is a complex of three growth stimulants (IAA, GA3 and zeatin). (41, 36) observed a mean increase in fruit, weight, chilli and tomato diameters plant in application of biozyme as foliar fertilizer. Early fruit development important for the production of large-size fruits include high amounts of cytokinins (42, 43), gibberellic acid (44), and auxin (45, 46) the partition of photosynthesis into fruit of biozyme treated plants could have changed these plant growth regulators.

Increasing the yield of okra after being treated with biozyme varies according to the location and cover, and that described by(47) which found that after treating tomato with a foliar fertilizer have shown increasing in production and yield, in the case of covering and open failed, it was explained that non-covering gave the best value in terms of yield. Relatively high minimum temperatures during the fruiting period, may favor the ripening of the pod. The harvest season is the optimum temperature for fruit set, Earlier investigations showed that pepper pollen at low temperatures become sterile (48). In the meantime, GA3 encourages other development and raises the amount of sustainable pollen. the plants where treated with three factors, location covering and both concentrations of Biozyme significantly increased the number of fruits in okra plants Biozyme at 0.75 ml.l⁻¹ produced the highest fruit weight, yield per hectare whereas 1.5 ml. Γ^{-1} gave the highest value in these cultivar. Although the nutrient value of fruits is higher for consumers, a high return for farmers is even more vital. Similarly, the ascorbic acid growth in pepper fruits by the use of NAA and GA3 in (49, 50) respectively was detected. Extension of the 5–6-week cover time negative affected the growth properties of okra plants. the fruit yield may decrease due to the lack of sufficient sunlight reaching the plants and may lead to a delay in flowering and failure to pollination (51). In this study, plant coverage period was affected by yield levels and pod weight, which is exactly contrary with the results of (52), who reported that the cover length of melon does not affect fruit yields substantially and fruit weight. The table (3) showed a significant increase in yield per hectare when plants grow in open field were (7.048 ton/ha). The open field temperature is observed to be higher compared to the shade net. non covering led to increase the average pod weight of okra and the share of marketable yield in the total yield. The highest production of okra was found in open field compared to shade net house. Since it provides unsuitable temperature for crop growth and development (53, 54, 55). So, the sun light in the covering or shade net house is less inside the house outside compared to. So, we have a good environment in the shade net house for cultivation. Morning and evening, the light intensity is decreased. This might be due to green color decreases the crop temperature. However, the study conducted by (56) found a 30% drop in the yield of pumpkin by covering, compared to non-cover cultivation. These effects of these foliar fertilizers may be due to their multi-nutrient content, which improved the growth properties of okra plants after absorption from the leaf tissues and consequently raised the yields of okra. Foliar fertilization not only increases plant development characteristics, crop yields and plant nutrient consumption (57) but also improves crop nutrient efficiency (58).

Increasing the yield of okra after being treated with biozyme varies according to the location and cover. Increased yield of okra, tomato and wheat (59) as reported foliar application of liquid fertilizer extracted from seaweed. The foliar fertilization method is therefore regarded as a strategy of environmental fertilization. Reduced floral drop and fruit growth may be related to delay of abscission (cytokinins and auxin effects) by preserving the loss of pectin material in the middle lamella (60) and improving tolerance to water as well as nutrient stress (61). A clear effect was observed with salicylic acid, which increased the yield characteristics at Dukan location, and this is the result of the temperature of the environment and the soil. Soaking of okra seed with SA significantly improved growth and yield characters, both in control with distilled water and treatment condition. The increase in yields results from improved vegetative growth features, which

led to the efficiency of the photosynthesis process being increased as a result of an increase in plant height and the number of leaves per plant and the increase in the amount of carbohydrates produced and transferred to the flowers during the growing period this is consistent with (62).

		Tre	eatments	Average of		1 4	C			
lesstien	location Type of		i Biozyme		Salicylic	Average of	total Average of covering			
location	covering	c acid	B0	B1	B2	acid effect	covering	covering		
		S 0	5.933 c-i	5.905 c-i	5.109 e-j	5.649 b				
	Covering	S 1	5.148 e-j	4.347 g-ј	5.975 c-i	5.157 bc				
		S 2	4.597 g-ј	6.537 b-g	6.171 c-h	5.769 b	5.525 b	Covering		
	Avera Bioz		5.226 b-d	5.596 bc	5.752 bc				4.770 b	
Dukan		S 0	8.801 ab	8.238 a-d	8.380 a-c	8.473 a	8.425 a			
	Non	S 1	9.491 a	8.810 ab	8.141 a-d	8.814 a				
	covering	S2	7.162 a-f	7601.847 a-e	9.201 a	7.988 a				
	Average of Biozyme		8.485 a	8.217 a	8.574 a		Average of Dukan effect		6.975 a	
	Covering	S 0	4.363 g-j	4.778 f-j	2.832 ј	3.991 c				
		S 1	3.656 h-j	4.618 g-j	3.516 ij	3.930 c	4.016 c			
		S2	4.026 g-j	4.299 g-ј	4.056 g-j	4.127 c				
Sulaimani	Average of Biozyme		4.015 de	4.565 с-е	3.468 e			Non covering	7.048 a	
Sulailliaill	Non	S 0	5.783 d-i	5.908 c-i	5.311 e-j	5.667 b				
	covering	S 1	6.047 c-i	6.302 c-g	5.214 e-j	5.854 b	5.671 b			
	covering	S2	5.378 e-i	6.278 c-g	4.819 f-j	5.492 b				
Averag Biozy		0	5.736 bc	5.736 bc 6.163 b 5.115 b-d		Average of	e of Sulaimani effect 4.843 b		343 b	
Total Average of Biozyme effect		B0			B1	B2	_			
	Total Average of Biozyme effect		5.865 a			6.135 a	5.727 a			
Total Average of Salicylic acid		S0			S 1	S2				
effect		5.945 a			5.939 a	5.844 a				

Table (3): Impact of Seed Soaking by salicylic acid, Biozyme Foliar Application and Covering Okra plant on the Yield Per Hectare (ton / hectare)

The averages with identical letters (same cells color) are not significantly different (P> 0.05) between them.

Results of location, covering and biozyme interaction had a direct effect on the yield per hectare of okra plants (8.574 ton/ hectare). However, favorable influence on the fruit of the chili was also detected. Maximum fruit yield of 9.77t ha-• was recorded with treatment T4. (63, 64) showed dramatically higher chili and bell pepper yields by biozyme applications. The data presented in table (2) showed that biozyme at over control concentration had significant effect on pod weight, it is assumed that this significant increase is due to the effect on the respiration process, accumulated nitrates, resistance and growth of plant pods to viruses and diseases (65), or may lead to a higher pod weight due to a good physiological effect and hormone activity as a result of plant cell metabolism, synthesis and raising photo the leaf chlorophyll and nitrogen content (66).

Seed Character

Number of Seeds Per Pod

Results in table (4) illustrated the number of seeds per pod, the tetra- interaction among locations, covering, salicylic acid and biozyme effect significantly (P<0.05) on the number of seeds per pod. Okra plant From Sulaymaniyah location with covering when the seeds treated with (0 ppm) salicylic acid and (0 ml.l-1) biozyme foliar application (SC*S0*B0) gave the highest value was recorded (78.467), while the lowest value was recorded in Okra plant from Dukan location with covering when the seeds treated with salicylic acid at (150 ppm) concentration and sprayed plants at (0 ml.l-1) biozyme (DC*S2*B0), which was (57.833).

Tabulated data showed that the tri-interaction among location, covering and salicylic acid effect significantly (P<0.05) on the number of seeds per pod, Okra plant in Dukan location with non-covering and the seeds treatment with (0 ppm) salicylic acid (DNC*S0) gave the highest value (74.389), while the lowest value was recorded in Okra plant from Sulaymaniyah with non-covering and seeds that treated with (0 ppm) salicylic acid (SNC*S0) (65.989).

Results in table (4) revealed that, the triinteraction among Location, covering and biozyme, significantly (P<0.05) increase the number of seeds per pod, Okra plant from Sulaymaniyah location with covering and biozyme foliar application at (0 ml.l-1) (SC*B0) (74.700), whereas the number of seeds per pod decrease dramatically in Okra plant from Dukan location at (0 ml.l-1) biozyme concentration (DC*B0) was recorded (66.322).

It's clear from same table, the interaction between locations and covering no significant (P>0.05) effect on the number of seeds per pod.

The results in the same table showed that there was no significant (P>0.05) difference on the number of seeds per pod, when the seeds treating with salicylic acid at different concentration on number of seeds per pod.

Regarding in the same table showed that when the okra plants treated with biozyme liquid as a foliar spray there were no significant effect (P>0.05) on the number of seeds per pod.

The results obtained from the present study revealed that the covering and non-covering for both locations had no significant effect (P>0.05) on the number of seeds per pod. The results in the same table showed that location no significant (P>0.05) effecting on the number of seeds per pod. Euphrates Journal of Agriculture Science-13 (4): 136-155, (2021) Abdulrahman and Mahmood

Treatments						Average of	total Average of		
location Type of S		Salicylic	Biozyme		- Salicylic acid effect	Average of	covering		
location	covering		B0	B1	B2	Salicylic actu effect	covering	covering	
		S 0	74.433 ab	69.667 ab	69.633 ab	71.244 a			
	Covering	S 1	66.700 ab	63.000 ab	71.833 ab	67.178 a	68.644 a		
		S2	57.833 b	72.367 ab	72.333 ab	67.511 a	00.044 a		
Dukan	Average of	Biozyme	66.322 a	68.344 a	71.267 a			Covering	698.667 a
Dukan	Non	S 0	76.767 a	70.400 ab	76.000 a	74.389 a	72.615 a	72.615 a	
	Non	S 1	74.533 ab	68.133 ab	72.967 ab	71.878 a			
	covering	S2	70.333 ab	72.267 ab	72.133 ab	71.578 a			
	Average of	Biozyme	73.878 a	70.267 a	73.700 a	Average of D	ukan effect	70.630 a	
		SO	78.467 a	67.533 ab	63.567 ab	69.856 a	71.089 a		
	Covering	S 1	76.600 a	73.867 ab	71.500 ab	73.989 a			
		S2	69.033 ab	73.000 ab	66.233 ab	69.422 a		/1.089 a	Non
Sulaimani	Average of	Biozyme	74.700 a	71.467 a	67.100 a			covering	701.444 a
Sulaillialli	Non	SO	61.600 ab	66.433 ab	69.933 ab	65.989 a		covering	
	Non	S 1	69.600 ab	69.933 ab	68.900 ab	69.478 a	67.674 a		
	covering	S2	69.733 ab	68.367 ab	64.567 ab	67.556 a			
Average of Biozyme		66.978 a	68.244 a	67.800 a	Average of Sulaimani effect		69.382 a		
Total Ayara	go of Piorum	a affaat		B0		B1	B2		
Total Avera	Total Average of Biozyme effect		70.469 a		69.581 a	69.967 a]		
Total Aver	Total Average of Salicylic acid			SO		S 1	S2]	
	effect			70.369 a		70.631 a	69.017 a		

Table (4): Impact of Seed Soaking by salicylic acid, Biozyme Foliar Application and Covering Okra plant on the Number of Seeds Per Pod

The averages with identical letters (same cells color) are not significantly different (P> 0.05) between them.

Weight of 1000 seed (g):

The data in table (5) reveals that the tetrainteraction among locations. covering, salicylic acid and biozyme had a significant (P<0.05) effect on the seed weight per pod. The results indicated that okra plant in Sulaymaniyah with non-covering, when the seeds treated with salicylic acid at (75 ppm) concentration and biozyme foliar application at (0.75 ml.l-1) concentration (SNC*S1*B1) gave the highest value (68.000 g), while the lowest value significantly decreased sharply in the seed weight per pod, was obtained from Okra plant in Dukan location with noncovering which were seeds treated with (75 ppm) of salicylic acid and (1.5 ml.l-1) with biozyme (DNC*S1*B2) it was (48.000 g).

In the same table indicated that the triinteraction among locations, covering and salicylic acid significantly (P<0.05) affected on the seed weight, the highest value (65.444 g) was recorded in okra plant from Sulaymaniyah location with non-covering when the okra seeds treated with salicylic acid at (75 ppm) (SNC*S1), while the lowest value recorded in okra plant from Dukan with noncovering and the seeds treated with salicylic acid at (75 ppm) (DNC*S1), it was (55.778 g).

The data in table (5) concerning the triinteraction among locations, covering and biozyme foliar application, significantly (P<0.05) affected on the seed weight per pod, okra plant in Sulaymaniyah location with noncovering and plants were treating with (0.75 ml.l-1) of biozyme (SNC*B1) recorded highest value(66.222 g). However, the lowest value recorded in okra plant from Dukan location with non-covering when okra plants treated with (0.75 ml. l-1) of biozyme (DNC*B1) (56.778 g).

In the same table the data showed that the interaction between location and covering significantly (P<0.05) effected on the seed weight per pod, okra plant in Sulaymaniyah location and non-covering (SNC) gave the highest value (64.407 g), whereas the lowest value obtained from okra plant in Dukan

location and non-covering of plants (DNC), it was (59.296 g).

Results in table (5) illustrated that there were no significant (P>0.05) differences on the seed weight per pod when okra seeds treating with salicylic acid at different concentration.

On the other hand, the results in the same table showed that when okra plants sprayed with biozyme at different concentration no significant (P>0.05) effect on the seed weight per pod.

Regarding the results, it indicated that there were no significant (P>0.05) differences on the seed weight per pod between covering and non-covering.

The results presence in the same table showed that the best value was obtained from Okra plant in Sulaymaniyah location (63.444 g) compared to Dukan location.

Results of tetra interaction (location, covering, salicylic and biozyme) indicated that the interaction (SC*S0*B0) was the most significant efficient treatment as it gave the highest value (78.467) that showed in table (4). Significantly, the highest value of this parameter was obtained from the tri-interaction (SC*B0) which was (74.700).

The improvement of the performance of the covering plants was attributed to a larger number of branches carrying more pods (67), a higher photosynthesis rating, accumulated more photosynthesis leading to improved pod count and seed per pod. The number of seeds ranged between (57.833 to 78.467) per pod attributed diversion may be to of photosynthetic and metabolites producer by leaves to strong carbohydrate (68). Number of pods is dependent on the seeds producing signal molecules that regulate cellular division and expansion. Auxin is a candidate for the signal molecules one of the biozyme (69). components Biozyme contains cytokinins which could also have encouraged cell division, boosted sink activity to enhance fruit growth in competition with resources and hence improve fruit production (70). The results in table (4-5) showed that the tetra among interaction (location, covering.

salicylic acid and biozyme) demonstrated that, the use of biozyme as foliar spray on okra plants reduces dramatically the number of seeds whereas seed weight increases considerably over control (36) were found contrary results in tomato plants.

More branches to carry more pods (67), salicylic acid and biozyme, increased photosynthetic rate, accumulation of more photosynthetic results, better pod dry number and the number of seeds, led to a larger rates for photosynthesis On plants sprayed with biozyme, the lowest number of seeds (57.833) was recorded at (0 ml.l⁻¹) concentration of biozyme, the correlation matrix of all independent variables contributing to the okra shows an important connection. Though there was no significant improvement of number of seeds per pod and seed weight when the plants treated with biozyme, these observations on the yield characteristics of okra in this study could be attributed to the synergistic effect of gibberellin, IAA, and cytokinin.

The interaction among three factors increased the number of seeds per pod in Sulaymaniyah location the plants grow under covering when the plants treated with biozyme at $(0 \text{ ml. } 1^{-1})$. However, the seed weight increased at the same location in open failed with treating plants with biozyme at $(0.75 \text{ ml.} 1^{-1})$. The differences between number of seed and seed weight were significant between the control and biozyme treatments. The larger seed numbers in the Biozyme control plants could be attributed to photosynthesis in fruit development, which would eventually support seed development.

Seed composition will lead to greater seed production through the shift of components of photosynthesis from vegetative to reproductive organs (72).

Biozymes at the control levels lead to increase the plant height and number of leaves to appear with good physiological effects, including effects of the plant cell metabolism and increased leaf chlorophyll levels (73). The application of biozyme at different concentration not significant differences on the number of seed and seed weight per plant, these results are contrary to the researchers' findings (74, 75). The covering and noncovering of plants had no substantial effect on the seed per pod. It can be owing to the maximum pod size containing more seeds. Similar findings were observed by (76, 77). Due to maximum seed size and per pod of seed, a relation exists between the number of seeds per pod and the weight of the seeds. Similar findings have been achieved (78, 79). Euphrates Journal of Agriculture Science-13 (4): 136-155, (2021)

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Treatments						Average of	· ·····			
location Type of		Salicylic			Salicylic	Average of covering	total Average of covering			
location	covering		B0	B1	B2	acid effect				
	Couring	SO	58.333 b-d	63.000 a-c	62.000 a-d	61.111 а-с	59.741 b			
	Covering	S 1	59.667 a-d	58.000 b-d	59.667 a-d	59.111 cd		41 b		
		S2	56.000 cd	59.667 a-d	61.333 a-d	59.000 cd			61 111 0	
Dukan	Average of	Biozyme	58.000 cd	60.222 b-d	61.000 b-d			Covering	61.111 a	
	Non	S 0	63.000 a-c	53.333 de	64.000 a-c	60.111 b-d	- 59.296 b			
	covering	S1	60.333 a-d	59.000 a-d	48.000 e	55.778 d				
		S2	65.667 ab	58.000 b-d	62.333 a-d	62.000 a-c				
	Average of	Biozyme	63.000 а-с	56.778 d	58.111 cd	Average	e of Dukan effect	59.519 b		
		S 0	67.333 ab	60.000 a-d	62.333 a-d	63.222 а-с	62.481 a			
	Covering	S 1	60.333 a-d	64.667 a-c	63.667 a-c	62.889 a-c				
		S2	61.333 a-d	62.667 a-c	60.000 a-d	61.333 а-с			Non	61.852 a
Sulaimani	Average of		63.000 a-c	62.444 a-c	62.000 a-c			covering	01.0 <i>J2</i> a	
Sulaman	Non	SO	65.000 a-c	65.667 ab	64.000 a-c	64.889 ab		Covering		
	covering	S1	63.000 a-c	68.000 a	65.333 ab	65.444 a	64.407 a			
		S2	65.000 a-c	65.000 a-c	58.667 b-d	62.889 a-c				
Average of B		Biozyme	64.333 ab	66.222 a	62.667 a-c	Average of	of Sulaimani effect	63.	.444 a	
Total Avera	Total Average of Biozyme effect		B0		B1	B2				
Total Average of Biozylie effect		62.083 a			61.417 a	60.944 a				
Total Average of Salicylic acid effect		SO		S1	S2					
		62.333 a			60.806 a	61.306 a				

Table (5): Impact of Seed Soaking	y by salicylic acid. Biozyme Folia	· Application and Covering Okra	plant on weight of 1000 seed (g)
	5 - 5		

The averages with identical letters (same cells color) are not significantly different (P> 0.05) between them.

Conclusion

According to the results obtained from this study, the following could be concluded:

- 1. Seed priming at various doses of salicylic acid promote germination and seed growth in okra seeds.
- 2. It is evident that the application of Shading, plant bio stimulants significantly improved plant growth, fruit set, and yield. So, it can be concluded that spray not only gave more yield to the farmers but also improve the fruit quality.

References:

- Naveed, A., Khan, A.A., & Khan, I.A. (2009). Generation mean analysis of water stress tolerance in okra (Abelmoschus esculentus L.). Pak. J. Bot., 41: 195-205.
- Oyelade, O.J., Ade-Omowaye, B.I.O., and Adeomi,V.F. (2003). Influence of variety on protein, fat contents and some physical characteristics of okra seeds. J. Food Eng., 57: 111-114.
- Andras, C.D., Simandi, B., Orsi, F., Lambrou, C., Tatla, D.M., Panayiotou, C., Domokos, J., &Doleschall. F. (2005). Supercritical carbon dioxide extraction of Okra (Hibiscus esculentus L.) seeds. J.Sci. Food Agric., 85: 1415-1419.
- Saifullah, M., & Rabbani, MG. (2009). Evaluation and characterization of okra (Abelmoschus esculentus L. Moench.) genotypes. SAARC J. Agric.7: 92-99.
- Sathish, D., and Eswar, A. (2013). A Review on:Abelmoschus esculentus (Okra). Int. Res J Pharm.App Sci., 2013; 3(4):129-132.
- Nzikou, J., Mvoula-Tsieri, M., & Matouba, E. (2006). A study on gumbo seed grown in Congo Brazzaville for its food and industrial applications. African Journal of Biotechnology Vol. 5 (24), pp. 2469-2475.
- Wild, A. (2003). Soils, land and food: Managing the land during the twentyfirst century (1st ed.). Cambridge, United Kingdom: Cambridge

University Press.

- National Research Council, (2006). "Okra". Lost Crops of Africa: Volume II: Vegetables. Lost Crops of Africa. 2. National Academies Press. ISBN: 0-309-66582-5, 378 pages Naveed, A., Khan, A.A., & Khan, I.A. (2009).Generation mean analysis of water stress tolerance in okra (Abelmoschus esculentus L.). Pak. J. Bot.,41: 195-205.
- 9. Zhu, J. K. (2002). Salt and Drought Stress Signal Transduction in Plants. Annual Reviews Plant Biology, 53, 247-273.
- Al-Khafajy R.A., AL-Taey D. K.A. and AL-Mohammed M.H.S. 2020. The impact of Water Quality, Bio fertilizers and Selenium Spraying on some Vegetative and Flowering Growth Parameters of *Calendula Officinalis* L. under Salinity Stress. *Int. J. Agricult. Stat. Sci.*, 16, Supplement 1:: 1175-1180.

https://connectjournals.com/03899.202 0.16.1175

- 11. Al- Duraid M.H., K. A. Al-Taey, and A. H. J. Al-Kikhani.(2019) Effect of Phenylalanine and Naphthalene Acetic Acid Growth, on Yield and Antioxidant Activity of Fenugreek foenum-graecum. Trigonella IOP Conf. Series: Earth and Environmental Science: 388 (2019)012073 doi:10.1088/1755-1315/388/1/012073.
- 12. Al-Taey, D.K.A., AL-Naely, I J.C., Kshash B.H. (2019). A study on effects of water quality, cultivars, organic and chemical fertilizers on potato (*Solanum tuberosum* L.) growth and yield to calculate the economic feasibility. *Bulgarian Journal of Agricultural Science*, 25 (6) 1239 -1245.
- 13. Manea, A.I., AL-Bayati, H.J. and AL-Taey, D.K.A . 2019. Impact of yeast extract, zinc sulphate and organic fertilizers spraying on potato growth and yield. *Res. on Crops* 20 (1) : 95-100. DOI : 10.31830/2348-7542.2019.013.

- 14. Al-Taey D.K.A. (2009).Effect of spraying acetyl salicylic acid to reduce the damaging effects of salt water stress on orange plants (*Citrus sinensis* L.). Journal of Kerbala University; 7 (2): 192-202.
- 15. Cutt JR, Klessig DF. Salicylic acid in plants a changing perspective. Pharmaceut Technol. 1992; 16:26-34.
- 16. Al-Taey, D.K.A. and Saadoon, A.H. (2012). Effect of treatment of kinetin to reduce the salinity damage by drainage water irrigation on the growth and nitrate accumulation in the leaves of spinach, *Spenacia oleracea* L. Euphrates Journal of Agriculture Science. 4(4):11–24.
- 17. El-Mergawi, R. A. en Abd El-Wahed, M. S. A. (2020) "Effect of exogenous salicylic acid or indole acetic acid on their endogenous levels, germination, and growth in maize", *Bulletin of the National Research Centre*. Bulletin of the National Research Centre, 44(1).
- Aberg B. Plant growth regulators. XLI.Monosubstituted benzoic acids. Swedish J. Agric. Res.1981; 11:93-105.
- 19. Attiya R.L., R.N. Al-Ubori, S. M. Alrubaye, M. M. Gani, A. S. Shakir and Duraid K.A. AL-Taey,(2020) Physiological Effect of the Water Sources and Salicylic Acid on the Germination and Some Seedling Traits of Sorghum. *Indian Journal of Ecology* ., 47 Special Issue (12): 275-280.
- 20. Hayat,S., Ali, B. & Ahmad, A. 2007. Salicylic Acid: Biosynthesis, Metabolism and Physiological Role in Plants. In: S. Hayat and A.Ahmad :Salicylic acid: A Plant Hormone. Springer, Netherlands. pp: 1-23.
- Argade, M. B., Kadam, J. H., Garande, V. K., Patgaonkar, D. R., Patil, V. S., & Sonawane, P. N. (2018). Effect of different shading intensities on growth and yield of cherry tomato. *Journal of Applied and Natural Science*, 10(1), 352-357.

- 22. Semida, W. M., Ammar, M. S. and EL-Sawah, N. A. (2017)'Effect of shade level and microenvironment on vegetative growth, physiological and biological characteristics of transplanted cucumber (*Cucumis sativus*)', *Archives of Agriculture and Environmental Science*, 2(4),pp.361-368.
- 23. Najmaddin, P. M., Whelan, M. J. and Balzter, H. (2017), Estimating daily reference evapotranspiration in a semiarid region using remote sensing data, Remote Sensing, 9(779), pp. 1-20.
- 24. Al-Maskri, A., Khan, M.M., AlManthery, O. and Al-Habsi, K .
 2002. Effect of accelerated aging on lipid peroxidation, leakage and seedling vigor (RGR) in cucumber (Cucumis sativus L.) seeds. Pakistan J.Agri. Sci. 39: 330–337.
- 25. AOSA. Official methods of analysis 15h Edition, Association of Official Analytical Chemists, Arlington, Washington D.C, 1990
- 26. Ellis, R.H., and E.H. Roberts. 1981.The quantification of aging and survival in orthodox seeds. Seed Science and Technology 9:373-409.
- Farooq, M., S.M.A. Basra, K. Hafeez, and N. Ahmad. 2005. Thermal hardening: A new seed vigor enhancement tool in rice. Journal of Integrative Plant Biology 47:187-193.
- 28. Association of Official Seed Analysts (AOSA). 1983. Seed Vigour Testing handbook. Contribution 32, Handbook on Seed Testing, AOSA, Lincoln, NE, USA.
- 29. Al- Rawi, K.M. Abd alaziz.M.K.2002.Design and analysis of Agriculture experiments. Library for printing and publishing –Univ. of Mosul. In Arabic, pp:88.
- 30. SAS Institute, Inc (2007). Statistical analysis system. SAS institute Inc., Cary, NC. USA.
- 31. Hussein H J (2015) Effect of Seed Priming treatment with Salicylic Acid on Viability of Okra (Abelmoschus

esculentus L.) Seeds. Euphrates Journal of Agriculture Science-7(2):1-9.

- Bayuelo-Jimenez J.S., Craig R., Lynch J.P. (2002). Salinity tolerance of Phaseolus species during germination and early seedling growth. Crop Sci 42: 1584–1594.
- 33. Rehman H, M Farooq, SMA, Basra I, Afzal (2011) Hormonal priming with salicylic acid improves the emergence and early seedling growth in cucumber. J. Agric. Soc. Sci. 7:109– 113.
- 34. Sakhabutdinova AR, DR Fatkhutdinova, MV Bezrukova, FM Shakirova, (2003) Salicylic acid prevents damaging action of stress factors on wheat plants. Bulgerian J. Plant Physiol. 314–319.
- 35. Buchanan, B. B., Gruissem, W., & Jones, R. L. (Eds.). (2015). *Biochemistry and molecular biology of plants*. John wiley & sons. Courier Companies, Inc., USA. 1367 pp.
- 36. Ofosu J. Anim, E. T. Blay& L.Bening (2007). Effect of Biozyme T.F. on yield and quality of tomato (*Lycopersicon esculentum*). Ghana Jnl agric. Sci. 40, 113-117.
- 37. Saimbhi, M. S., Padda, D. S. and Singh, G. (2012). Ascorbic acid content of chilli varieties as affected by fruit maturity. J. Res., 9: 248-50.
- 38. Quintalan, L., & Rojas Garciduenas, M. (1990). Effects of three phytoregulators on the development and yield on Tomato (Lycopersicon esculentum). Serie Fitociencia.
- 39. Abdel Mawgoud, A. M. R.; N. M. El Greadly; Y. I. Helmy and S. M. Singer (2007). Responses of tomato plants to different rates of humic based fertilizer and NPK fertilization. Jornal of Applied Sciences Research 3(2): 169-174.
- 40. Sarhan, Z. T. (2011). Effect of humic acid and seaweed extracts on growth

and yield of potato plant (Solanum tubersum L) Desiree cv. Mesopotamia Journal of Agriculture, 39(2), 19-25.

- 41. Manna, D., Sarkar, A. and Maity, T.K. (2012) Impact of biozyme on growth, yield and quality of chilli (Capsicum annuum L.). Journal of Crop and Weed 8(1): 40-43.
- 42. Cutting, J. M. (1993) The cytokinin complex as related to small fruit. Acta Hort. 329, 147-149.
- 43. Cowan, A. K., Moore-Gordon, C. S., Bertling, I. & Wolstenholme, B. N. (1997) Metabolic control of avocado fruit growth. Pl. Physiol. 114, 511-518.
- 44. Salazar-Garcia, S. & Lovatt, C. J. (2000) Use of GA3 to manipulate flowering and yield of the 'Haas' avocado. J. Am. Soc. Hort. Sci. 125, 25-30.
- 45. Vanderpuije, S. N. (1998) Effect of auxin on fruit size and quality of tomato (*Lycopersicon esculentum*) (B Sc Dissertation). Department of Crop Science, University of Ghana, Legon..
- 46. Lovatt, C. J. (1999) Timing citrus and avocado foliar nutrient applications to increase fruit set and size. Hort. Technol. 9, 607-612.
- 47. Padem, H., Ocal, A., Senguin, A. & Bieche, B. J. (1999) Effect of foliar fertilizers on yield and some Charscteristics of processing tomato. Acta Hort. 487, 225-228.
- 48. Polowick, P.L. and Sawhney, V.K., 1985. Temperature effects on male fertility and flower and fruit development in Capsicum annuum L. Sci. Hortic., 25: 117-127.
- 49. Patil, U.B., Sangale, P.B. and Desai, B.B., 1985. Chemical regulation of yield and composition of chilli (Capsicum annuum L.) fruits. Cum Res. Rep., 1: 39-41. (Hot-tic. Abstr., 56: 345.).
- 50. Zayed, E.A., El-Zawily, AI., Nofal, E.S. and Hassan, M., 1985. Studies on growth, productivity and some physiological aspects of hot pepper (Capsicum annuum L. var. Red

Cherry). I. Effect of morphactin, gibberellic acid and their combination. J. Agric. Sci. Mansoura Univ., 10: 193-190.

- 51. Rodrigo Gómez S, Ornosa C, Selfa J, Guara M, Polidori C. Small sweat bees (Hymenoptera: Halictidae) as potential major pollinators of melon (Cucumis melo) in the Mediterranean. Entomol Sci. 2016;19(1):55–66. <u>https://doi.org/10.1111/ens.12168</u>.
- 52. Santos FGB, de Negreiros MZ, de Medeiros JF, de Sousa Nunes GH, de Medeiros DC, Grangeiro LC. Produção e qualidade de melão Cantaloupe em cultivo protegido temporariamente com agrotêxtil em Mossoró, Rio Grande do Norte. Revista Ceres. 2015;62(1):93–100. https://doi.org/10.1590/0034-

737x201562010012.

- 53. Bastias Richard M., Manfrini Luigi and Grappadelli Luca Corelli. 2012. Exploring the potential use of photoselective nets for fruit growth regulation in apple. Chilean J. Agril. Res. 72(2): 224-231.
- 54. Bhardwaj, R. 2013. Effect of mulching on crop production under rainfed condition, Agricultural Reviews. Vol. 34 Issue 3, p188.
- 55. Bhadur A., Singh K. P., Rai A., Verma A. and Rai, M. 2009. Physiological and yield response of okra (Abemoschuse sculentus) to irrigation scheduling and organic mulching. Indian journal of agri. sci., 79(10): 813-15.
- 56. Gordon GG, Wheeler GF, Stewart TR, Brown JE, Vinson E, Woods FM. Plastic mulches and row covers on growth and production of summer squash. International Journal of Vegetable Science. 2008;14(4):322– 338. https://doi.org/10.1080/193152608022

https://doi.org/10.1080/193152608022 15830

57. Maitlo, A., Zia-ul-hassan, A. N. Shah and H. Khan. 2006. Growth, yield and nutrient uptake of wheat (Triticum aestivum L.) in relation to foliar and soil application of urea. Int. J. Agri. Biol., 8: 477-481.

- 58. Fageria, N. K., M. P. B. Filhoa, A. Moreirab and C. M. Guimaresa. 2009.
 Foliar fertilization of crop plants. J. Plant Nutr., 32 (6): 1044 –1064.
- 59. Singh P K & Chandel A S, Effect of Biozyme on yield and quality of wheat (*Triticum aestivum*), Indian J Agron, 50 (2005) 58-60.
- 60. Kachave, D.B. and Bhosale, A.M.2007. Effect of plant growth regulators and micronutrients on fruitng and yield parameters of Kagzi lime (Citrus urantifoliaswingle) fruits.Asian Journal of Horticulture 2: 75-79.
- 61. Fujioka, S.S.A.1997. Biosynthesis and metabolism of brassinosteroids. Physiologic plant arum 100: 710-715.
- 62. Al-Mamkagh,A. 2009. Effect of Tillage Time and plastic mulch on Growth and yield of okra (Abelmoschus esculentus) Grown under Rain – fed Conditions. Int.J. Agric. Biol., 11(4):453-457.
- 63. Gore, A. K., Jadhav, S. B., Gore, A. K. and Ghuge, T. D. 2007. Effect of different bioenzymes on growth, flowering and yield of green chilli (*Capsicum annuum* L.) variety "Pusa Jwala". J. Soils and Crops. 17: 105-09.
- 64. Kumar, P., Dube, S. D. and Chauhan, V. S. 2000. Photosynthetic response of bell -pepper to Biozyme in relation to fruit yield. Veg. Sci., 27: 54-56.
- 65. Unlu, O; H. Unlu; Y. Karakurt and H. Padem (2011). Changes in fruit yield and quality in response to foliar and soil humic acid application in cucumber. *Scientific Research and Essays*, 6: 2800 - 2803
- 66. Sure,S; H. Arooie; K. Sharifzade and R. Dalirimoghadam (2012).Response of productivity and quality of cucumber to application of the two biofertilizer (humic acid and nitroxin) in fall planting. *Agricultural Journal*, 7(6): 401-404.

- 67. Omer, E.A., Khattab,M.E. & Ibrahim, M.E. (1997). Effect of pinching and foliar application of some growth regulators on two new early mature varieties of *Hibiscus sabdariffa* L., Egypt J.Hort. 24, pp. 117-130.
- 68. Tripathi. M. K.; B. Chaudhary; S.R. Singh and H. R. Bhandari (2013). Growth and yield of sunnhemp (*Crotalaria juncea* L.) as influenced by spacing and topping practices. African Journal of Agricultural Reserch Vol.8(28),pp. 3744-3749.
- 69. Magnus, V., Ozga, J. A., Reinecke, D. M., Pierson, G. L., Larue, T. A., Cohen, J. D. & Brenner, M. L. (1997)
 4-chloroindole-3-acetic acid in Pisum sativum. Phytochem. 46, 675-681.
- 70. Bower, J. P. & Cutting, J. G. (1988) Avocado fruit development and ripening physiology. Hort. Rev. 10, 229-271.
- 71. Rahmani, M.; D. Habib; A. H. Shiranirad; J. Daneshian; S. A. R. Valadabadi; M. A. Boujar and A. H. Khalatbari (2010). The effect of super absorbent polymer on yield, antioxidant enzymes (catalase and superoxide dismustase) activity and cell membrane stability in mustard under water deficiency stress.
- 72. Nardi, S.; D. Pizzeghello; A.Muscolo and A. Vianello (2002). Physiological effects of humic substance on higher plants. Soil Biochem. 34: 1527-1536.Hort. Sci., 29 (3/4): 266-267.
- 73. Khafaga, E. E. E.; S. A. Hasanin and R. M. El- Shal (2014). Effect of foliar application with Ascorbic, Humic acid and Compost Tea on nutrients content and Faba bean productivity under sandi

soil conditions. J. Soil Sci. and Agric. Eng., Mansoura Univ., Vol. 5(6): 767 – 778.

- 74. Barakat M.A.S.; A. Sh. Osman, W.M. Semida and M.A.H. Gyushi (2015) influence of potassium humate and ascorbic acid on growth, yield and chemical, composition Journal of Academic Research Vol. 7. No. 1. Pp. 192 – 199.
- 75. Olorunmaiye, K. S. (2010). Reproductive Performance of Two Cowpea ('Vigna unguiculata'(L) Walp) Varieties Ife Brown and TVX3236 as Influenced by Imidazolinone and Dinitroaniline Herbicides. *Australian Journal of Agricultural Engineering*, 1(3), 101-105.
- 76. Choudhary, A. K., Kumar, S., Patil, B. S., Sharma, M., Kemal, S., Ontagodi, T. P., ... & Vijayakumar, A. G. (2013). Narrowing yield gaps through genetic improvement for Fusarium wilt resistance in three pulse crops of the semi-arid tropics. SABRAO Journal of Breeding and Genetics, 45(03), 341-370.
- 77. Madukwe D. K.; H.C. Ogbuehi and M.
 O. Onuh (2012). Effects of weed control methods on the growth and yield of cowpea (Vigna unguiculata (L.) Walp.) under rain-fed conditions of Owerri. American- Eurasian Journal of Agricultural and Environmental Sciences 12 (11): 1426-1430.
- 78. Usman, I. (2013). Effect of pre emergence herbicides on weed control and performance of cowpea in Samaru. Journal of Agricultural Sciences (Sri Lanka) 8(2): 76-8.