

EVALUATION OF THE EFFICIENCY OF SOME INTEGRATED PEST MANAGEMENT AGENTS AND THEIR INTERACTION FOR CONTROLLING EGGPLANT ROOT ROT DISEASE CAUSED BY SOME FUNGI ISOLATED FROM DIFFERENT AGRICULTURAL AREAS

Rehab Adil A. AL-Ajeely and Ahed A. H. Matloob*

Al-Mussaib Technical College, Al-Furat Al-Awsat Technical University, 51009 Babylon, Iraq. E-mail: com.ahd@atu.edu.iq

Abstract

The study aimed to evaluate the effectiveness of *Pseudomonas fluorescens* and the chemical inducer Chitosan in combating some causes of eggplant root rot disease. The results of isolation and diagnosis showed the presence of several genera of fungi isolated from the roots of eggplant plants infected with root rot disease. The results showed the ability of *P. fluorescens* bacteria to inhibit the growth of the pathogenic fungi *F. solani* and *R. solani* Fs2 and Rs3 on PDA medium and the percentage of inhibition reached 100.00%. The results showed that all the factors used in the experiment, including *Pseudomonas fluorescens* and chitosan, whether applied individually or in combination, had a significant effect in reducing the percentage and severity of eggplant root rot disease compared to the pathogenic fungi treatments alone (*R. solani* and *F. solani*), where the infection rate reached 100% and the disease severity was 80% and 75%, respectively. The integration treatment between *P. fluorescens* and chitosan in the presence of pathogenic fungi showed significant superiority over other treatments in reducing the infection percentage and severity, with infection rates of 16.67% for both, while the disease severity was 5% and 8.33%, respectively. The results showed that all treatments involving the biocontrol agent *Pseudomonas fluorescens*, chitosan, led to an increase in certain growth parameters of eggplant, including plant height, root length, root volume, and the fresh and dry weights of both the shoot and root systems, compared to the treatment with the pathogenic fungi alone (*R. solani* and *F. solani*), which significantly reduced all the studied growth parameters.

Key words: Eggplant, root rot, biological control, *Pseudomonas fluorescens*, Chitosan

-1Introduction:

Eggplant *Solanum melongena* L. crop in open and protected cultivation is affected by many pathogens such as fungi, bacteria, viruses and nematodes that cause a reduction in the yield in quantity and quality [34]. The most important of these diseases is root rot disease. Symptoms of seedling death and root rot disease are characterized by a decrease in the number of emerging plants due to the attack of fungal pathogens on seeds and seedlings [11]. It can also cause the appearance of different

degrees of rot on the primary and secondary roots and reduce the formation of nodules, which affects the absorption of water and nutrients [15]. The modern trend in combating various agricultural pests tends towards using integrated pest management to reduce the use of chemical pesticides and limit their environmental and economic harms. Therefore, the efforts of most researchers at present have turned to using various possible methods to move away from the use of

chemical pesticides, such as using biological agents as a practical and safe solution to control diseases, especially root diseases, including root rot disease, due to the many complex problems it causes. Biological control agents are used as an alternative to pesticides, as these organisms act as biopesticides that improve production and protect the crop [32,13]. Among the biological agents used to combat pathogenic fungi are Plant Growth Promoting Rhizobacteria (PGPR) [27]. Also, many recent studies have shown the importance of using chitosan as a component of the cell walls of some living organisms such as crustaceans such as crabs and shrimp and some insects, and it is also a major component of the cell walls of some fungi such as *Aspergillus* and *Mucor*. Chitosan plays an important role in inducing plant resistance against pathogens, including eggplant, cowpea, grapes, cucumbers, and sugarcane [31,22]. This study aims to assess the efficacy of Plant Growth Promoting Rhizobacteria (PGPR) and the chemical inducer Chitosan in mitigating root rot disease in eggplant, addressing the significant impact of seedling drop and root rot on the crop. It seeks to fill the research gap regarding biological control methods compared to a chemical pesticide and to evaluate the effectiveness of these control agents in inducing systemic resistance and enhancing growth parameters in the plant.

-2Materials and methods

-21Isolation and diagnosis of fungi associated with the roots of infected eggplant plants

The isolation process was carried out from eggplant samples that showed symptoms of root rot disease represented by wilting, yellowing of leaves and general weakness in growth, with the presence of brown rots on the main and secondary roots, which were

collected in the field survey. The sterilized pieces of infected roots were transferred using sterile forceps to Petri dishes with 4 pieces in each dish (dish diameter 9 cm) containing the culture medium Potato Dextrose Agar (PDA), the plates were incubated at $25\pm 1^{\circ}\text{C}$ for three days. Then, the diagnosis and purification process was carried out, and the genera were identified by Dr. Ahed Abd Ali Hadi based on the taxonomic keys [19,33,9,12]. The percentage of fungal appearance was calculated according to the following equation [20-:]

Fungi appearance = (Number of root pieces in dishes in which pathogenic fungi appear / The total number of root pieces used for each sample) * 100

2-2Pathogenicity Test:

1-2-2Detection of pathogenic isolates of *Rhizoctonia solani*, *Fusarium solani* and *Macrophomina phaseolina* using cabbage seeds on Potato Dextrose Agar (PDA) culture medium

The best pathogenic fungal isolates were selected, and the best among them was chosen. the pathogenicity of 7 isolates of *F. solani*, 7 isolates of *R. solani* and 6 isolates of *M. phaseolina* was tested according to the method of [8]. The PDA dishes were inoculated in their center with a 0.5 cm diameter disk taken from the edges of the colonies of *F. solani*, *R. solani* and *M. phaseolina*. The dishes were then incubated in an incubator at a temperature of $25\pm 2^{\circ}\text{C}$ for three days, after which the cabbage seeds at a rate of 10 seeds in each dish. The dishes were incubated in the incubator at a temperature of $25\pm 1^{\circ}\text{C}$ until the seeds in the comparison treatment were fully germinated. The percentage of germination was calculated.

2-2-2the pathogenicity of the fungal isolates on eggplant seeds

This experiment was carried out under the conditions of the net shade (Siran) affiliated with the Department of Biological Control Technologies at Al-Mussaib Technical College in 2024. The fungal isolates (Fs-2, 3Fs, and Fs-4) of the pathogenic fungus *F. solani* and the fungal isolates (Rs-3, Rs-4, and Rs-5) of the fungus *R. solani* (which showed the highest inhibition rate on the amaranth seeds) were added in section 2-2-1, loaded onto local millet seeds [10]and added to sterile mixed soil distributed in plastic pots. Ten sterilized local eggplant seeds were planted in each pot, and the pots were carefully watered. The germination rate was calculated after the seeds in the control treatment had fully germinated.

2-3Determination of the effective concentration of the *Pseudomonas fluorescens* suspension that inhibits the growth of the two pathogenic fungi *Fusarium solani* and *Rhizoctonia solani*

Pseudomonas fluorescens bacterial inoculum was obtained from plant diseases laboratory (These bacteria were isolated from healthy eggplant plants rhizosphere that showed signs of good growth and production. They were diagnosed using PCR technology, tested for their effectiveness, and published in previous studies by the researcher., the bacterial isolate was grown on the liquid activation medium Nutrient Broth, The inoculated flasks were incubated at a temperature of 37°C for 48 hours [7]. A series of dilutions of the bacterial suspension of *P. fluorescens* were prepared from 10⁻¹ to 10⁻⁵, then the plates containing the PDA culture medium were inoculated at a rate of 1 ml/plate for each dilution of the bacterial suspension. The plate was moved with a rotating motion to distribute the

bacterial inoculum. A disk was taken from the edge of the fungal colony with a diameter of 0.5 cm from the fungal colony of *F. solani* isolate Fs 2 and *R. solani* isolate Rs 3, which were grown on the PDA medium . The plates were incubated at a temperature of 28±2°C. After that, the amount of inhibition was calculated after reaching The comparison treatment of the edge of the plate was done by calculating the diameter of the growing fungal colony and the percentage of inhibition was calculated according to the equation [25.]

$$\% \text{Inhibition} = [(R - r) / R] \times 100.$$

Where, r is the radius of the fungal colony against the bioagents and R is the radius of the fungal colony without the bioagents .

According to these results, one bacterial isolate that showed high antagonistic ability to inhibit pathogenic fungi was selected for subsequent experiments.

2-4 Evaluation of the efficiency of *Pseudomonas fluorescens* bacteria, the chemical inducer chitosan, and the pesticide Beltanol in resisting *Fusarium solani* and *Rhizoctonia solani* fungi, which cause root rot diseases in eggplant plants, and some growth parameters under greenhouse conditions.

This experiment was conducted in the plastic greenhouse belonging to the Department of Biological Control Technologies at Al-Mussaib Technical College for the autumn season of 2024-2025. A mixture of soil and peat moss (1:2) was prepared and sterilized using commercial formalin at a rate of 20 ml/liter of water (with a commercial formalin concentration of 40%). The formalin was sprayed onto the soil after it was gathered on nylon, mixed, and then covered well with transparent nylon for 7 days under sunlight. Afterward, the soil was left to ventilate for three days before use [5]. The soil was then distributed into plastic pots with a capacity of

4 kg/pot. Experimental treatments were added with three replications for each treatment. Four 30-day-old eggplant seedlings

(Barcelona variety) were planted in each pot, and the treatments were applied as shown in the table below.

Table 1: the treatment used in the woody canopy experiment

<u>No.</u>	<u>Treatments</u>	<u>No.</u>	<u>Treatments</u>
1	Rs3	8	Fs2 + Pf
2	Rs3 + Pf	9	Fs2 + Pf + Ch
3	Rs3 + Ch	10	Fs2 + Bel
4	Rs3 + Pf + Ch	11	Pf
5	Rs3 + Bel	12	Ch
6	Fs2	13	Pf+ Ch
7	Fs2 + Pf	14	Control

Rs = *R. solani* , Fs = *F. solani* Pf= *Pseudomonas fluorescens* , Ch = chitosan, Bel = Beltanol

The experiment was conducted using a Completely Randomized Design (CRD). The inoculum of the pathogenic fungi *Fusarium solani* (Fs2) and *Rhizoctonia solani* (Rs3) was introduced into the designated treatments by loading it onto millet seeds at a weight-to-weight ratio five days before planting. For the biological control agent *Pseudomonas fluorescens*, a bacterial suspension was applied to the pots at a rate of 10 mL per planting hole. Regarding Chitosan, the planting holes were initially sprayed, followed by a subsequent foliar application at a concentration of 5% one month after planting, ensuring full coverage of the vegetative parts. The chemical fungicide Beltanol was applied as a soil drench at a concentration of 1% one day after inoculating the pathogenic fungi [18]. The results were evaluated three months post-planting by assessing the incidence and severity of root rot disease caused by *R. solani* and *F. solani*.

3 Results and Discussion

1-3 Isolation and diagnosis of fungi associated with the roots of eggplant plants infected with root rot disease

The results of isolation and diagnosis showed the presence of several genera of fungi isolated from the roots of eggplant plants infected with root rot disease, which showed symptoms of the disease represented by brown discoloration of the roots and rotting of part or all of the root, with yellowing and wilting of the plant leaves (Table 2). The most frequently occurring pathogenic fungi was *Fusarium solani*, which was isolated from 7 areas covered by the survey, with an incidence rate of 74.14% and the highest incidence rate of 95%, followed by *Rhizoctonia solani*, which was isolated from 7 areas with an incidence rate of 36.2% and the highest incidence rate of 81%. These results are consistent with many studies that confirmed the association of these fungi with root rot of many crops. The diagnostic results showed the presence of many fungi associated with eggplant roots at a lower frequency, including *Trichoderma* spp., *F. oxysporum*, *Aspergillus niger*, *Penicillium* spp., *Sclerotium seclerotinia*, *Alternaria alternata*, and *Mucor* spp., which appeared at a rate of 12%, 8%, 6%, 25%, 30%, 22%, and 14%, respectively. These results were consistent with what [4] found that these fungi *F. solani* and *R. solani* infect eggplant plants.

and cause root rot diseases and seedling damping off.

Table (2) Percentage of fungi associated with eggplant roots infected with root rot disease

<u>Names of fungi</u>	<u>Rashidiy</u>	<u>Akir</u>	<u>Imam</u>	<u>Muwailh</u>	<u>Al-</u>	<u>Tali'ah</u>	<u>Rashid</u>	<u>Zubaidi</u>	<u>Kish</u>	<u>Abu Sha'ir</u>	<u>Sumoud</u>	<u>Rate (%)</u>	<u>Highest presence rate (%)</u>
<i>Fusarium solalni</i>	60	95	93	93	-	-	-	50	66	-	62	74.14	95
<i>Rhizoctonia solani</i>	25	30	25	-	81	37	25	-	-	31	-	36.2	81
<i>Macrophomina phaseolina</i>	30	25	-	68	31	38	-	10	-	-	-	33.66	68
<i>Trichoderma spp.</i>	8	8	-	-	-	12	-	-	-	-	-	10	12
<i>Fusarium oxysporum</i>	10	8	-	-	10	-	-	-	-	6	-	8.5	10
<i>Aspergillus nigeria</i>	4	4	-	-	6	-	-	6	1	6	-	4.5	6
<i>Pencillium spp.</i>	8	13	-	-	-	-	-	-	25	6	6	11.6	25
<i>Scleroctinia sclerotium</i>	-	30	-	-	-	-	-	-	-	-	-	30	30
<i>Alternaria alternata</i>	10		22	-	-	-	-	12	-	-	--	14.67	22
<i>Mucor spp.</i>	-	14	12	-	-	-	-			11	-	12.33	14

2-3Pathogenicity Test

1-2-3Pathogenicity Test of Pathogenicity of Pathogenic Fungal Isolates Using Cabbage Seeds on PDA Media

The results of Table (3) showed that all tested pathogenic isolates led to a significant reduction in the germination percentage, compared to the comparison treatment. in which the percentage of germinated seeds reached 100%, as the *Fusarium solani* isolate (Fs2) (Akir isolate) outperformed the rest of the isolates in reducing the germination percentage, as the germination percentage rate reached 0.00%. The results also indicated that all tested *Rhizoctonia solani* isolates caused a clear significant reduction in the germination of Cabbage seeds compared to the comparison treatment. The results also indicated that the fungus isolates *Macrophomina phaseolina*

caused a reduction in the germination percentage. The reason for the variation of isolates in their effect on the percentage of germination of cabbage seeds is due to the genetic differences between these isolates, which were collected from different regions, or the difference between these isolates in their ability to produce toxins or enzymes that decompose pectin and cellulose in the early stages of infection. These enzymes play a major role in penetrating the plant host and causing infection, such as Phosphatase, Pectinase, Cellulase, and pectin lyase. This was confirmed by [38]. These results also agree with what [28,23] indicated, that these fungi are important and major causes of root rot disease, and that they are among the most important pathogens of many plant families .

Table (3) Detection of pathogenic isolates associated with the roots of eggplant plants infected using cabbage seeds

No.	Isolate	Site	No. of seed germinated	% Germination
1	Control	-	10.00	100.00
2	Fs1	Rashidiya	5.00	50.00
3	Fs2	Akir	0.00	0.00
4	Fs3	Imam	1.00	10.00
5	Fs4	Muwailha	1.33	13.33
6	Fs5	Zubaidi	4.00	40.00
7	Fs6	Kish	2.33	23.33
8	Fs7	Sumoud	2.00	20.00
9	Rs1	Rashidiya	3.66	36.67
10	Rs2	Akir	2.66	26.67
11	Rs3	Imam	0.00	0.00
12	Rs4	Al-Azzawiya	1.00	10.00
13	Rs5	Tali'ah	1.33	13.33
14	Rs6	Rashid	3.33	33.33
15	Rs7	Abu Sha'ir	2.66	26.67
16	Mp1	Rashidiya	3.66	36.67
17	Mp2	Akir	3.00	30.00
18	Mp3	Muwailha	4.33	43.33
19	Mp4	Al-Azzawiya	5.33	53.33
20	Mp5	Tali'ah	2.66	26.67
21	Mp6	Zubaidi	3.33	33.33
LSD=0.05			1.0170	10.170

Each number represents the average of three replicates, Fs represents the symbol of the fungus *Fusarium solani*, Rs represents the fungus *Rhizoctonia solani* Mp=*Macrophomina phaseolina*, the numbers near the symbol represent the isolate number.

2-2-3 Testing the pathogenicity of the pathogenic fungi *Fusarium solani* and *Rhizoctonia solani* in the germination of eggplant seeds in plastic pots

The results of the pathogenicity in plastic pots, Table (4), showed that the fungal isolates under study were able to infect eggplant seeds, as the germination percentage reached 0.00%

for the isolate Fs2 compared to the comparison treatment, in which the germination percentage reached 100%, while the germination percentage reached 23.3% and 36.7% for the isolates Fs3 and Fs4, respectively. The germination percentage of eggplant seeds reached 0.00% for the isolate Rs3 compared to By treating the fungus alone, which was 100%. The results of this study are consistent with what many studies have reached regarding the emergence of pathogenic soil fungi and their spread on eggplant and their causing root rot on eggplant and other crops, most notably the fungus *Fusarium* sp. And *R.solani* [29,24. [

Table (4) Testing the pathogenicity of pathogenic fungi *Fusarium solani* and *Rhizoctonia solani* in the germination of eggplant seeds in plastic pots

<u>No.</u>	<u>Isolate</u>	<u>No. of seed germinated</u>	<u>% Germination</u>
1	Control	10.00	100.00
2	Fs2	0.00	0.00
3	Fs3	1.00	23.3
4	Fs4	1.33	36.7
5	Rs3	0.00	0.00
6	Rs4	1.00	36.7
7	Rs5	1.33	40.00
0.05=LSD		1.146	11.46

*Each number in the table represents the average of three replicates, Fs = *F. solani*, Rs = *R. solani* and the number near the isolate symbol represents the isolate number

3-3 Testing the antagonistic ability of *Pseudomonas fluorescens* bacteria against the pathogenic fungi *Fusarium solani* and *Rhizoctonia solani* on PDA culture medium . The results showed in Table 5 and Figure 1 the ability of *P. fluorescens* bacteria to inhibit the

growth of the pathogenic fungi *F. solani* and *R. solani* Fs2 and Rs3 isolation on PDA culture medium and *P. fluorescens* bacteria showed its highest effect at concentration 10⁻¹ on the growth of the pathogenic fungi *F. solani* and *R. solani* as the percentage of inhibition reached 100.00%. These results are consistent with the results of a number of studies that confirmed the effectiveness of *P. fluorescens* bacteria in inhibiting the growth of the fungi *F. solani* and *R. solani* [2,6,14.]

Table 5 Testing the antagonistic ability of *Pseudomonas fluorescens* bacteria against the pathogenic fungi *Fusarium solani* and *Rhizoctonia solani* on the PDA culture medium

<u>Treatment</u>	<u>Concentration</u>	<u>Colony diameter.cm</u>	<u>Inhibition (%)</u>
P.f+ Fs2	Control	9.00	0.00
	10 ⁻¹	0.00	100.00
	10 ⁻²	1.33	85.33
	10 ⁻³	2.50	73.00
	10 ⁻⁴	3.00	67.00
	10 ⁻⁵	3.50	62.00
Rs3+ P.f	Control	9.00	0.00
	10 ⁻¹	0.00	100.00
	10 ⁻²	1.00	89.00
	10 ⁻³	2.00	78.00
	10 ⁻⁴	2.50	73.00
	10 ⁻⁵	3.00	67.00
0.05= L.S.D		0.2809	3.089

* Each number in the table represents the average of three replicates, Fs = *F. solani*, Rs = *R. solani* and Pf = *P. fluorescens* bacteria and the number near the isolate symbol represents the isolate number

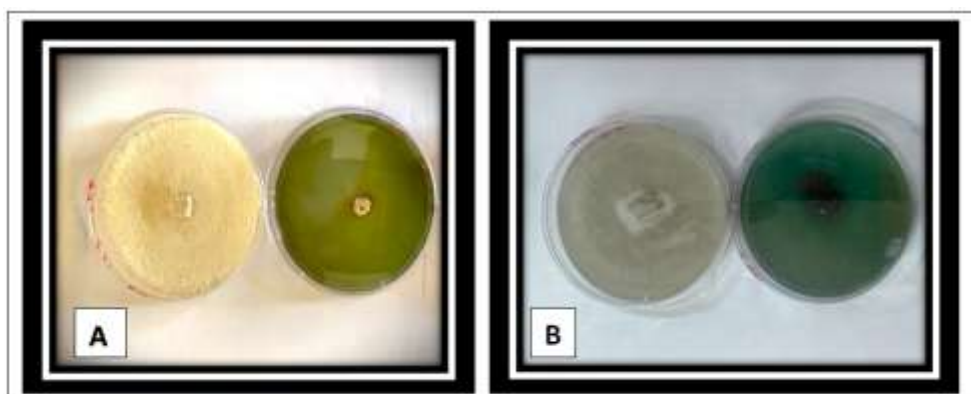


Figure 1 shows the antagonistic ability of *P. fluorescens* bacteria against the two pathogenic fungi A = with the pathogenic fungus *F. solani* B = with the pathogenic fungus *R. solani* on the PDA culture medium.

These results are also consistent with what [37] found regarding the inhibitory effectiveness of *P. fluorescens* bacteria against the growth of the pathogenic fungi *R. solani* and *F. oxysporum* on the PDA culture medium, as it reached The inhibition rate was 80% and 40%, respectively. These results were close to what [30] found using *P. fluorescens* bacteria as a biocontrol agent against the pathogenic fungus *F. solani*, as the bacteria inhibited fungal growth by 70.29% compared to the control treatment, which had an inhibition rate of 0.00%.

3-4 Evaluation of the efficacy of the biological agent *Pseudomonas fluorescens* and chitosan in controlling the pathogenic fungi *Fusarium solani* and *Rhizoctonia solani*, which cause root rot disease in eggplant, and their effects on certain growth parameters under greenhouse conditions.

The results (Table 6) showed that all the factors used in the experiment, including *Pseudomonas fluorescens* and chitosan, whether applied individually or in combination, had a significant effect in reducing the percentage and severity of eggplant root rot disease compared to the pathogenic fungi treatments alone (*R. solani*

and *F. solani*), where the infection rate reached 100% and the disease severity was 80% and 75%, respectively. The integration treatment between *P. fluorescens* and chitosan in the presence of pathogenic fungi showed significant superiority over other treatments in reducing the infection percentage and severity, with infection rates of 16.67% for both, while the disease severity was 5% and 8.33%, respectively. This was followed, with a significant difference, by the treatment of *P. fluorescens* alone in the presence of the pathogenic fungi *R. solani* and *F. solani*. This effect is attributed to its production of cell wall-degrading enzymes that break down the pathogenic fungi, as well as its ability to synthesize various antibiotics and growth regulators, which inhibit many microorganisms and promote plant growth. These include cytokinins, gibberellins, and auxins, which improve the root growth environment, enhance nutrient availability, fix atmospheric nitrogen, and produce HCH compounds that contribute to inhibiting the growth of pathogenic fungi [40] Similarly, the treatment with chitosan in the presence of pathogenic fungi was effective in reducing the infection percentage and severity, reaching 50% for both, with disease severity values of

20% and 21.66%, respectively. The antifungal activity of chitosan is attributed to its ability to increase fungal cell membrane permeability due to the interaction between its positively charged molecules and the negatively charged fungal membrane. Additionally, chitosan inhibits the synthesis of essential proteins and enzymes [39]. It also stimulates plants to produce low-molecular-weight proteins known

as pathogenesis-related proteins, such as chitinase, β -1,3-glucanase, peroxidase, and polyphenol oxidase [17]. These results are consistent with the findings of [35], who reported that adding chitosan to the soil or applying it as a foliar spray at different concentrations significantly reduced the percentage and severity of tomato root rot disease caused by *F. solani* and *R. solani*.

Table (6): Evaluation of the Efficiency of the Biocontrol Agent *Pseudomonas fluorescens* and Chitosan Against the Pathogenic Fungi *Fusarium solani* and *Rhizoctonia solani*, the Causal Agents of Eggplant Root Rot Disease Under Greenhouse Conditions.

<u>NO.</u>	<u>treatments</u>	<u>Disease Incidence %</u>	<u>Severity %</u>
1	Rs3	100	.0075
2	Rs3 + Pf	25.00	15.00
3	Rs3 +Ch	50.00	20.00
4	Rs3 + Pf +Ch	16.67	5.00
5	Rs3 + Bel	0.00	0.00
6	Fs2	100	80.00
7	Fs2 + Pf	25.00	16.66
8	Fs2 + Ch	50.00	21.66
9	Fs2 + Pf + Ch	16.67	8.33
10	Fs2 + Bel	0.00	0.00
11	Pf	0.00	0.00
12	Ch	0.00	0.00
13	Pf+ Ch	0.00	0.00
14	Control	0.00	0.00
LSD=0.05		9.124	4.470

Each number represents the average of three replicates, Rs3 = *Rhizoctonia solani*, Fs2 = *Fusarium solani* Pf= *Pseudomonas fluorescens*, ch= Chitosan, Bel= Beltanol

The results (Table 7) showed that all treatments involving the biocontrol agent *Pseudomonas fluorescens*, chitosan, and the fungicide Beltanol led to an increase in certain growth parameters of eggplant, including plant height, root length, root volume, and the fresh and dry weights of both the shoot and root systems, compared to the treatment with the pathogenic fungi alone (*R. solani* and *F.*

solani), which significantly reduced all the studied growth parameters. The recorded values for the pathogenic fungi treatments alone were 18.53 and 17.17 cm for plant height, 21.66 and 20.33 cm for root length, 3.10 and 3.00 mL for root volume, 11.20 and 10.03 g for fresh shoot weight, 2.50 and 2.00 g for dry shoot weight, 3.53 and 3.36 g for fresh root weight, and 0.33 and 0.21 g for dry root weight, respectively. The combination treatment of *P. fluorescens* and chitosan applied to soil contaminated with the pathogenic fungi significantly outperformed all other treatments in enhancing growth parameters, reaching 35.87 and 34.03 cm for

plant height, 44.10 and 44.00 cm for root length, 8.00 mL for root volume in both fungal treatments, 35.00 and 34.00 g for fresh shoot weight, 6.40 and 6.00 g for dry shoot weight, 8.53 and 8.11 g for fresh root weight, and 1.70 and 1.67 g for dry root weight, respectively, compared to the pathogenic fungi treatments alone. This was followed by the treatment of *P. fluorescens* alone in soil contaminated with *R. solani* and *F. solani*, which also led to an increase in growth parameters, recording 35.06 and 33.93 cm for plant height, 40.00 and 39.00 cm for root length, 7.10 and 7.00 mL for root volume, 32.36 and 31.50 g for fresh shoot weight, 5.80 and 5.60 g for dry shoot weight, 6.94 and 6.70 g for fresh root weight, and 1.32 and 1.31 g for dry root weight, respectively. The efficiency of *Pseudomonas fluorescens* in promoting plant growth is attributed to several mechanisms, including competition with soil microorganisms, production of various antibiotics that enhance plant growth, secretion of siderophores and volatile compounds, and its ability to withstand environmental stresses.

Additionally, this bacterium has the capability to fix atmospheric nitrogen and convert it into ammonia, which becomes available to plants through the production of the enzyme nitrogenase. Furthermore, *P. fluorescens* can solubilize phosphorus, potassium, and other complex minerals in the soil [16,26,36]). The findings of this study align with several previous studies that demonstrated the effectiveness of chitosan in enhancing crop growth parameters. For instance, [3] reported in a study on tomato plants that chitosan application resulted in an increase in plant height and fresh and dry shoot biomass by 1.5–2 times compared to untreated plants. These results are consistent with multiple studies indicating that the interaction between biotic and abiotic factors enhances their role in controlling various plant pathogens by inducing systemic resistance in plants. Additionally, these factors stimulate plant growth by promoting the production of growth regulators and other bioactive compounds [21,1

Table (7): Evaluation of the Efficiency of the Biocontrol Agent *Pseudomonas fluorescens* and Chitosan Against the Pathogenic Fungi *Rhizoctonia solani* and *Fusarium solani* and their Effect on the Growth Parameters of Eggplant Under Greenhouse Condition

NO.	Treatment	Shoot Length	Shoot (g)	Weight	Root Length	Root Volume	Root (g)	Weight
		(cm)	Fresh	Dry	(cm)	(ml)	Fresh	Dry
1	Rs3	18.53	11.20	2.50	21.66	3.10	3.53	0.33
2	Rs3 + Pf	35.06	32.36	5.80	40.00	7.10	6.94	1.32
3	Rs3 + Ch	33.25	31.33	5.23	37.20	6.80	6.25	1.21
4	Rs3 + Pf + Ch	35.87	35.00	6.40	44.10	8.00	8.53	1.70
5	Rs3 + Bel	30.21	30.00	4.00	35.00	5.15	6.00	1.21
6	Fs2	17.17	10.03	2.00	20.33	3.00	3.36	0.21
7	Fs2 + Pf	33.93	31.50	5.60	39.00	7.00	6.70	1.31
8	Fs2 + ch	33.03	31.03	5.00	37.00	6.20	5.15	1.18
9	Fs2 + Pf + Ch	34.03	34.00	6.00	44.00	8.00	8.11	1.67
10	Fs2 + Bel	30.20	30.00	4.00	34.00	5.00	6.00	1.21
11	Pf	40.50	37.00	8.00	46.00	10.33	10.50	2.96
12	Ch	35.07	35.46	7.00	45.00	9.00	9.00	2.16

13	Pf+ Ch	43.07	39.86	9.00	49.00	12.00	10.70	3.34
14	Control	32.20	30.50	4.37	35.40	5.20	6.50	1.51
LSD=0.05		0.894	0.9466	0.8010	0.999	0.894	1.315	0.1325

Each number represents the average of three replicates, Rs3 = *Rhizoctonia solani*, Fs2 = *Fusarium solani* Pf= *Pseudomonas fluorescens* ch=, Chitosan, Bel= Beltanol

References:

- .1 Al-Aamil, Ali Nasser Ali. 2023. The integrated efficacy of the biological control agents *Trichoderma harzianum* and *Pseudomonas fluorescens*, and chelated iron in controlling wilt disease in pepper. Department of Plant Protection / Plant Diseases, College of Agriculture, University of Baghdad.
- .2 Al-Fadhal, F. A., Al-Abedy, A. N., & Alkhafije, D. A. (2019). Isolation and molecular identification of *Rhizoctonia solani* and *Fusarium solani* isolated from cucumber (*Cucumis sativus* L.) and their control feasibility by *Pseudomonas fluorescens* and *Bacillus subtilis*. *Egyptian Journal of Biological Pest Control*, 29, 1-11
- .3 Algam SAE, Xie G, Li B, Yu S, Su T, Larsen J (2010) Effects of *Pae-nibacillus* strains and chitosan on plant growth promotion and control of *ralstonia* wilt in tomato. *J Plant Pathol* 92(3):593–60
- .4 Al-Janabi, Rami Abdul Rahman Abdullah. (2022). Evaluation of the efficiency of some plant extracts and the bio-resistance *Trichoderma harzianum* in controlling eggplant root rot disease. Master's thesis, Al-Musayyab Technical College, Al-Furat Al-Awsat Technical University
- .5 Al-Karawi, Kazem Zghair Khudair (2021). Study of the Fungus *Sclerotinia sclerotiorum* (Lib) Causing White Mold Disease in Eggplant in Some Provinces of Central and Southern Iraq, Identification of Its Strains, and the Effect of Certain Biological Factors in Control and Induction of Systemic Resistance. Ph.D. Dissertation, University of Basrah.
- .6 Alsudani, A. A. (2020). Biocontrol of *Rhizoctonia solani* (Kühn) and *Fusarium solani* (Marti) causing damping-off disease in tomato with *Azotobacter chroococcum* and *Pseudomonas fluorescens*. *Pakistan Journal of Biological Sciences: PJBS*, 23(11), 1456-1461
- .7 Bakker, P.A.H.; P.J. Weisbeek; and B. Schippers.1986. The role of siderophores in plant growth stimulation by *Fluorescent Pseudomonas*. *Med. Fac. Land. Bouww Rijks Univ. Gent*.51/3b (Abstract.(
- .8 Bolkan, H. H., and E. E. Butler .(1974). Studies on heterokaryosis virulence of *Rhizoctonia solani* . *Phytopathology* 64: 513 – 522
- .9 Booth, C. (1971). The Genus *Fusarium*. Commonwealth Mycological Institute, Kew, Surrey, 237.
- .10 Dewan, M. M. (1989). Identify and frequency of occurrence of fungi in roots of wheat and rye grass and their effect on take – all and host growth. Ph. D. Thesis Univ. Wes. Australia. 201
- .11 Donnnelly, J. (2022). Integrated Pest Management Strategic Plan for Oregon.
- .12 Dugan, F.M. (2006). The identification of fungi an illustrated introduction with keys, glossary, and guide to literature. u.s. department of agriculture . agricultural

research service Washington State University Pullman .176 pp.

.13 Elnahal, A. S., El-Saadony, M. T., Saad, A. M., Desoky, E. S. M., El-Tahan, A. M., Rady, M. M., ... and El-Tarabily, K. A. (2022). The use of microbial inoculants for biological control, plant growth promotion, and sustainable agriculture: A review. *European Journal of Plant Pathology*, 1-34

.14 El-Sharkawy, E. E., & Abdelrazik, E. (2022). Biocontrol of *Fusarium* root rot in squash using mycorrhizal fungi and antagonistic microorganisms. *Egyptian Journal of Biological Pest Control*, 32(1), 13.

.15 Feiyan Yu, Yuxuan Chen, Xiaowei Huang, Jiachun Shi, Jianming Xu, Yan He, 2023, Does straw returning affect the root rot disease of crops in soil? A systematic

.16 Gaba, S., Singh, R. N., Abrol, S., Yadav, A. N., Saxena, A. K., & Kaushik, R. (2017). Draft genome sequence of *Halolamina pelagica* CDK2 isolated from natural salterns from Rann of Kutch, Gujarat, India. *Genome Announcements*, 5(6), 10-1128.

.17 Ghule, M. R., Ramteke, P. K., Ramteke, S. D., Kodre, P. S., Langote, A., Gaikwad, A. V., ... & Jambhekar, H. (2021). Impact of chitosan seed treatment of fenugreek for management of root rot disease caused by *Fusarium solani* under in vitro and in vivo conditions. *3 Biotech*, 11, 1-12.

.18 Hasson, Ibrahim Khalil (2005). Biological and Chemical Control of the Causal Agent of Potato Stem Canker, *Rhizoctonia solani* Kühn. Ph.D. Dissertation, College of Agriculture, University of Baghdad, Iraq.

.19 Holliday, P.; and E. Punithalingam. (1970). *Macrophomina phaseolina*. C.M. Descriptions of pathogenic fungi and bacteria 275 commonwealth mycological Institute Kew Surrey England

.20 Hussein, S. and K. Juber. (2014). First report of identification *Fusarium solani* f.sp. *cucurbitae* race 1 and 2 the causal agent of crown and root rot disease of watermelon in Iraq. *International Journal of Agriculture Innovations and Research*, 3: 2319-1473

.21 Jumaa, Ahmad Mohammed Salman. 2022. Evaluation of the efficacy of some plant extracts and propolis in controlling the fungus *Fusarium solani*, which causes root rot and basal stem rot in pepper. Master's Thesis. Department of Plant Protection / Plant Diseases, College of Agriculture, University of Anbar.

.22 Makhlof, B. S. I., Khalil, S. R. A. E., and Saady, H. S. (2022). Efficacy of Humic Acids and Chitosan for Enhancing Yield and Sugar Quality of Sugar Beet Under Moderate and Severe Drought. *Journal of Soil Science and Plant Nutrition*, 1-16.

.23 Matloob, A. A. H. K. F. Alwan and S. H. Segar. (2019). Control of the causes of the damping off disease on pepper by some biological agents in Babylon province-Iraq. *Journal of Biopesticides*, 12 (2):215-223.

.24 Mishra, P. K. (2017). Study on *Macrophomina phaseolina* (Tassi) Goid causing charcoal rot of soybean *Glycine max* (L.) Merrill and its management. Ph.D. Thesis. Indira Gandhi krishi Vishwavidyalaya, Raipur. 216pp

.25 Montealeger, J. R., R. Rodrigo, P. M. Luz, H. Rodrigo, S. Polyana and B. Ximena. (2003). Selection of bioantagonistic bacteria to be used in biological control of *Rhizoctonia solani* in tomato. *J. Biotech.* 6:115-127.

.26 Morris, C. E., Lamichhane, J. R., Nikolić, I., Stanković, S., & Moury, B. (2019). The overlapping continuum of host range among strains in the *Pseudomonas syringae* complex. *Phytopathology Research*, 1, 1-16.

- .27 Muhammad, S., and A. Amusa. 2003. In-vitro inhibition of growth of some seedling blight inducing Pathogens by compost-inhabiting microbes. *Journal of Biotechnology* 2: 161 –164.
- .28 Mulan, Younis Yousef, Alaa Saleh El-Din and Saleh El-Din El-Hussein Mohamed. (2006). Natural chemical and biological control of pathogens that cause wilting and death of seedlings of some vegetable crops in greenhouses in the Riyadh region. King Saud University.
- .29 Mwaniki, P. K. M. M. Abang, I. N. Wagara, J. N. Wolukau and S. Hans-Josef. (2015). Response of African eggplants to *Fusarium* spp. and identification of sources of resistance. *Afr. J. Biotechnol.* 15(11), pp. 390-400.
- .30 NAIN, Y., CHAWLA, N., MEENA, P. K., & SHEKHAWAT, D. S. .2023 BIOCONTROL OF PEA ROOT ROT INCITED BY *FUSARIUM SOLANI* F. SP. *PISI*.
- .31 Nedved, E. L., Kalatskaja, J. N., Ovchinnikov, I. A., Rybinskaya, E. I., Kraskouski, A. N., Nikalaichuk, V. V., ... and Laman, N. A. (2022). Growth Parameters and Antioxidant Activity in Cucumber Seedlings with the Application of Chitosan and Hydroxycinnamic Acids Conjugates under Salt Stress. *Applied Biochemistry and Microbiology*, 58(1), 69-76.
- .32 O'Brien, P. A. (2017). Biological control of plant diseases. *Australasian Plant Pathology*, 46(4), 293-304
- .33 Parameter, J. R.; Whitney, H. S. (1970). Taxonomy and nomenclature of the imperfect state. In: *Rhizoctonia solani* Biology and Pathology. Parmeter, J. R. (Editor). University of California Press, California
- .34 Sadeghi, M. S. S. A. A. Behjatnia, M. Masumi and K. Izadpanah (2008). Characterization of a strain of potato virus Y causing eggplant mosaic in southern Iran. *Austral. Plant Pathol.* 37 (1): 79-86.
- .35 Satar, Shahla Abd Al Kareem. (2022). Effectiveness of some normal and nano plant resistance inducer and effective microorganism preparations against some causes of tomato root rot.. Master's thesis. Al-Furat Al-Awsat University, Al-Mussaib Technical College.
- .36 Singh, P., Singh, R. K., Li, H. B., Guo, D. J., Sharma, A., Verma, K. K., ... & Li, Y. R. (2024). Nitrogen fixation and phytohormone stimulation of sugarcane plant through plant growth promoting diazotrophic *Pseudomonas*. *Biotechnology and Genetic Engineering Reviews*, 40(1), 15-35
- .37 Singh, S., Balodi, R., Meena, P. N., & Singhal, S. (2021). Biocontrol activity of *Trichoderma harzianum*, *Bacillus subtilis* and *Pseudomonas fluorescens* against *Meloidogyne incognita*, *Fusarium oxysporum* and *Rhizoctonia solani*. *Indian Phytopathology*, 74(3), 703-714.
- .38 Wrather, J. A.; Anderson, T. R., Arsyad, D. M., Gai, J., Ploper, L. D.; Porta-Puglia, A.; Ram, H. H. and Yorinori, J. T. (1997). Soybean disease lossestimates for the top 10 soybean producing countries in 1994. *Plant Dis.* 81: 107-110
- .39 Younes, I., & Rinaudo, M. (2015). Chitin and chitosan preparation from marine sources. Structure, properties and applications. *Marine drugs*, 13(3), 1133-1174.
- .40 Zafar, M., Ahmed, S., Munir, M. K., Zafar, N., Saqib, M., Sarwar, M. A., ... & Gulnaz, A. (2023). Application of Zinc, Iron and Boron enhances productivity and grain biofortification of Mungbean.