Anti-bacterial activity of Syzygium cumini (L.) Skeels leaves extract.

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Abstract

It has been determined that *Syzygium cumini L.*, also known as jamun and a member of the Myrtaceae family, has anticancer properties. Bacterial infection a significant problem around the globe. Therefore, the primary objective of this work is to use S. cumini leaf extract as antibacterial agent. Fresh plant samples of the *S. cumini* species were collected Al-Zafaraniyah - Botanical Garden. Methanol was used for extraction. The effectiveness of the extract was evaluated as antibacterial agent by using agar well diffusion with different concentration of extract against two Gram negative bacteria (*Escherichia coli*, *Pseudomonas aeruginos*a) and two Gram positive (*Streptococcus pneumonia*, *Staphylococcus aureus*). The zones of inhibition were measured by using a caliper micrometer against the back of the petri plates . Leaf extract significantly exhibit the most potent as antibacterial with Inhibition zone diameter range 22.1mm -11.5mm against Gram positive and 21.8mm- 12.4mm against Gram negative. The minimum inhibitory concentrations (MICs) of extract were determined. The MIC for Gram positive bacteria was 104 μ g/ml while the MIC for Gram negative bacteria was 208 μ g/ml. The synergistic effect of *S. cumini* leaves extract was investigated with six antibiotics against the higher resistance isolates which appeared increase the antibiotic effect .

Key words : Syzygium cumini , Myrtacea , Antibacterial activity .

Introduction

There are around 150 genera and 3,600 species in the Myrtaceae plant family, which is found in the tropics and subtropics. (1). Jamun, also referred to as *Syzygium cumini* L., is a significant and widely cultivated member of this family (2). For many years, South Asian traditional medicine has employed the fruits, leaves, and seeds of S. cumini to treat diabetes mellitus. The leaves are employed as a diuretic and astringent. (3). They have antioxidant, hypolipidemic, anti-inflammatory, antipyretic, psychopharmacological, and hypoglycaemic effects. (4). *S. cumini's* fruits, stems, and

leaves have all been researched for their antibacterial properties,

The antimicrobial activity of the Syzygium *cumini* leaves hydroalcoholic extract may be tannins and phenolic due to other constituents. Syzygium cumini is known to be very rich in gallic and ellagic acid polyphenol derivatives. Syzygium species have been reported to possess antibacterial activity (5). Aqueous and methanol extracts of Syzygium species have been shown to inhibit the growth of some fungal microorganism implicated in skin diseases, such as Candida albicans, and Trichophyton rubrum and Staphylococcus aureus (6).

Bacteria are microorganisms that are simple in structure and vary in shape and size. They are single-celled and considered prokaryotic. Bacteria are found in all different environments in the air, water, and soil, as they play an important role in life cycles on the surface of the Earth. Pathogenic bacteria differ among themselves in the extent of their virulence according to the virulence factors they possess and their ability to bypass the host's defenses, as well as their ability to resist antibiotics.

Among the most important genera of pathogenic bacteria that are resistant to many antibiotics *are E. coli, S. aureus, P. aeruginosa*, and others (7).

the objectives of the present study to evaluate the *in vitro* antibacterial properties of *S. cumini* against Gram positive and Gram negative bacterial.

Material and Methods

1: Collect plant samples

Fresh plant samples of the S. cumini species were collected for a period from 12/7/2023 until 8/1/2024, from Baghdad Governorate (Al-Zafaraniyah - Botanical Garden). Samples were placed in cardboard boxes Write on it the name of the plant, the place of collection, and the date of collection. Then the samples were washed with water Regular twice, then with filtered water once, then placed on paper to dry for (3-5) Days in the shade and at room temperature with constant stirring to prevent any rotting of parts The plant was obtained to obtain dry samples, which were then ground using an electric grinder (blender To obtain dry plant powder, then it was collected in clean glass containers until use (8).

2: preparation of Plant extraction

The plant extract was prepared based on the method [9] with some minor modifications. The solvent was used Aqueous-Alcoholic (Methanol) 50% for the purpose of preparing plant extract.

The extraction process was done as follows: (10).

- Dry leaves of the *S.cumini* were ground using an electric grinder to obtain Powder.
- Place 80 gm. of leaves powder in a 1 liter volumetric flask in a mixture consisting of (water, methanol (400/400 ml)) and close the opening of the container tightly with aluminum foil to avoid evaporation of the mixture and air oxidation.
- The flasks were placed in a water bath at 37 C° for 2 hour at high speed.
- The extracts were filtered in two stages, the first using medical gauze and the second using a kit Layers of Whatman type (1) filter paper.
- Place the filtrate in the centrifuge at 2500 rpm for 10 minutes.
- Place the filter in a clean glass dish and then place it in the Oven at 40 C° for 2-5 days. To be dried well.
- Scrape the extract after it dries completely, then store it in clean containers until use.

3: Collect pathogenic bacteria

Four pathogenic bacteria, 2 gram-positive and 2 gram-negative, were collected from microbiology laboratories at the University of Babylon\ College of Science\ Department of Biology. The diagnosis has been confirmed by VITEK-2 Compact System.

4: Identification of bacteria by VITEK-2 Compact System

Suspension of bacteria was prepared according to the manufacturer's instructions. An adequate number of colonies was obtained by transferring an overnight pure culture and suspending it in 3.0 ml of sterile saline in (polystyrene) test tube. Adjustment of turbidity to 0.5 McFarland was made. Employing a Densi-Chek turbidity meter. Finally, the vitek-2 chamber with the specimen suspension tubes was loaded with the Gram negative -ID, and Gram positive -ID cassette (10).

5: Inocula Preparation (Turbidity Standard)

To prepare the inocula , colonies from overnight culture of bacteria isolates were transferred to 5 ml tube of normal saline to obtain culture with 1.5×108 CFU/ml by adjusting to 0.5 McFarland standard (11).

6: Antibiotic Susceptibility Test (AST)

Lists of antibiotic susceptibility testing were created using documents and breakpoints from the Clinical Laboratory Standards Institute (CLSI, 2020) the European Committee on Antibacterial Susceptibility Testing (EUCAST) and the United States Food and Drug Administration (FDA). This study included 10 antibiotics was determined by discdiffusion method, which are Azithromycin , Nitrofurantoin, Imipenem, Amikacin, Nalidixic acid Ciprofloxacin . Doxycycline , Amoxicillin / calvulanic acid, Trimethoprime / sulphamethoxazole

, Ampicillin against (*E.coli*, *p. aeruginosa*, *St. pneumonia*, *S. aureus*). (12).

7: Antibacterial activity of leaves extract

This method was done on Muller Hinton agar (13).

- Turbidity of each bacterial isolates compared to McFarland 0.5 standard to get the right concentration for each of them .
- A 0.1 ml of each bacterial isolates were added to petri dish containing Muller Hinton agar and spread by spreader and left the dishes for an 1 hr.
- Wells were made by using cork borer (2 mm diameter) as it was equal distance between the well and the other.
- The extract were dissolved to get various concentrations (1000 as stock and by dilution got the three concentration(500, 250, 125) mg/ ml.
- A 40 microliter of each concentration were added to each well and incubated in the incubator at 37 ° C for 24 hr . inhibition zones were measured by a ruler .

8: Minimum Inhibitory Concentrations (MICs) Determination:

The minimum inhibitory concentrations (MICs) were determined by a serial dilution technique. MICs were defined as the lowest concentration of an antimicrobial that inhibits growth of a microorganism after their incubation for overnight (14) . The MIC for Gram positive bacteria was 104 μ g /ml while the MIC for Gram negative bacteria was 208 μ g /ml . as showed in table (4).

9: Synergistic effect

The antagonistic activity of antibiotic and extract combination were determined by the modified disc diffusion method according to the NCCLS guidelines.

The MHA plates were seeded with the above antibiotic disc impregnated with exract (104 μ g/ml) for (*St.pneumonia* and *S. aureus*) and (208 μ g/ml) for (*P. aeruginosa and E.coli*) along with plain antibiotic disc taking as positive control, the MHA plates were kept at 4C° for 1 hr to allow the proper diffusion, after that kept at 37 C° for 24 hr. The zones of inhibition were measured by using a caliper micrometer against the back of the petri plates (15].

Results and Discussion 1: Morphological characteristics of

Syzygium cumini (L.) Skeels .

The species *S. cumini* is a large evergreen tropical tree, 25-30 meters high, with thick, brown bark. One of the fast growing trees. The leaves of the plant are simple, opposite . The leaf blade is broadly elliptic and may be oblong-ovate, or elliptical in shape , with a length of 6-12 cm. The leaf is smooth and

sometimes somewhat leathery. The leaf petiole is 3 cm long. The flowers are white pink or greenish-white, about 1 cm. They are arranged in branched groups at the ends of the stem. The calyx leaves are funnelshaped or cup-like, (4 mm) long, with 4 lobes (teeth). Petals: small fused petals that fall together as a small disk, white or light purple. Stamens are multiple and longer than the calyx leaves. Fruits are oval in shape, 1.5-3.5 cm long, dark purple or almost black, and dark red. The fruit contains a single seed approximately 2 cm long. . (15).

Taxonomic Tree : (16)

Kingdom: Plantae Phylum : Spermatophyta Subphylum : Angiospermae Class : Dicotyledonae Order : Myrtales Family : Myrtaceae Genus : Syzygium Species : Syzygium cumini

2 : Bacterial isolates according to Vitek 2 Advanced Expert System

In VITEK2 Advanced Expert System system (bioMerieux, Marcy l'Etoile. France), the isolates were identified as S. aureus with probability of (91) %, E.coli 99%, St. pneumonia 99% and P.aeroginosa with probability of 89% . table (1). Biological characteristics of all bacterial isolates were done also by VITEK2 compact system, figure 1. These results are similar to (16) who found that S. aureus was the most common pathogens ,followed by Р. aeruginosa.

morphology and Vitek 2 system.						
Bacterial species	No. of isolates	Probability in Vitek 2 system				
Streptococcus pneumonia	15	99 %				
Staphylococcus aureus	20	91 %				
Escherichia coli	10	99 %				
Pseudomonas aeruginosa	10	89 %				
TOTAL	55					

Table (1): Identification of pathogenic bacteria depended on the colonial morphology and Vitek 2 system .

C.t					A	Analysis Time: 91% Probability Bionumber:			1.91 hours							-	
Selected Organism ID Analysis Messages				9 8	3.93 hours Status Staphylococcus aureus 250402066761111				t Final		inal	7					
Bi	orhand an				_	_	_	_		11-1				-	_		-
2	ochenical I	Detail	5					-	-	-	_	_		-	-		-
	AMY		4	PIPLC	+	5	dXYI	T	Te.	Lance	-	-					
20	APPA	+	14	CDEX		15	AunA	÷	8	ADHI	+	9	BGAL.		11	AGLU	
20	LeuA		23	ProA		24	BGUR	÷	16	BGAR		17	AMAN		19	PHOS	•
-8	AlaA		29	TyrA		30	dSOR	+-	25	AGAL	-	26	PyrA	+	27	BGUR	E
38	dRIB	-	39	ILATK		42	LAC	-	31	URE		32	POLYB	+	37	JGAL	Ŀ
47	NOVO	-	50	NC6.5	+	52	duan	÷	44	NAG	+	45	dMAL.	+	46	BACI	
57	dRAF	+	58	0129R		100	SAL	·	53	dMNE.	+	54	MBdG	+	56	PUL	Ŀ
54	OPTO	+			-	100	5/11.	+	60	SAC	+	62	JTRE	+	63	ADH2s	Ŀ
iele	cted Organi	ism				99% Bio	Probabili number:	ty		Escherich 04056105	nia col 50524	li 1610					
D 4	nalysis Me	ssages	;														
U P																	
Bio	chemical De	tails															
Bio	hemical De	tails	3	ADO	- 1	4	РугА	-	5	IARL	-	7	dCEL	-	9	BGAL	+
Bio	chemical De APPA H2S	tails - -	3 11	ADO BNAG	-	4	PyrA AGLTp	-	5 13	IARL dGLU	-+	7	dCEL GGT	-	9 15	BGAL OFF	+++
Bior 0 7	APPA H2S BGLU	tails - -	3 11 18	ADO BNAG dMAL	- - +	4 12 19	PytA AGLTp dMAN	- - +	5 13 20	IARL dGLU dMNE	- + +	7 14 21	dCEL GGT BXYL	-	9 15 22	BGAL OFF BAlap	+++
Bio4	hemical Do APPA H2S BGLU ProA	- - - -	3 11 18 26	ADO BNAG dMAL LIP	- - +	4 12 19 27	PyrA AGLTp dMAN PLE	- - +	5 13 20 29	IARL dGLU dMNE TyrA	• + +	7 14 21 31	dCEL GGT BXYL URE	-	9 15 22 32	BGAL OFF BAlap dSOR	+++++++++++++++++++++++++++++++++++++++
Bio:	chemical De APPA H2S BGLU ProA SAC	etails - - - - +	3 11 18 26 34	ADO BNAG dMAL LIP dTAG	- - + -	4 12 19 27 35	PyrA AGLTp dMAN PLE dTRE	• • • •	5 13 20 29 36	IARL dGLU dMNE TyrA CIT	• + + +	7 14 21 31 37	dCEL GGT BXYL URE MNT	-	9 15 22 32 39	BGAL OFF BAlap dSOR 5KG	+++++++++++++++++++++++++++++++++++++++
Bio 0 7 3 0	hemical Do APPA H2S BGLU ProA SAC ILATk	- - - + +	3 11 18 26 34 41	ADO BNAG dMAL LIP dTAG AGLU	- - + -	4 12 19 27 35 42	PyrA AGLTp dMAN PLE dTRE SUCT	- - + + +	5 13 20 29 36 43	IARL dGLU dMNE TyrA CIT NAGA	• + + +	7 14 21 31 37 44	dCEL GGT BXYL URE MNT AGAL	- - -	9 15 22 32 39 45	BGAL OFF BAlap dSOR 5KG PHOS	+ + + + + + + + + + + + + + + + + + + +
Bio 0 7 3 3 0 6	APPA H2S BGLU ProA SAC ILATk GlyA	- - - + +	3 11 18 26 34 41 47	ADO BNAG dMAL LIP dTAG AGLU ODC	- - - -	4 12 19 27 35 42 48	PyrA AGLTp dMAN PLE dTRE SUCT LDC	• + • + +	5 13 20 29 36 43 53	IARL dGLU dMNE TyrA CIT NAGA IHISa	• + + •	7 14 21 31 37 44 56	dCEL GGT BXYL URE MNT AGAL CMT	- - + +	9 15 22 32 39 45 57	BGAL OFF BAlap dSOR 5KG PHOS BGUR	+ + + + + + + + + + + + + + + + + + + +

Figure (1): Picture clear biochemical characteristics of all bacterial isolates were done also by VITEK2 compact system.

3 : Antibiotic Susceptibility Test (AST)

This study included 10 antibiotics was determined by disc-diffusion method, which are Azithromycin , Nitrofurantoin , Imipenem , Amikacin , Nalidixic acid , Ciprofloxacin, Doxycycline , Amoxicillin/calvulanic acid, Trimethoprime / sulphamethoxazole , Ampicillin against (*E.coli* , *p.aeruginosa* , *St.pneumonia* , *S. aureus*). The pathogenic bacterial isolates used for the study showed high resistance to most of the antibiotics used, as shown in Table 2 .

Antibiotic	Antibiotic susceptibility of pathogenic bacteria (%)							
	E.coli	P.aeruginosa	St.pneumonia	S. aureus				
AZM	R	S	S	R				
AK	R	Ι	S	S				
NA	Ι	R	S	S				
CIP	R	Ι	R	R				
F	R	S	R	R				
IPM	S	Ι	S	S				
DO	Ι	R	R	S				
SXT	Ι	R	R	Ι				
AMC	Ι	R	Ι	S				
AM	R	R	S	R				

 Table (2): Antibiotic Susceptibility Test for pathogenic bacteria.

The emergence of antibiotics resistance in the majority of pathogenic bacterial strains is a cause of utmost concern in infectious bacterial diseases. Therefore, there is an inevitable need to identify the effective antibacterial agents which are more effective against microbial ailments with minimal side effects on host cells (17).

the phenotypic prevalence of resistance to Azithromycin and Ampicillin was reported in more than 30% of cases, which is which matches perfectly with the results of this study (18) ,while it was not consistent with the study of (18). in Kuwait , and (19) . in Qom, and (20) in Tabriz ,who demonstrated that highest antibiotic resistance was found in Nalidixic acid, which included .

4 : Antibacterial activity of *S. cumini* leaves extract

In well diffusion method various concentrations (2000, 1000 ,500 mg\ml) from methanol leaves extract were test against 4 pathogenic bacteria . It was found that at concentration 500 mg\ml gave the highest inhibition effectiveness and all concentrations were significant at P-value 0.05 as showed in table 4 and figure 2.

leaves extract may bind into proteins, thereby inhibiting the assembly of proteins. This, in turn, suppresses the formation of inhibition zone lead to inhibit cytokinesis and bacterial proliferation[]. The results shown in Table (3) show that the concentration of 2000 mg/ml) has the highest effect on pathogenic bacteria, and all concentrations were significant at P-value 0.05. This suggests that the observed differences in bacterial growth inhibition between different extract concentrations were statistically significant.

Concen. mg\ml	Inhibition zone diameter mm.					
Bacteria type	2000	1000	500	C+	C-	
S. aureus	22.1±2.5 b	20.6±2.9 b	18.3±1.1 b	17.7±4.4 b	0±0 a	
St.pneumonia	21.1±3.8 c	20.8±1.8 c	11.5±0.9 b	10.4±2.2 b	0±0 a	
E. coli	21.8±1.8 c	21.4±0.9 c	15.7±1.9 b	15.4±2.5 b	0±0 a	
P. aeruginosa	20.4±1.9 d	20.6±1.3 d	12.4±1.4 b	15.4±1.5 c	0±0 a	

 Table (3): Antibacterial effect of methanol S. cumini leaves extract.

Different letters refer to significant difference at p≤0.05.

One study published in the Journal of Microbiology and Biotechnology in 2017 evaluated the antibacterial effects of *S. cumini* leaves extract against clinical isolates of *S. aureus* and *P.aeruginosa* (21). The results demonstrated that *S. cumini* leaves extract exhibited significant inhibitory activity against the tested strains. Another study published in the Journal of Natural Products in 2019 found *S. cumini* leaves extract was effective in inhibiting the growth of *S. aureus* biofilms, which are communities of bacteria that can be particularly resistant to antibiotics (22). It is believed that *S. cumini* leaves extract can disrupt the bacterial cell membrane, interfere with DNA replication and protein synthesis, and inhibit enzymes necessary for bacterial survival (23).



Figure 2: Antibacterial effect of *S.cumini* . leaves extract on some pathogenic bacterial strains..

5 : Minimum Inhibitory Concentrations Determination

The minimum inhibitory concentrations (MICs) were determined by a serial dilution technique. MICs were defined as the lowest

concentration of an antimicrobial that inhibits growth of a microorganism after their incubation for overnight. The MIC for Gram Negative bacteria was 208 μ g/ml. The MIC for Gram positive bacteria was 104 μ g/ml, as showed in Table 4.

Test organisms	Minimum Inhibition
	Concentration (MIC)
	(µg/ml)
E. coli	208
P. aeruginosa	208
S. aureus	104
St.pneumonia	104

Table (4): Minimum Inhibitory Concentrations and (MICs) of S. cumini leaves extract.

By measuring the minimum inhibitory concentration of the extract against the pathogenic bacteria, it was found that the Gram-positive bacteria were more sensitive to the extract, as they require the inhibition of the positive bacteria to $104 \mu g/ml$, while they require the inhibition of the negative bacteria was 208 µg/ml. The difference between the sensitivity of Gram-positive and Gram-negative bacteria may be due to the difference between the bacteria in terms of wall structure and the presence of the peptidoglycan layer, in addition to the nature of Gram-negative bacteria and their containment of a high percentage of fats, which delays the absorption and assimilation of substances.

6 : Synergistic effect of *S. cumini* leaves extract with antibiotic .

The synergistic effect of *S. cumini* leaves extract was investigated with six antibiotics

against the higher resistance isolates which appeared increase the antibiotic effect as showed in figure (3). The MHA plates were seeded with the above antibiotic disc impregnated with exract (104 µg /ml) for (*St.pneumonia* and *S. aureus*) and (208 µg /ml) for (*P. aeruginosa and E.coli*) along with plain antibiotic disc taking as positive control , the MHA plates were kept at 4C° for 1 hr to allow the proper diffusion , after that kept at 37 C° for 24 hr . The zones of inhibition were measured by using a caliper micrometer against the back of the petri plates (24).

S. cumini leaves extract increased the effectiveness of antibiotics and increased the efficiency of their work by causing modifications in the structure of the antibiotic to make it more effective.



Figure 3 : The synergistic effect of *S. cumini* leaves extract was investigated with six antibiotics against the higher resistance isolates

Conclusion

The Aqueous-alcoholic extract.of the leaves of *S. cumini* is of great importance as an antidote to pathogenic bacteria, both positive and negative, of the Gram stain. The use of medicinal plants can contribute to solving the problem of antibiotic resistance.

References

1- Thampi N, Shalini JV. Bioprospecting the *in-vitro* antioxidant and anti-cancer activities of silver nanoparticles synthesized from the leaves of *Syzygium* samarangense. Int J Pharm Pharm Sci. 2015;7:269–74.

- 2- Fiqri H, Gaisani A, Adrebi K, Proborini W, Widyanto R. Evaluation of total phenolic, total flavonoid, and *in vitro* cytotoxic activity of *Syzygium cumini* extract in cervical cancer cell. *J Phys Conf Ser.* 2020;1665:12033. 10.1088/1742-6596/1665/1/012033 [CrossRef] [Google Scholar]
- 3- Khandaker MM, Mat N, Abdulrahman MD, Ali MA. Morphological and anatomical studies of Syzygium polyanthum (Wight) Walp. (Myrtaceae). MNJ. 2018;70:309–22.

- 4- Abdulrahman MD. Review of ethnopharmacology, morphoanatomy, biological evaluation and chemical composition of *Syzygium polyanthum* (Wight) Walp. *Plant Sci Today*. 2021;9:167–77.
- 5- Abdulrahman MD, Hasan Nudin N, Khandaker MM, Ali AM, Mat N. In vitro biological investigations on Syzygium polyanthum cultivars. Int J Agric Biol. 2019;22:1399–406.
- 6- Abdulrahman M. Antioxidant, alpha glucosidase and antibacterial evaluation of *Syzygium mytifolium* (Roxb.) Walp. *Plant Sci Today*. 2021;8:410–5. 10.14719/pst.2021.8.2.1113
- 7- Chua LK, Lim CL, Ling APK, Chye SM, Koh RY. Anticancer potential of Syzygium species: a review. Plant Foods Hum Nutr. 2019;74:18–27. 10.1007/s11130-018-0704-z
- 8- Abdulrahman MD, Fatihah HNN, Abdul M, Ali M, Mat N, Khandaker M. Phenetic and unsupervised multivariate analysis Syzygium polyanthum (Wight) Walp. Iraqi JAS. 2021;52:249–58. 10.36103/ijas.v52i1.1255
- 9- Jena S, Ray A, Sahoo A, Das PK, Dash KT, Kar SK, et al. Chemical composition and biological activities of leaf essential oil of Syzygium cumini (L.) from eastern India. J Essent Oil Bear Pl. 2021;24:582– 95.

10.1080/0972060X.2021.1947897

- 10- Ayyanar M, Subash-Babu P. *Syzygium cumini* (L.) Skeels: a review of its phytochemical constituents and traditional uses. *Asian Pac J Trop Biomed*. 2012;2:240–6. 10.1016/S2221-1691(12)60050-1
- 11-Kristanti AN, Aung EE, Aminah NS, Takaya Y, Aung HT,

RamadhanR. BioactivetriterpenoidsfromIndonesianmedicinalplant Syzygiumcumini (L.). OpenChem.2022;20:204–11.10.1515/chem-2022-01382022-0138

- 12- Aung E, Kristanti A, Aminah N, Takaya Y, Ramadhan R, Aung H. Anticancer activity of isolated compounds from *Syzygium cumini* (L.). stem bark. *Rasayan J Chem.* 2021;14:312–8. 10.31788/RJC.2021.1416106
- 13- Rocchetti G, Lucini L, Ahmed SR, Saber FR. *In vitro* cytotoxic activity of six *Syzygium* leaf extracts as related to their phenolic profiles: an untargeted UHPLC-QTOF-MS approach. *Food Res Int.* 2019;126:108715.
 10.1016/j.foodres.2019.108715
- 14-Aung , E.E. ; Kristanti , A.N. ; Aminah , N.S. ; Takaya, Y. ;and Ramadhan , R. (2020) . Plant description, phytochemical constituents and bioactivities of *Syzygium* genus: A review . De Gruyter . Open Chemistry ; 18: 1256–1281 .
- 15- Ayyanar, M. and Subash-Babu, P. (2012). *Syzygium cumini* (L.) Skeels: A review of its phytochemical constituents and traditional uses. Asian Pac J Trop Biomed. 2(3): 240–246. doi: 10.1016/S2221-1691(12)60050-1.
- 16-Townsend, C.C. and Guest, E. (1974), Flora of Iraq, Vol. 3. Ministry of Agriculture and Agrarian reform, Baghdad, Iraqi.
- 17-Ruan, Z.P.; Zhang, L.L. and Lin, Y.M. (2008) . Evaluation of the antioxidant activity of *Syzygium cumini leaves*. Molecules. ;

13(10):2545–56.

https://doi.org/10.3390/molecules13 102545 PMID: 18927517.

- 18-Simões-Piresa, C.A.; Vargasa, S.; Marstona, A.; Ioseta, J.R.; Paulo, M. Q. An Matheeussend and Maesd, L. (2009). Ellagic Acid Derivatives from Syzygium cumini Stem Bark: Investigation of their Antiplasmodial Activity. Natural Product Communications Vol. 4 (10) No. 10.1371 – 1376.
- 19-Sarma, N.; Begum, T.; Pandey, S.K.; Gogoi, R.; Munda, S. and Mohan Lal, M. (2020). Chemical Composition of Syzygium cumini (L.) Skeels Leaf Essential Oil with Respect to its Uses from North East Region of India . Journal of Essential Oil Bearing Plants . DOI: 10.1080/0972060X.2020.1796822.
- 20-Das, G.; Nath, R.; Talukdar, A.D. ; `gagündüz, D.A.; Yilmaz, B.; Capasso, R.; Shin, H.S. and Patra, J.K. (2023). Review Major Bioactive Compounds from Java Plum Seeds: An Investigation of Its Extraction Procedures and Clinical Effects. Plants, 12, 1214. https://doi.org/10.3390/plants12061

25-

214

https://www.mdpi.com/journal/plant s.

- 21-Lini J.J. and Anju S.B. (2021). Phytochemical analysis, nutritional profile and antibacterial studies of *Syzygium cumini* seeds . Journal of Emerging Technologies and Innovative Research (JETIR) . Volume 8, Issue 2 . 75-92. www.jetir.org (ISSN-2349-5162) .
- 22-Selvaganesh C. and Sadhana B. (2022). Phytophenolics and functional group analysis of *Syzygium cumini* L. seeds for antibacterial activity. International Journal of Advanced Research in Biological Sciences. 9(6): 132-140.
- 23- Chattopadhyay D, Sinha BK, Vaid LK (1998) . Antibacterial activity of *Syzygium* species. *Fitoterapia* 69: 356–367.
- 24- Chandrasekaran M, venkatesalu V (2004) Antibacterial and antifungal activity of Syzygium jambolanum seeds. J Ethnopharmacol 91(1): 105–108.