

**BIOLOGICAL CONTROL AND INDUCED SYSTEMIC
RESISTANCE IN CHICKPEA *Cicer arietinum* L. AGAINST
Fusarium oxysporum F.SP. *ciceri***

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ABSTRACT

Bio-control agents, Bio Root Care (BRC), *Trichoderma harzianum*, *Pseudomonas fluorescens*, and Non-pathogenic *Fusarium oxysporum* (N_{fo}) showed clear inhibition of *Fusarium oxysporum* f.sp.ciceri (*Foc*) growth on artificial medium. *T.harzianum* significantly ($p = 0.05$) scored the highest rate of growth inhibition, 59.7% compared with BRC, N_{fo}, *P.fluorescens*, *R.leguminisarum* treatments. Seed or soil treatments with local *T.harzianum* and *P.fluorescens* showed clear ability to control chickpea wilt disease by significantly reducing the disease incidence compared with the commercial, *T.harzianum* and *P.fluorescens* products and with control treatment. While seed and soil treatments with the local *T.harzianum* and *P.fluorescens* caused 7 and 9 disease incidence, 7 and 12% respectively. The commercial products, *T.harzianum* and *P.fluorescens* caused 16 and 19,18, 19% respectively. Systemic induced resistance in chickpea plants against *Foc* was achieved by N_{fo}, BRC, *R.leguminisarum* with soil and seed treatments. Soil treatments with BRC, N_{fo} and *R.leguminisarum* caused significantly less disease incidence, 11,16 and 39% respectively, and 8, 12 and 41% when seed treatment respectively compared with 69% for the control treatment(*Foc*). In field experiment, treatment with bio control agents, BRC, local and commercial *T.harzianum*, *P.fluorescens* and *R.leguminisarum* significantly reduced wilt disease incidence compared with control treatment. The most effective treatment was BRC, 13 % disease incidence compared with 45 % in control treatment. The tested bio control agents increased plant height, wet and dry weights and yield of chickpea plants. The greatest height of chickpea plant 38.2 and 38.3cm were recorded for BRC and *T.harzianum* + *P. fluprescens* treatments. Seed treatment with BRC scored the highest average plant weights, 41.4 and 13.4 g respectively. BRC and *T.harzianum* + *P.fluprescens* treatments were significantly superior over *T.harzianum*, *P.fluprescens* (local and commercial product) and *R.leguminisarum* in yield weight 342 and 346 g m⁻² respectively.

INTRODUCTION

Chickpea (*Cicer arietinum* L.) is one of the most important pulse crops worldwide. The global chickpea area was about 11 million ha to produce 8.8 million tons with an average yield of nearly 800 kg ha⁻¹ (17). Chickpea importance is due to it's nutrition role in human food and soil fertility. Iraq is producing about 1% of the world production of chickpea (17). However, the cultivation of chickpea in Iraq is still limited because of the low yield of the crop, not mechanically

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harvested and the high susceptibility to pathogens. *Fusarium* wilt caused by *Fusarium oxysporum* f.sp. *ciceri* (Padwick) Matu and Satu (*Foc*) was recorded in more than 33 countries is considered as one of the most important diseases which limit chickpea production (32). The high incidence of chickpea *Fusarium* wilt was often observed in Nineveh province, Iraq. The increased demands for food prompted the use of chemical pesticides and fertilizers which achieved part of this aim. But because of the negative effects of these chemicals on the environment, human and animal health and its high cost and the emergence of resistance in the pathogen, researchers have focused on alternative control methods such as biological control and induced systemic resistance in plant against many plant pathogens (23). The fungus *Trichoderma* was considered as one of the most important bio control agents for soil, seed borne and other pathogens (10,11). Seeds of chickpea treated with *T. harzianum* reduce *Fusarium* wilt incidence by 25-27% and increased yield by 13% (24). Recent study indicated that *T. harzianum* and *T. viridi* was able to inhibit *Foc* growth and reduce wilt disease incidence under field conditions and increase seed germination, root length and shoot height (14). *Pseudomonas fluorescens* has a major role as bio control of soil borne pathogenic fungi, producing several compounds which effects plant pathogens like phenazin (40,18), pyrolnitrin (7), phloroglucinol (20) and chelating agent (35). Recent reports indicated that chickpea seeds treated with *P. fluorescens* before inoculation with *Foc* showed reduced wilt disease incidence and increased yield (22, 24,41).

Induced systemic resistance was reported against different plant pathogens since the thirties of the last century (9). Numerous studies have confirmed the ability of non pathogenic *Fusarium* to suppress *Fusarium* wilt (26,33,36, 37,38). Chickpea seedling treated with *Rhizobium* isolates led to reduce wilt disease incidence and increased plant height, weight and yield of chickpea plant (2,3,4,5).

This study was undertaken to evaluate the biocontrol and induced systemic resistance ability of different bio agents in chickpea plants against *Fusarium oxysporum* f. sp. *ciceri*.

MATERIALS AND METHODS

Biological Materials

Chickpea seeds, *Cicer arietinum* L. cv "Marakishi" susceptible to *Foc* was obtained from the local market and was used in the experiments.

Fusarium oxysporum f.sp. *ciceri*

This pathogen was isolated from chickpea plants with the characteristic *Fusarium* wilt symptoms from Nineveh province (400 km north of Baghdad, Iraq) and propagated on potato dextrose agar medium. The fungus was stored in autoclaved soil at 4 C and used in the experiments.

Non Pathogenic *Fusarium oxysporum*

A non pathogenic *F.oxysporum* isolate (N Fo) was isolated from chickpea plants from Nineveh province. This isolate caused no disease symptoms on inoculated susceptible chickpea cv Marakishi.

Rhizobium leguminisarum (R.I)

This R.I isolate was obtained from the Integrated Management of Plant Production and Protection Center, Plant Protection Directorate, Ministry of

Agriculture, Baghdad, Iraq. This isolate was originally isolated from chickpea plants with active bacterial nodules.

Bio-Root Care (BRC)

BRC bio pesticides (Dr. Ragan laboratories, Chennai, India) was a mixture consisting of various bio control agents such as: *P. fluorescens*, *Trichoderma viride*, *T. harzianum*, *Bacillus subtilis* and *Paceilomyces lilacinus*.

Pseudomonas fluorescens

Local isolate of *P. fluorescens* was obtained from Dr. Audai Najem (Department of plant protection, college of agriculture, university of Baghdad). Commercial product of this bacterium was obtained from Dr. Ragan laboratories, Chennai, India.

Trichoderma harzianum

Local isolate of *T. harzianum* was obtained from DR. Hadi M. Aboud (Ministry of science and technology, Baghdad, Iraq). Commercial product of *T. harzianum* was obtained from Dr. Ragan laboratories, Chennai, India

In vitro* evaluation of bioagents against *Fusarium oxysporum* f.sp. *ciceri

P. fluorescens, *T. harzianum*, *R. leguminisarum*, N_{Fo}, and the bio pesticide BRC were used to evaluate their antagonistic activity against *Foc*. Each antagonist and the pathogen were simultaneously inoculated at the opposite ends of 9cm diam. Petri dishes containing Potato Dextrose Agar (PDA). Discs of 5 mm diam from actively growing culture of *Foc* and *T. harzianum*, N_{Fo} and 50µl of BRC (5g l⁻¹) was placed at equal distance from the ends of PDA plates. *P. fluorescens* and *R. leguminisarum* were streaked on one side of *Foc* inoculated plates. Each test was replicated three times and inoculation with *Foc* only served as control. Diameter of the pathogen was measured after incubation at 25C (23).

Biocontrol of *Foc*: Seed treatment with antagonists

Seeds were surface sterilized with 1% sodium hypochlorite solution for 10 min, and washed with sterile distilled water. Seeds (10g) were then thoroughly mixed with 1ml of local *P. fluorescens* (1x10⁸ CFU ml⁻¹) and local *T. harzianum* (10⁶ spores ml⁻¹) and 5g kg seed⁻¹ for the commercial products. One % carboxymethyl cellulose was added as sticking agent. Loamy sand soil was used for the experiment. Sterilization of the soil was done at 121C at 1.5 kg cm⁻² for 30 min 2days interval prior to sowing and later on mixed with 100 ml of 1x10³ spore ml⁻¹ kg soil⁻¹ *Foc*. Three treated seeds of chickpea were sown in 18 cm diam pots filled with inoculated soil. Each treatment was triplicated. Untreated seeds were used as control. Wilt incidence, was recorded 30 days after sowing.

Soil treatment with antagonists

Autoclaved soil in pots (1 kg) were inoculated with 100 ml of 1x10³ spore ml⁻¹ kg soil⁻¹ *Foc* 2 days prior to inoculation with antagonists. The inoculums 100 ml kg soil⁻¹ of local *P. fluorescens* (1x10⁸ CFU g soil⁻¹) and *T. harzianum* (10⁶ spore.g soil⁻¹), 5g kg soil⁻¹ for commercial products was mixed with soil. Three treated seeds of chickpea were sown in pots (18 diam) filled with inoculated soil. Treatments were replicated three times and untreated seeds were used as control. Wilt incidence, was recorded 30 days after sowing.

Induced resistance in chickpea plants against *Foc*

Chickpea seeds were surface sterilized with 2% sodium hypochlorite solution for 10 min, and washed with sterile distilled water. Seeds then were planted in plastic pots (18cm diam, 3 seeds pot⁻¹) and maintained in a plastic house. When seedlings were 30 days old, the soil was treated with 100ml of 1×10⁶ spores ml⁻¹ of *NFo*, 1×10⁸ spores ml⁻¹ of *R.l* and BRC at 5g kg soil⁻¹. After 24 hours the pots were treated with spore 1×10³ ml⁻¹ spores suspension of *Foc*. Soil treated with *NFo*, *R. leguminisarius*, BRC, *Foc* and intact healthy plant were control treatments.

Bioassay under field conditions

Eight experimental units, 10×25 m were chosen in chickpea cultivated field in Alkosh, Nenivah province, known to encounter Fusarium wilt disease in previous seasons. Chickpea seeds, pretreated with 100ml kg seed⁻¹ suspension of local *P. fluorescens*, *R. leguminisarius* (10⁸ cfu ml⁻¹) and *T. harzianum* (10⁶ spore ml⁻¹) and 5g kg seed⁻¹ of the commercial products (*P. fluorescens*, *T. harzianum* and BRC) were planted in the experimental units. Wilt incidence, shoot length, wet and dry weight of plant was recorded.

RESULTS DISCUSSION

In vitro evaluation of bioagents against *Foc*

All the tested biocontrol agents inhibited the growth of the pathogenic fungus *Foc* (Table 1). *T.harzianum* caused significantly (P=0.05) the highest inhibition percentage, 59.7% of *Foc* compared with other test bio control agent. This was followed by 53.3 and 52.0% inhibition when *P.fluoresens* and *NFo* were used respectively. The latter treatments were significantly different compared with BRC and *R.leguminisarius* treatments which recorded 32.6 and 35.3 % inhibition of *Foc* respectively.

Table1: Effect of bio agents on *Fusarium oxysporum* f.sp.*ciceri* (*Foc*) growth on PDA 7 days after inocubation at 25C

Treatment	Grow inhibition(%)
<i>Foc</i> +Bio-Root care	32.6
<i>Foc</i> + <i>NFo</i>	52.0
<i>Foc</i> + <i>T.harzianum</i>	59.7
<i>Foc</i> + <i>P. fluorescens</i>	53.3
<i>Foc</i> + <i>R. leguminisarius</i>	35.3
<i>Foc</i> (Control)	0.0
LSD (P = 0.05) =4.8	

Discs of 5 mm diam. of *Foc* and *T. harzianum*, *NFo* and 50µl of BRC was placed at equal distance from the periphery of PDA plates. *P. fluorescens* and *R. leguminisarius* were streaked to one side. Each test was replicates three times. *Nfo* = non pathogenic *Fusarium oxysporum*.

Bio control of Fusarium wilt

Results revealed the ability of *P.fluoresens* and *T.harzianum* to significantly (P = 0.05) reduce chickpea wilt incidence compared with the control treatment for both seed and soil treatments (Table 2). Treated chickpea seeds with the local isolates of *P.fluoresens* and *T.harzianum* caused significantly more disease control compared with the commercial product of these isolates. The disease incidence was 7 and 9 % when local *P.fluoresens* and *T.harzianum* were used respectively, while it was 16 and 19 % for commercial product respectively. Treatment of both local and commercial *P. fluoresens* and *T.harzianum* significantly reduced wilt disease incidence compared with control treatment (*Foc* only) which recorded 62% disease incidence (Table2). Soil treatment with the local *T.harzianum* and *P. fluoresens*

caused significantly ($p = 0.05$) reduced disease incidence of 7 and 12% respectively compared with 18 and 19 % for the commercial product of the bio control agent respectively.

Table 2: Bio control of *Fusarium wilt* in Marakishi chickpea, *Cicer arietinum* L.

Soil treatment	Disease incidence (%)
<i>T. harzianum</i> local	7
<i>P. fluorescens</i> local	12
<i>T. harzianum</i> (commercial product)	18
<i>P. fluorescens</i> (commercial product)	19
Control (<i>Foc</i> only)	62
LSD ($P = 0.05$) for treatment = 5.9	
Seed treatment	Disease incidence (%)
<i>T. harzianum</i> (local)	7
<i>P. fluorescens</i> (local)	9
<i>T. harzianum</i> (commercial product)	16
<i>P. fluorescens</i> (commercial product)	19
Control (<i>Foc</i> only)	62
LSD ($P = 0.05$) for treatment = 4.8	

Numbers represent three replicates. 100 ml kg soil⁻¹ of *Foc* spore suspension was added (10^3 spore ml⁻¹) 2 days after local *P. fluorescens* (10^8 spore ml⁻¹), (100ml kg soil⁻¹), and 50 g kg soil⁻¹ of local *T. harzianum* propagated on millet, 5g kg soil⁻¹ of commercial products were added. Sowing was 2 days after treatments. Seeds treated with 1ml 10g seed⁻¹ of *P. fluorescens* (10^8 spore ml⁻¹) and (10^6 spore ml⁻¹) of *T. harzianum* and 5g kg seed⁻¹ of commercial products, planted in pots contain soil inoculated with 100ml of *Foc* was added (100ml kg soil⁻¹).

Induced systemic resistance in chickpea plants

Induced systemic resistance (ISR) of chickpea plants was achieved when soil was treated with bio agent BRC, N Fo and *R. leguminisarum* before *Foc*. This was manifested by the reduction of wilt disease incidence in bio agent treatments (Table 3). The disease incidence was 11 and 16% when soil was treated with BRC and N Fo respectively with significant ($p=0.05$) difference compared with 39% for *R. leguminisarum*. Similarly ISR was observed in chickpea when soil was treated with the bio agents (Table 3).

Field experiment

Seed treatments with BRC, *T. harzianum*, *P. fluorescens* local and commercial product, *R. leguminisarum*, was effective in reducing wilt disease incidence in chickpea plants significantly ($p = 0.05$) compared with control treatment (Table 4). BRC was the most effective treatment recorded disease incidence 13% which is significantly difference compared with the other rest of treatment (*T. harzianum*, *P. fluorescens* local and commercial product, *R. leguminisarum*). This was followed by local *T. harzianum*, *P. fluorescens* and commercial.

Table 3: Bio control and induced resistance in chickpea, *Cicer arietinum* L. against *Fusarium oxysporum* f.sp.ciceri (*Foc*)

Soil treatment	Disease incidence (%)
Bio-Root Care	11
Nonpathogenic <i>Fo</i> (N Fo)	16
<i>R. leguminisarum</i>	39
<i>Foc</i> (Control)	62
LSD ($P = 0.05$) = 7.4	
Seed treatment	Disease incidence (%)
Bio-Root Care	8
Nonpathogenic <i>Fo</i> (N Fo)	12
<i>R. leguminisarum</i>	41
<i>Foc</i> (Control)	62
LSD ($P = 0.05$) = 5.5	

Numbers represent three replicates. Bioagents (100 ml kg soil⁻¹ as follows: N Fo (10^8 spore ml⁻¹), *R. leguminisarum* (10^8 spore ml⁻¹), BRC (5g kg⁻¹) were added 7 days after sowing. Seeds were similarly treated with the bio agent. When plants were 2 – 3 leaves, 100 ml kg soil⁻¹ of spore suspension of *Foc* was added (10^3 spore ml⁻¹). Wilt disease incidence was recorded 15 days after inoculation with *Foc*.

Table 4: Effect of bioagents on Fusarium wilt disease incidence in chickpea, *Cicer arietinum* L. under field conditions

Treatment	Disease incidence (%)
Bio-Root care (BRC)	13
<i>T.harzianum</i> (local)	20
<i>T.harzianum</i> (commercial product)	24
<i>P. fluorescens</i> (local)	21
<i>P. fluorescens</i> (commercial product)	22
<i>T.harzianum</i> + <i>P. fluorescens</i>	12
<i>R.leguminisarum</i>	30
Control	45
LSD (p =0.05) = 5.8	

Numbers represent four replicate, each replicate is 10 plants in Alkosh, Nenivah province. Chickpea seeds were pre-treated with 100 ml kg seed⁻¹ of local *P. fluorescens*, *R. leguminisarum* (10⁸ cfu ml⁻¹) and *T. harzianum* (10⁶ spore ml⁻¹) and 5g kg seed⁻¹ from the commercial product (*P. fluorescens*, *T. harzianum* and BRC). Wilt incidence was recorded at the end of the growing season.

product treatment which was recorded significantly difference disease incidence of 20 and 24% respectively compared with *R. leguminisarum* treatment which recorded infection rate 30%.

Treatments with bioagent used in this study induced significant increment in most plant growth parametery (Table5). The maximum average of plant height was observed when chickpea seeds treated with BRC, *T.harzianum* + *P. fluorescens* (local and commercial product) and *R.leguminisarum*. The plant height was 38.2 and 38.3cm with BRC and *T.harzianum*+*P. fluorescens* respectively (Table 5). Other treatment with bio agent did not show significant differences which ranged between 34.2 and 36.1cm in commercial product treatment of *P. fluorescens* and *T.harzianum* respectively. All treatments outperformed the control treatment which recorded 28.3cm. All the tested bioagent increased the wet and dry weight of the plant. The highest significant increased (p =0.05) of wet and dry weight was 41.4 and 13.4g respectively with BRC treatment compared with other test treatment (Table 5). Other test bio agent treatment did not cause significant differences in wet and dry weight. The wet and dry weight of these treatments ranged between 30.1 and 35.3g for wet weight, and 10.6 to11.4g for dry weight. All treatments outperformed the control treatment which recorded 23.7 and 7.4g for wet and dry weight respectively. The bioagent affected yield, significantly (p = 0.05) maximum yield weight was 342 and 346 g m⁻² when BRC and *T.harzianum* + *P. fluorescens* treatment were used respectively compared with 292 and 294.3 for local *T.harzianum*, *P. fluorescens* and with 287.4 , 277.2 g for commercial *T.harzianum*, and *P. fluorescens* products and with 289.5 for *R.leguminisarum* treatment (Table 5). No significant differences were recorded between *T.harzianum*, *P. fluorescens* (local and commercial product) and *R. leguminisarum* treatment which ranged between 277.2 and 289.5g m⁻² with *P. fluorescens* and *R.leguminisarum* treatments respectively. All treatment with the bioagent outperformed the control treatment which was 233.8 g m⁻².

Table 5: The influence of bioagents on chickpea, *Cicer arietinum* L. growth and yield under field conditions

Treatment	Plant height (cm)	Wet weight (g)	Dry weight (g)	Yield weight (g m ⁻²)
Bio-Root Care	38.2	41.4	13.4	342
<i>T.harzianum</i> (local)	35.8	31.6	10.6	292
<i>T.harzianum</i> (commercial product)	35.1	31.1	10.7	287.4
<i>P. fluorescens</i> (local)	36.0	33.6	11.5	294.3
<i>P.fluorescens</i> (commercial product)	34.2	30.1	10.7	277.2
<i>T.harzianum</i> + <i>P. fluorescens</i>	38.3	35.3	11.4	346
<i>R.leguminisarum</i>	36.9	34.0	11.4	289.5
Control	28.3	23.7	7.4	233.8
LSD (P = 0.05)	1.8	5.5	2.5	45.4

Numbers represent four replicate, each replicate is 10 plants in Alkosh, Neniwah province. Chickpea seeds were pre-treated with 100 ml kg seed⁻¹ of local *P. fluorescens*, *R. leguminisarum* (10⁸ cfu ml⁻¹) and *T. harzianum* (10⁶ spore ml⁻¹) and 5g kg seed⁻¹ from the commercial product (*P. fluorescens*, *T. harzianum* and BRC).

Results of this study indicated the ability of bioagent used in this study to inhibit *Foc* growth on artificial medium and confirmed, previous studies on the ability of *Trichoderma* isolates to inhibit *Foc* growth (14,28,30). Other studies, reported on the ability of *P. fluorescens* and *Rhizobium* to inhibit *Foc* growth for more than 30-40% in dual culture test (2,4,23,42). The superiority of the local *T.harzianum* and *P. fluorescens* in reducing wilt disease incidence on chickpea plants compared with the commercial products of these microorganisms may be due to the adaptation of the local isolate to environmental condition or to their directly use after propagation without passing through storage duration. Result of this study support previous studies on the ability of *T. harzianum* to *in vitro* suppress *Foc*, and control of the disease under greenhouse and field conditions (34). Treatment of chickpea seeds with *Trichoderma* isolates under greenhouse conditions reduced wilt disease incidence, increased plant height and wet and dry weight (28). The disease incidence of chickpea wilt was reported to be reduced to about 40% after treatment with *T. harzianum* (29). Similarly seed treatment with *P.fluorescens* led to control chickpea wilt disease and reduce disease incidence under green-house and field conditions and increased chickpea yield (22,23,24,41). Furthermore, the use of *P.fluorescens* against chickpea wilt disease led to delay development of the disease symptoms (25). The biocontrol agents were known to suppress pathogens directly through parasitism, antibiosis and lytic enzymes and indirectly through nutrient competition, element chelating and induced plant resistance (31).

Results of this study confirmed the ability of BRC, NFO and *R.leguminisarum* when exceeded *Foc* inoculation to reduce wilt disease incidence in chickpea plant. This is may be resulted by stimulating the systemic resistance in chickpea plants through the production of enzymes and phenolic compounds, promote plant growth and the strengthening of physical defensive barriers. This study indorse the results of previous studies stating that seeds or seedling of leguminous crop treated with *Rhizobium* caused significant reduction in some root diseases caused by soil born fungi (8,12). Mechanism of resistance in the plant is activated as a result of *R.leguminisarum* and NFO treatment due to production of phytoalexin by roots, biotoxins and pathogenesis related proteins, PRP (15,39). Chickpea seedlings treated with *Rhizobium* isolates before *Foc* was found to increase phenylalanine ammonia lyase, chalcon, isoflavon reductase, peroxidase, poly phenol oxidase and phenolic

compound production (2,4,5). Chickpea seedlings treated with *NFo* led to reduce wilt disease incidence and disease severity caused by *Foc* (19).

Result of biocontrol chickpea wilt disease under field conditions was similar to the results of the plastic house. The bioagent used in this study BRC, *T.harzianum*, *P.fluorescens*, *T.harzianum* + *P.fluorescens* (local and commercial product) and *R. leguminisarum* were able to biocontrol chickpea wilt disease. The successful biocontrol of wilt disease incidence was reflected on the increased plant height, wet and dry weight and yield of chickpea plants. Results of this study confirmed that of previous studies which indicated that chickpea seeds treatment with bioagent caused decreased Fusarium wilt disease incidence (1,6,21,24). Soil treated with *T.harzianum*, *T.koningii*, *T.pseudokoningii* delayed disease appearance and reduce disease severity (27). The increase of some growth and yield criteria could be due to nitrogen fixation by symbiotic microorganisms with plant roots. These microorganisms have the ability to transform not available elements to available elements for plants (43). These microorganisms could regulate the action of hormone and other factors which are responsible for the growth and development of plant. Bacteria were able to produce plant hormone like auxins, ethylene, gibberellins and cytokinine (16).

Further research is needed to establish a successful and biocontrol alternatives and decreased the demands on chemicals.

REFERENCE

- 1-Animisha, Z.S.; K.K. Jaiswal and P. pandy (2012). Integrated management of chickpea wilt Incited by *Fusarium oxysporum f. sp. Ciceri* International J. of Agric. Res., 7: 284-290.
- 2-Arfaoui, A.; B. Sifi; A. Boudab-ous; I. El Hadrami and M. cherif (2006). Effect of *Rizobium* isolates on isoflavonoid levels in chickpea plants infected with *Fusarium oxysporum f. sp. Ciceri*. Phytopathol. Mediterr, 45: 24-34.
- 3-Arfaoui, A.; A. El Hadrami; Y. Mabrouk; B. Sifi; A. Boudabous; I. El Hadrami; F. Daayf and M. Cherif (2007). Treatment of chickpea with rizobium isolates enhances the expression of phenyl propanoid defense related genes in response to in infection by *Fusarium oxysporum f. sp. Ciceri*. Plant physiology and Bio., 451. 6-7 :.470-479.
- 4-Arfaoui, A.; B. Sifi; M. El Hassni; I. El Hadrami; A. Boudabbous and M. cherif (2005). Biochemical Analysis of chickpea protection against Fusarium wilt Afforded by two Rhizobium isolates. Plant pathology J., 4(1): 35-42.
- 5-Arfaoui, A.; B. Sifi; A. Boudabous; I. El Hadrami and M. cherif (2006). Identification of Rhizobium isolates possessing Antagonistic Activity Against *Fusarium oxysporum f. sp. Ciceri* the causal agent of Fusarium wilt of chickpea. J. of plant pathology, 88:67-75.
- 6-Bouregghda, H. and Z. Bouznad (2009). Biological control of Fusarium wilt of chickpea using isolates of *Trichoderma atroviride*, *T. harzianum* and *T. longibrachiatum*. Acta phytopathologica et entomologica hungarica, 44: 25-38.
- 7-Burkhead K. and M.J. Geoghegan (1994). Antibiotics. IN: Soil borne plant pathogens. 9 Burkhead , k. ed.) Macmillon, New York, NY, USA, 368 pp.

- 8-Chakraborty, U. and B.N. Chakraborty (1989). Interaction of *Rhizobium leguminosarum* and *Fusarium solani* f.sp. *pisi* in pea affecting disease development and phytoalexin productions. Canadian J. of Botany, 67:1698-1702.
- 9-Chester, K.S. (1933). The problem of acquired physiological immunity in plants. Quat Rev Bio., 8: 275-324.
- 10-Chet, I.; Y. Hadar; Y. Elad; J. Katan and Y. Henis (1979). Biological control of soil borne pathogens by *Trichoderma harizianum*. In: soil Borne plant pathogens (B.Schippers), Academic press, London, UK 585.
- 11-Chet, I. and R.R. Baker (1981). Isolation and biocontrol potential of *Trichoderma hamatum* in suppression of *Rhizoctonia* damping-off in a bark compost amended container medium. Phytopathology, 80: 73-77.
- 12-Dar, H.; G.M.Y. Zargar and G.M. Beigh (1997). Biocontrol of *Fusarium oxysporum* root rot in the common bean (*Phaseolus vulgaris* L.) by using symbiotic *Glomus mosseae* and *Rhizobium leguminisarum*. Microbial ecology, 34:74-80.
- 13-Dubey, S.C.; M. Suresh and B. Singh (2006). Evaluation of *Trichoderma* species against *Fusarium oxysporum* f.sp. *206icero* for integrated management of chickpea wilt Biological control, 40: 118-12.
- 14-Duijff, B. J.; D. Pouhair; C. Olivain; C. Alabouvette and P. Lemanceau (1998). Implication of systemic induced resistance in the suppression of *Fusarium* wilt of tomato by *Pseudomonas fluorescens* WCS417r and nonpathogenic *Fusarium oxysporum* Fo47.
- 15-Frankenberger, W. T. and M. Arshad (1995). Phytohormones in soils - microbial production and function. New York:Marcel Dekker.
- 16-Gaur, P.M.; S. Tripath; C.L.L. Gowda; R. Ranga; H.C. Sharma; S. Pande; and M. Sharma (2010). Chickpea seed production manual. Patancheru 502 324, Andhra Pradesh, India : International Crops Research Institute for the Semi-Arid Tropics. 28pp.
- 17-Gurusiddaiah, S.; D.M. Weller; A. Sarkar and R.J. Cook (1986). Characterization of an antibiotic produced by a strain of *P. fluorescens* inhibitory to *Gaeumannomyces graminis* var *tritici* and *pythium* spp. Antimicrobial agents and chemotherapy, 29: 488-495.
- 18-Hervas, A.; J.L. Trapero-casas and R.M. Jimenez-Diaz (1995). Induced resistance against *Fusarium* wilt of chickpea by nonpathogenic races of *Fusarium oxysporum* f. sp. *ciceri* and non pathogenic *F. Oxysporum* plant Dis. 79: 1110-1116.
- 19-Howell, C.R. and R.D. Stipanovic (1980). Suppression of *pythium ultimum*-induced damping-off of cotton seedling by *Pseudomonas fluorescens* and its antibiotics pyoluteorin. Phyto pathology, 70: 712-715.
- 20-Karimi, K.; J. Amini; B. Harighi and Bahramnejad (B.2012). Evaluation of biocontrol potential of *Pseudomonas* and *Busillus* spp. Against *Fusarium* wilt of chickpea. Australian J. of crop Sci., 6(4):695-703.

- 21-Kaur, R.; J. Kaur; R.S. Singh and C. Alabouvette (2003). Evaluation of non pathogenic *Fusarium* and *pseudomonas fluorescent* against *Fusarium oxysporum* f. sp. *ciceri* proceedings Indian Phytopathological society (NZ): 69-74.
- 22-Kaur, R. and R.S. Singh (2007). Study of Induced Systemic resistance in *cicer arietinum* L. du to non pathogenic *Fusarium oxysporum* Using a Modified split root technique. J. Phytopathology, 155:694-698.
- 23-Khan, M.R.; S.M. Khan and F.A. Mohiddin (2004). Biological control of fusarium wilt of chichepa through seed treatment with the commercial formulation of *Trichoderma harzianum* and/or *Pseudomonas fluorescens*. Phytopathologia mediterranea, 43 (1).
- 24-Landa B.B.; J.A. Navas-cortes and R.M. Jimenez-Diaz (2004). Integrated management of fisarium wilt of chickpea with sowingdate, host resistance and Biological control. Phytopatholog y, 94: 964-960.
- 25-Larkin, R.R. and D. Fravel (R.1996). Ecological characteristics of biological control of Fusarium wilt of tomato using non pathogenic. Fusarium spp. Phytopathology, 86:59.
- 26-Meki, S.; S. Ahmed and P. K. Sakhuja (2011). Control of chickpea wilt (*Fusarium oxysp-orum* f.sp.*ciceris*) using *Trichoderma* spp.in Ethiopia. Archives of Phytopathology and plant protection. 44(5):432-440.
- 27-Merkuz, A. and A. Getachew (2012). Management of chickpea wilt (*Fusarium oxysporum* f. sp. *ciceri*) using *Trichoderma* spp. International J., of current Res., 4: 128-134.
- 28-Moradi,H., B. Bahramnejad; J. Amini; A. Siosemardeh and K. Haji Allahverdipoor (2012). Suppres- sion of chickpea (*cicer arietinum* L.) Fusarium wilt by *Bacillus subtillius* and *Trichoderma harzianum*. plant omics J., 5: 68-74.
- 29-Nikam, P.S.; G.P. Jagtap and P.L. Sonkakke (2007). Manage- ment of chickpea wilt caused by *Fusarium oxysporum* f. sp. *ciceri*. African journal of Agric. Res., 2:692-697.
- 30-Pal, K.K. and B.M. Gardener (2006). Biological control of plant pathogen. The plant health instructor DOI:10.1094IPHI-A-1117-02.
- 31-Pande, S.; M. Sharma; A. Nagavardhini and T Rames-hwar (2012). High Throughput Phenotyping of Chickpea Diseases: Stepwise identification of host plant resistance. Information Bulletin No. 92 Patancheru 502 324, Andhra Pradesh, India: International Crops Research Institute for the Semi-Arid Tropics. 56 pp. ISBN 978-92-9066-552-6. Order code: IBE 092
- 32-Paulitz, T.C.; C.S. Park and R. Baker (1987). Biological control of Fusarium wilt of cucumber wilt non pathogenic isolates of *Fusarium oxysporum* .Can. J. Microbial., 33: 349-353.
- 33-Ramezani, H. (2009). Efficacy of some fungal and bacterial bioagents against *Fusarium oxysporum* f. sp. *ciceri* on chickpea. Plant Protection J., 1:108-113.
- 34-Sakthivel, N.; E. Sivamani; N. Unnmalai and Ganamanikam (1986). Plant growth promoting rhizobacteria in enhancing plant growth and suppressing plant pathogen. Current Sci., 55:22-25.

- 35-Schneider, R.W. (1984). Effect of non pathogenic Strains of *Fusarium oxysporum* on celery infection by *Fusarium oxysporum* f. sp. api and a novel use of line weaver- burk double reciprocal plot technique. *Phytopathology*, 47: 646-653.
- 36-Singh, R.S.; J. Kanr; R. Kaur and C. Alabouvette (2002). Effect of amendment with farm yard manure on biocontrol potentiality of non-pathogenic *Fusarium* against chickpea wilt. *plant Dis. Res.*, 17: 207.
- 37-Tamietti, G. and C. Alabouvette (1986). Resistance des sols aux maladies: x111- Role des *Fusarium oxysporum* non pathogènes dans les mécanismes de résistance d'un sol de noirmoutier aux fusarioses vasculaires. *AGRONOMIE*, 6:541-548.
- 38-Tamietti, G.; L. Ferraris; A. Matta and I.A. Gentile (1993). Physiological responses of tomato plants grown in *Fusarium* suppressive soil. *J. Phytopathol.*, 138:66-76.
- 39-Toohey, J.I.; C.D. Nelson; G. Krotkov (1965). Toxicity of phenazine carboxylic acid to some bacteria, algae, higher plants, and animals. *Canadian J. of Botany* 43: 1151–1155.
- 40-Ul-Haq, M.I.; S. El-Hassan; S. Growen, and N. Javed (2009). Effect of two rhizobacterial isolates and Neem cake Application on control of chickpea wilt caused by *Fusarium oxysporum* f. sp. *208icero*. *Arab. J.PI.Prot.* 27: 103-110.
- 41-Vannia, R.P.; S. Muhammad and M.S. Pallavi (2013). In vitro evaluation of bio-agents, fungicides and herbicides against *Fusarium oxysporum* f.sp. *ciceri* causing wilt of chickpea. *A Quarterly journal of life Sci.*, 10:403-405.
- 42-Vessey, J. K. (2003). Plant growth promoting rhizobacteria as biofertilizers. *Plant and Soil*, 255: 571–586.

المكافحة الاحيائية وتحفيز المقاومة الجهازية في الحمص (*Cicer aratinum*)

L. ضد الفطر *Fusarium oxysporum* f.sp. *ciceri*

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الملخص

أظهرت عوامل المكافحة الإحيائية (BRC, Bio-Root Care و *Trichoderma harzianum* و *Pseudomonas fluorescens* و *Rhizobium leguminisarum* وعزلة غير ممرضة من NFO, *Fusarium oxysporum* فعالية واضحة في تثبيط نمو الفطر *Fusarium oxysporum* f.sp. *ciceri* (Foc) فقد بلغ أعلى معدلاً لتثبيط نمو Foc عند استخدام *T.harzianum* ، إذ كانت نسبة التثبيط 59.7% وبفارق معنوي عن باقي المعاملات (BRC و NFO و *P.fluorescens* و *R. leguminisarum*). أظهرت النتائج قدرة كل من *T.harzianum* و *P.fluorescens* المحليين على مكافحة مرض ذبول الحمص من خلال خفض نسبة الإصابة معنويًا مقارنة مع معاملة المستحضرين التجاريين *T.harzianum* و *P.fluorescens* ومعاملة السيطرة سواء أكانت عند معاملة البذور أم التربة. فقد كانت نسبي الإصابة 7 و 9% عند معاملي البذور 7 و 12% عند معاملة التربة على التوالي للعزلتين المحليتين، بينما كانت نسبي الإصابة 16 و 19% عند معاملي البذور 18 و 19% عند معاملة التربة على التوالي بالمستحضرين التجاريين. كما أوضحت النتائج مقدرة العوامل الإحيائية NFO و BRC و *R. leguminisarum* في تحفيز المقاومة الجهازية لنبات الحمص عند معاملي التربة والبذور ضد Foc. إذ انخفضت نسبة الإصابة بمرض الذبول معنويًا فكانت نسبة الإصابة عند معاملة التربة 11% عند معاملة BRC و 16% عند معاملي NFO و 39% عند معاملة *R. leguminisarum*. بينما كانت نسبة الإصابة عند معاملة البذور 8% عند معاملة BRC و 12% عند معاملة NFO و 41% عند معاملة *R. leguminisarum* مقارنة 69% عند معاملة السيطرة (Foc). بينت نتائج التجربة الحقلية فعالية عوامل المكافحة الإحيائية لكل من BRC و *T.harzianum* و *P. fluorescens* (محلي ومستحضر تجاري) و *R.leguminisarum* في خفض نسبة الإصابة بمرض الذبول معنويًا مقارنة بمعاملة السيطرة. وكان أكثرها فعالية معاملة BRC التي سجلت نسبة إصابة 13% مقارنة بنسبة إصابة 45% لمعاملة السيطرة. وأدت معاملات المكافحة الإحيائية إلى زيادة ارتفاع النباتات والوزن الرطب والجاف ووزن الحاصل لنباتات الحمص. إذ كان أعلى ارتفاعاً للنبات 38.2cm في معاملي BRC و 38.3cm في معاملة *P. fluorescens* + *T.harzianum*. وسجلت معاملة BRC لبذور الحمص أعلى متوسطاً للوزنين الرطب والجاف وبفارق معنويًا عن باقي المعاملات إذ بلغ الوزن الرطب 41.4g والجاف 13.4g. وتفوقت المعاملتان BRC و *P.fluorescens* + *T.harzianum* معنويًا على المعاملات *T.harzianum* و *P.fluorescens* (محلي ومستحضر تجاري) و *R.leguminisarum* في وزني الحاصل g m^{-2} 342 و 346 على التوالي.

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