# PHYTOCHEMICAL IDENTIFICATION OF PELARGONIUM GRAVEOLENS L. AND STUDYING ITS BIOLOGICAL ACTIVITY

Suad. F. Dhifiar

Researcher

H. M. Habeeb Assist. Prof.

Dept. of Biolo. Colle. Sci. for women, University of Baghdad, Baghdad

soad.fadel1202a@csw.uobaghdad.edu.iq

# ABSTRACT

This study was aimed to determine the active compounds in alcoholic extracts (methanolic 75% and ethanolic 75%) of Pelargonium gravolens leaves and flowers as well as their antibacterial and antioxidant activities. The methanolic extract of dry leaves and fresh flowers (30 g each) gave 4.8650, 2.8015 g respectively. While ethanolic extract for the same parts gave 5.5787, 2.1832g respectively. Spectrophotometric analysis showed that, the flavonoids content in leaves and flowers methanol extract were 20.38, 70.90 µg rutin equivalent/g of extract, respectively, as for ethanol extract those values were 21.89, 71.63 µg rutin equivalent/g of extract, respectively. According to the above findings, it has been noticed that the flowers ethanol extract recorded the highest value for flavonoids (71.63 µg rutin equivalent /g of extract). The results indicate that the free radical scavenging activity ( RSA) of ethanolic extract of the flowers and leaves was lower than that of methanolic extract. Where the RSA percentages of the leaves methanolic extract were (12, 13, 14, 16, 19 %) at a concentration (0.25, 0.50, 0.75, 1.00, 1.25 mg/ml) respectively, and for the flowers were (142, 162, 162) 164, 166%) at the same concentrations respectively.

Keyword: Active compounds, antioxidant, extract. \*Part of Ms. Dissertation of the 1<sup>st</sup> author.

هدفت هذه الدراسة إلى تحديد المركبات الفعالة في المستخلصات الكحوابية ( الميثانول 75٪ و الإيثانول 75٪) لأوراق وأزهار نبات العطرة، بالإضافة إلى دراسة نشاطها المضاد للبكتيريا ومضادات الأكسدة. أعطى المستخلص الميثانولي للأوراق الجافة والزهور الطازجة (30 جم لكل منهما) 4.8650 و 2.8015 جم على التتابع. بينما أعطى المستخلص الإيثانولي لنفس الأجزاء 5.5787، 2.1832 جرام على التوالي. أظهر تحليل مقياس الطيف الضوئي أن محتوى الفلافونوبد في مستخلص الميثانولي للأوراق والازهار كان بواقع 20.38 ، 70.90 ميكروغرام مكافئ روتين/ غرام من المستخلص) على التوالى، أما بالنسبة للمستخلص الإيثانولى، فقد كانت تلك القيم 21.89، 71.63 ميكروغرام روتين مكافئ/ غرام من المستخلص على التوالى. وفقًا للنتائج المذكورة أعلاه ، فقد لوحظ أن مستخلص الإيثانول للزهور سجل أعلى قيمة لمركبات الفلافونوبد (71.63 ميكروغرام روتين مكافئ/ جرام من المستخلص). تشير النتائج إلى أن نشاط إزالة الجذور الحرة (RSA) للمستخلص الإيثانولي للزهور والأوراق كان أقل من نشاط المستخلص الميثانولي. حيث كانت نسب RSA للمستخلص الإيثانولي للأوراق (12، 13، 14، 16، 19٪) بتركيز (0.25، 0.50، 1.00، 1.25 مجم/ مل) على التوالى، وبالنسبة للزهور كانت (142، 162).، 162) 164، 166٪) وعند نفس التراكيز، على التوالى.

الكلمات المفتاحية: المركبات الفعالة، مضاد للأكسدة، مستخلص.

البحث مستل من رسالة ماجستير للباحث الأول.



This work is licensed under a Creative Commons Attribution 4.0 International License. Copyright@ 2025 College of Agricultural Engineering Sciences - University of Baghdad.

Received:8 /1/2023, Accepted:8/3/2023, Published:30 June.

## **INTRODUCTION**

Medicinal plants have a long history of remediation of various types of diseases worldwide because they include several chemical substances that act to prevent, relieve and treat illnesses (2, 3, 6). Pelargonium graveolens, is an aromatic and medicinal plant belonging to Geraniaceae family. They are an erect, multi-branched shrub that grows up to 1.5 m and has a spread of 1 m. The leaves are deeply incised, velvety and soft to the touch. The flowers vary from pale pink to almost white. Some plants are strongly scented, and others have little or no scent (22, 32). Flavonoids are a type of naturally organic compound found in plants. Flavonoids, as a group, comprise upwards of 8000 diverse compounds (27). From a chemical structure standpoint, all flavonoids stemmed from the main skeleton they share and differentiate from each other based on the substituent attached to any part of the structure. Flavonoids possess phenolic and pyrene rings in their structures and have many subclasses, as flavonols, flavones, flavanones, such chalcones, and anthocyanidins (20). The flavonoid skeleton comprises an aromatic ring linked on one side with a six-membered heterocyclic ring, which bears an oxygen atom instead of carbon next to the common side. The two rings connect to another aromatic ring to form the skeleton. Flavonoids have a wide range of pharmacological activities that include anti-oxidant, antimicrobial, antiinflammatory, antimutagenic, antitumor, and their effects on human health are very often ascribed to their potential ability to act by diminishing free radical steady-state concentration in biological systems and so providing antioxidant protection (13, 35). Antioxidant activity was displayed by a number of extracts of representative species and cultivars of *Pelargonium*. Miller (35) established that the flavonoids isolated from P. reniforme produced higher anti-oxidant activity than ascorbic acid. Potent anti-oxidant activity was observed for the extracts of P. betulinum (IC50: 4.13 $\pm$ 0.14 µg/ml) and P. *crispum* (IC50 : 4.49±0.18 µg/ml). cordifolium and P. scabrum also showed potent radical scavenging activity. The antimicrobial activity of extracts of Pelargoniums and their constituents is reported against bacterial (Staphylococcus *Streptococcus* pneumoniae, aureus, Klebsiella pneumoniae, Escherichia coli, Proteus mirabilis, Pseudomonas aeruginosa Haemophilus influenzae), and fungi (Microsporum canis, Microsporum gypseum, Aspergillus fumigatus, Mucor racemosus, Rhizopus nigricans) and pathogens as well as opportunistic yeasts such as Candida albicans (24, 31). Pelargonium glutinosum, Pelargonium pseudoglutinosum, Pelargoniums scabrum and Pelargonium sublignosum exhibited considerable antimicrobial activity against the Gram-positive bacteria (S. aureus and Bacillus cereus) and Gram-negative bacteria (K. pneumonia) (21). This research aims to extract and purify flavonoids from the leaves and flowers of Pelargonium graveolens L. and study their biological activity as antioxidants and antibacterial.

#### MATERIALS AND METHODS

**Plant collection;** Plant was collected from Baghdad nurseries plant. The plant was classified by Dr. Zainab Abed Aoun Department of Biology / College of Science for Women / University of Baghdad. The leaves were dried in the shade at room temperature and ground by the electric grinder, and kept at the refrigerator temperature until use (while fresh (wet) flowers were used).

**Extraction of the active compounds;** The active compounds from leaves and flowers were extracted by Soxhlet using methanol (75%) and ethanol (75%) as extraction solutions. Mixing ratio 1:10 (weight / volume) at a temperature of 60-70  $^{\circ}$  C for 6-8 hours. Then the extract was filtered and dried by a rotary evaporator at a temperature of 40  $^{\circ}$ C (4) (1).

Qualitative detection of the active compounds crude in the extract of Pelargonium graveolens L. leaves and flowers: The active compounds alkaloids, flavonoids, terpenes, tannins and saponins were detected according to Ali et al.(5). Each active compound was detected by using two different specific qualitative reagents (Table1).

| Table 1. The types of detections and the |
|--|
| reagents that used in the current study  |

| Ν  | Type of detection | Type of reagents         |
|----|-------------------|--------------------------|
| 1  | Alkaloids         | A- Mayer reagent         |
| 2  |                   | <b>B-</b> Wagner reagent |
| 3  | Flavonoids        | A- Magnesium             |
|    |                   | crystals and 1% HCl      |
| 4  |                   | B- H2SO4 reagent         |
| 5  | Terpenes          | A- Chloroform and        |
|    |                   | H2SO4                    |
| 6  |                   | B- Anace aldehyde        |
|    |                   | reagent                  |
| 7  | Tannins           | A- FeCl3 reagent         |
| 8  |                   | <b>B-</b> Lead acetate   |
| 9  | Saponins          | A- Foam reagent          |
| 10 |                   | B- HgCl2 reagent         |

#### **Total flavonoids content**

The total flavonoids content in the crude extract was determined by the colorimetric method of aluminum chloride (7) . fifty  $\mu$ l of crude extract (one mg/ml ethanol) were made

up to one ml with methanol, mixed with four ml of distilled water and then 0.3 ml of 5% NaNO<sub>2</sub> solution; 0.3 ml of 10% AlCl<sub>3</sub> solution was added after five min of incubation(20 C), and the mixture was allowed to stand for six min. Then, two ml of NaOH (1M) solution were added, and the final volume of the mixture was brought to 10 ml with doubledistilled water. The mixture was allowed to stand for 15 min, and absorbance was measured at 510 nm. Finally, the total flavonoid content was calculated from the standard curve equation, and the result was expressed as the equivalent of mg rutin per gram of dry weight.

#### **Standard curve of Rutin**

Fifty mg Rutin was dissolve in 25 ml distilled water, the dilutions from this standard solution was prepared as shown in the table (2).

| Table 2. Concentration of rutin | used in preparing the standard curve to estimate the content |
|---------------------------------|--|
|                                 | of flavonoids.   |

| Tube No. | Rutin<br>concentration (µl) | Distilled water<br>(µl) | Concentration<br>mg/ml | Absorbance at 510 nm |  |
|----------|-----------------------------|-------------------------|------------------------|----------------------|--|
| 1        | 0                           | 2000                    | 0                      | 0                    |  |
| 2        | 50                          | 1950                    | 0.050                  | 0.199                |  |
| 3        | 100                         | 1900                    | 0.100                  | 0.28                 |  |
| 4        | 150                         | 1850                    | 0.150                  | 0.499                |  |
| 5        | 200                         | 1800                    | 0.200                  | 0.63                 |  |
| 6        | 250                         | 1750                    | 0.250                  | 0.821                |  |
| 7        | 300                         | 1700                    | 0.300                  | 0.96                 |  |
| 8        | 350                         | 1650                    | 0.350                  | 1.117                |  |
| 9        | 400                         | 1600                    | 0.400                  | 1.28                 |  |
| 10       | 450                         | 1550                    | 0.450                  | 1.389                |  |
| 11       | 500                         | 1500                    | 0.500                  | 1.43                 |  |



Fig 1. Standard curve of rutin concentration (mg/ml).

## **Purification of flavonoids**

The crude extract was concentrated to 60 ml and transferred to a separating funnel; 30 ml of distilled water and 300 ml of ethyl acetate was added, and shake several times. Finally, the organic layer was separated from the aqueous layer and dried using a rotary evaporator at 40 °C. The dry matter, which represents the flavonoids, was collected (16, 12).

High-performance liquid chromatography analysis (HPLC): Quantifying individual flavonoid compounds was performed by reversed-phase HPLC analysis, using an SYKAMN HPLC chromatographic system equipped with a UV detector, (Che station, and a Zorb ax Eclipse Plus-C18-OSD .25cm, 4.6mm column). The column temperature was 30°C the gradient elution method, with eluent A (methanol) and eluent B (1% formic acid in water (v/v)) was performed, as follows: initial 0-4 min, 40 % B; 4-10 min, 50 % B; and flowrate of 0.7 ml/min. The injected volume of samples was 100  $\mu$ l, .and the standard was 100 µL and was done automatically using an auto sampler. The spectra were acquired at 280 nm (30).

#### Determination of antioxidant activity: DPPH Radical-Scavenging Activity (RSA)

The RSA was measured according to Hammood *et al.* (15) with some modulations. First, one ml sample (1 mg/ml) was mixed with 1 ml DPPH solution (0.1 M). The mixture was kept in the dark at room temperature for 30 minutes and then centrifuged at 10,000x g for 5 min. The absorbency was measured at 517 nm. The percentage of the scavenging activity was calculated according to the following equation (15):

## Radical Scavenging Activity = [C - (B-A )/ C] x100

A (Absorbency) = Spectrophotometer reading of the tested sample at 517 nm wavelength.

B = the Absorbance of the control sample at 517 nm (prepared by mixing 1 ml of ethyl alcohol with 1 ml of the sample under study).

C = Absorbance of the positive control at 517 nm (obtained from mixing 1 ml of DPPH with 1 ml of distilled water).

# **RESULTS AND DISCUSSION**

**Crude extraction:** Table 3. shows the extraction yield of active compounds from leaves and flowers of *Pelargonium graveolens*.

The active compounds were extracted by two methods. The first method 75% methanol was applied at 70°C. The results showed that 30 g of dry leaves gave 4.8650 g of active compounds, while 30 g of fresh flowers gave 2.8015 g of active compounds. For the second method, ethanol (75%) was used at 70°C, When 30 g of dry leaves gave 5.5787 g of active compounds, while 30 g of wet flowers gave 2.1832 g of active compounds.

# Table 3 .The yield of active compounds extracted from leaves and flowers of Polynomium angualous plant

| Extraction solution | Leaves     | flowers    | L.S.D |  |
|---------------------|------------|------------|-------|--|
| Methanol            | 16.2166 %  | 9.3383 %   | 3.81* |  |
| (75%)               | (4.8650 g) | (2.8015 g) |       |  |
| Ethanol             | 18.5956 %  | 7.2773 %   | 4.06* |  |
| (75%)               | (5.5787 g) | (2.1832 g) |       |  |
| L.S.D               | 2.75 NS    | 2.29 NS    |       |  |

Mizzi *et al.* (25), studied two types of *Pelargonium*, found that *Geranium atlanticum* contained a higher amount of crude alcoholic extract (19.05%) compared to *Geranium lucidum*, whose content was (16.95%).

Detection of the active compounds of Pelargonium graveolens: The qualitative detection of some active compounds from four plant extracts of Pelargonium graveolens showed that the plant contained alkaloids, terpenes, saponins, tannins and flavonoids, as shown in table 3. These results are consistent with Robert and Philip 2003, findings which the presence of terpenes, mentioned to saponins, tannins and flavonoids in the extracts of Pelargonium graveolens. While Saraswathi et al. (33) referred to the presence of terpenes, tannins and flavonoids in the aqueous and alcoholic extracts of the aerial parts (flowers, stems and leaves) of the plant Pelargonium graveolens. Boukhris et al. (9), reported that the aqueous and alcoholic extract of the leaves and flowers of this plant contain flavonoids and phenols. The variance in the published results may be due to genetic factors and environmental factors such as soil components, pH, temperature, light intensity and photoperiod that affect the type and quantity of these compounds which affect the metabolic pathways of the active compounds (10).

| Table 4. Qualitative detection of active compounds of alcoholic extract of <i>Pelargonium</i> | n |
|---|---|
| arayoolong  |   |

|           |                   |             |                   | graveolens  |                   |             |                   |                        |
|-----------|-------------------|-------------|-------------------|-------------|-------------------|-------------|-------------------|------------------------|
| Active    | Methanol extract  |             | Methanol extract  |             | Ethanol extract   |             | Ethanol extract   |                        |
| compounds | 75% (Leaves)      |             | 75% (Flowers)     |             | 75% (Leaves)      |             | 75% (Flowers)     |                        |
|           | Reagent A*        | Reagent B*  | Reagent A*        | Reagent B*  | Reagent A*        | Reagent B*  | Reagent A*        | Reagent B <sup>*</sup> |
| Alkaloid  | (+) White         | (+) Brown   | (+) White         | (+)Brown    | (+) White         | (+)Brown    | (+) White         | (+)Brown               |
|           | precipitate       | precipitate | precipitate       | precipitate | precipitate       | precipitate | precipitate       | precipitate            |
| Saponins  | (+) Thick         | (+) White   | (+) Thick         | (+) White   | (+)thick          | (+) White   | (+)thick          | (+) White              |
|           | foam              | precipitate | foam              | precipitate | foam              | precipitate | foam              | precipitate            |
| Terpenes  | (+) Brown -       | (+)Brown    | (+)Brown –        | (+)Brown    | (+)Brown -        | (+)Brown    | (+)Brown –        | (+)Brown               |
|           | reddish           | precipitate | reddish           | precipitate | reddish           | precipitate | reddish           | precipitate            |
| Tannins   | (+) Green -       | (+)Yellow   | (+) Green –       | (+)Yellow   | (+) Green -       | (+)Yellow   | (+) Green –       | (+)Yellow              |
|           | bluish            | precipitate | bluish            | precipitate | bluish            | precipitate | bluish            | precipitate            |
| Flavonoid | (+)Red-<br>orange | (+) Red                |

(+) Presence of the active compound. \*The reagents mentioned in Table (1).

#### **Determination of total Flavonoids**

**Fig 2.** shows the total flavonoid extracted ( $\mu$ g of Rutin equivalent/g of extract) by methanol (75%) and ethanol (75%) form *Pelargonium graveolens* leaves and flowers. The data show that total flavonoid concentrations ranged from 21.89 to 71.63 ( $\mu$ g Rutin equivalent/g of extract). The high concentration of total flavonoids was observed in flowers ethanol extract (71.63  $\mu$ g rutin equivalent/g of extract), followed by flowers methanol extract (70.90  $\mu$ g rutin equivalent /g of extract) as compared to the leaves extracts.Pradeepa *et al.* (29) found that the ethanol extract of *Pelargonium* 

gravolens leaves exhibited high amounts of flavonoid content. While Hsouna and Hamdi (17) found that the total flavonoid content ranged from 10.90 to 21.75 mg quercetin equivalent /g, for *Pelargonium tomb* leaves extract. The aqueous extract had the highest level, 21.75 mg kaempferol equivalent /g, while hexane extract had the lowest amount 10.90 mg quercetin equivalent /g. As can be seen, a high phenol content was not always accompanied by high concentrations of flavonoids. These results proved that methanol is the most suitable solvent for the extraction of phenolic compounds (8).





**Quantitative analysis of** *Pelargonium graveolens* **flavonoid:** HPLC technique is widely applied to quantify and separate antioxidants, mainly phenolic and flavonoid compounds in recent years (25, 19). Table 5. shows that six types of available standard flavonoid compounds were used, and compared to what is available in the *Pelargonium graveolens* alcoholic extract, namely, Catechin, Quercetin, Rutin, Galic acid, Kaempferol and Apigenin.

| Flavonoids<br>compounds | Retention<br>time (min) | Methanol (7             | 75%) extract            | Ethanol (75%) extract   |                         |        |
|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|-------------------------|--------|
| -                       | of standard             | Leaves                  | Flower                  | Leaves                  | Flower                  |        |
|                         | flavonoid<br>compounds  | Concentration<br>(µg/g) | Concentration<br>(µg/g) | Concentration<br>(µg/g) | Concentration<br>(µg/g) |        |
| Catechine               | 3.15                    | 103.1727                | 38.5018                 | 50.4578                 | 50.0293                 | 8.03*  |
| Quercetine              | 4.78                    | 55.1714                 | 28.6243                 | 36.3421                 | 12.6263                 | 7.94*  |
| Rutin                   | 8.74                    | -                       | 250.7024                | -                       | 99.9849                 | 23.57* |
| Galic acid              | 10.10                   | 127.7065                | 44.8720                 | 62.6096                 | 100.1716                | 21.08* |
| Kaempferol              | 6.69                    | -                       | 35.1521                 | -                       | 12.2676                 | 8.52*  |
| Apigenin                | 12.68                   | 177.5652                | 119.800                 | 104.2250                | 104.2100                | 21.07* |

Cable 4. Quantitative analysis of flavonoids in *Pelargonium graveolens* alcoholic extract using HPLC

Table 4. also shows the concentrations of flavonoids extracted from Pelargonium graveolens leaves and flowers by methanol and ethanol. The results of HPLC analysis elucidated that the flowers alcoholic extract contained the following compound (catechin, quercetin, rutin, gallic acid, kaempferol and apigenin) and their concentrations were 28.6243, (38.5018,250.7024, 44.8720, 35.1521, 119.800 µg /gm) respectively. Rutin recorded the highest concentration (250.7024  $\mu g$  /g), and quercetin recorded the lowest concentration in comparison to the other  $(28.6243 \ \mu g \ /g)$ . While in the ethanolic extract of the flowers, all flavonoids were appeared at concentrations of (50.0293, 12.6263, 99.9849, 100.1716, 12.2676, 104.2100  $\mu g/g$ ), respectively. It is obvious that the apigenin gave the highest value in this extract (104.2100  $\mu$ g/g), where as kampferol was the lowest (12.2676  $\mu$ g/g). From the above results, the methanol extract of the leaves showed fewer active compounds represented by catechin, quercetin, gallic acid and apigenin at concentrations of (103.1727,55.1714, 127.7065. 177.5652  $\mu g/g$ ), respectively. Apigenin recorded the highest concentration among the active compounds (177.5652  $\mu g/g$ )

while quercetin recorded the lowest concentration (55.1714  $\mu g/g$ ). The ethanol extract of the leaves showed the same types of compounds but with different active concentrations (50.4578, 36.3421, 62.6096, 104.2250 μg/g) respectively. Similarly apigenin recorded the highest concentration among the active compounds (104.2250  $\mu g/g$ ) quercetin while recorded the lowest concentration (36.3421 µg/g). Boukhris et al. (9) identified nine flavonoids by highliquid chromatography performance in aqueous and methanolic extracts of Pelargonium graveolin leaves and flowers. The concentration of flavonoids ranged between 29.9 and 78.2 mg/g in the aqueous and alcoholic extracts of the flowers, respectively, while the concentration of these compounds was 22.5 and 71.2 mg/g in the aqueous and alcoholic extracts of the leaves, respectively. From the results of the previous table, it can be concluded that the flowers and leaves of the Pelargonium graveolens plant are promising sources for flavonoid compounds and effective antioxidants. Furthermore, the flowers have a higher content of active flavonoid compounds compared to the leaves (Figures 3, 4, 5, 6).



Figure 3. Quantitative analysis of flavonoids in Pelargonium graveolens flowers methanolic extract using HPLC



Figure 4. Quantitative analysis of flavonoids in Pelargonium graveolens flowers ethanolic extract using HPLC



Figure 5. Quantitative analysis of flavonoids in Pelargonium graveolens leveas methanolic extract using HPLC





#### **Radical-Scavenging Activity (RSA)**

Figure 7. shows radical scavenging activity (using DPPH) for extracted flavonoids of leaves and flowers of *Pelargonium graveolens* by methanol (75%) and ethanol (75%). Flavonoids are natural compounds in the plant system that inhibit oxidation activity, and play a significant role in absorbing, decomposing the free radicals and neutralizing and

quenching (11, 18). Therefore, the DPPH radical has been used widely to test the potential compounds as free radical hydrogen donors and to investigate plant extracts' antioxidants compounds. The results indicate that the free radical scavenging activity of methanolic extract of the flowers and leaves of the *pelargonium graveolens* plant was lower than that of ethanolic extract. Where the effect of scavenging free radicals for the methanolic extract of leaves were 16, 18, 19, 17, 20 %, at concentrations 0.25, 0.50, 0.75, 1.00, 1.25 mg/ml, respectively, and for flowers were 46, 84. 122, 144. 170 % at the same concentrations. The effectiveness of ethanolic extract for leaves were 12, 13, 14, 16, 19 at concentrations 0.25, 0.50, 0.75, 1.00, 1.25 mg/ml, respectively, and for flowers were 142, 162 ,162, 164, 166 % , at the same concentrations. The higher activity in the resulting methanol extract compared with ethanol extract may be attributed to the polarity indicated by the published literature (28). As the effectiveness of free radical scavenging for flowers extracts (in both ethanol and methanol) were higher than that of leaves extracts, so the flowers contain higher percentage of flavonoids as compared to leaves. Previous literature studied Р. graveolens essential oil found significant antioxidant and biological activities (26). Maria et al. (23) reported that the ethanol extract of the aerial parts of this plant gave a high potency as an antioxidant.



Figure 7. Radical scavenging activity (using DPPH) of flavonoids extracted from leaves and flowers of *Pelargonium graveolens* by methanol (75%) and ethanol (75%).

## **CONFLICT OF INTEREST**

The authors declare that they have no conflicts of interest.

## **DECLARATION OF FUND**

The authors declare that they have not received a fund.

## REFERENCES

1. Alara O. R.; A. Nour and C. I. Ukaegbu. 2018. Soxhlet extraction of phenolic compounds from Vernonia cinerea leaves and its antioxidant activity. J. of Applied Res. on Medicinal and Aromatic Plants. V. 11, pp. 12-17. DOI:10.1016/j.jarmap.2018.07.003

2. Ansari Prawej; A. D. Reberio ;N. J. Ansari; N. J. Ansari; S. Kumar ; J. T. Khan ; S. Chowdhury ; F. M. Abd El-Mordy ; and J. M. A. Hannan. 2024. Therapeutic Potential of Medicinal Plants and Their Phytoconstituents in Diabetes, Cancer, Infections, Cardiovascular Diseases, Inflammation and Gastrointestinal Disorders. Biomedicines. 13(2), 454;

https://doi.org/10.3390/biomedicines13020454

3. Abd El-Rahman, A. A. A.; I. M. Abd El-Aleem,; L. A. Refahy and M. A. El-Shazly. 2016. Total phenolic content, cytotoxic and antioxidant activities of Quisqualisindica (Linn.) growing in Egypt. Der Pharma Chemica. 8 (3):53-59.

4. Al-Delaimy, K. S. and S. H. Ali. 1970. Antibacterial action of vegetable extract on the growth of pathogenic bacteria . J. Sci. Food Agric. 21:110-111. doi.org/10.1002 /jsfa. 2740210214

5. Ali Z. A. A.; M. H. Hadeel and A. J. Liqaa. 2022. Morphological, anatomical and chemical study of an exotic plant *Jatropha integerrima jacq.* 1763 (*euphorbiaceae*) in Iraq. Bull. Iraq nat. Hist. Mus. 17 (1): pp129-140. DOI: https ://doi.org/10.26842 /binhm. 7. 2022.17.1.0129 6. Al-Ezzy, R.M., M.H. Ahmed, H. M. Khalaf, 2024. In vivo study the cytogenetic effect of ammi majus methanolic extract on mitotic index, micronucleus formation and dna damage on mitoxantrone. Baghdad Science Journal, 21(8): 2522–2530. https://doi.org/10.21123/bsj.2024.9034

7. Al-Saffar A. Z.; A. F. Al-Shanon; S. L. Al-Brazanchi; F. A. Sabry; F. Hassan and N. A. Hadi. 2017. Phytochemical Analysis, Antioxidant and Cytotoxic Potentials of Pelargonium graveolens Extract in Human Breast Adenocarcinoma (MCF-7) Cell Line. Asian Journal of Biochemistry. 12 (1) :16-26. DOI: 10.3923/ajb.2017.16.26.

8. Boeing J. S. ; É. O. Barizão; B. C. Silva; P. F. Montanher; V. C. Almeida and J. V. Visentainer.2014. Evaluation of solvent effect on the extraction of phenolic compounds and antioxidant capacities from the berries: application of principal component analysis. Chemistry Central Journal. 8:48, pp 1-9.

9. Boukhris M. ; S. J. S. Monique ; S. Sami and B. Mohamed. 2012. Chemical composition and biological activities of polar extracts and essential oil of rose-scented geranium, *Pelargonium graveolens*. Phytotherapy Res. 27 (8) : 1206-1213.

DOI: 10.1002/ptr.4853. Epub 2012 Oct 2.

10. Cervantes L.; M. Ariza; J.A. Gómez-Mora and L. M. Enamorado. 2019. Light exposure affects fruit quality in different strawberry cultivars under field conditions. Scientia Horticulturae 252(4):291-297.

DOI:10.1016/j.scienta.2019.03.058.

11.Duffy C. F. and R. F. Power .2001. Antioxidant and antimicrobial properties of some Chinese plant extracts . Int. J. Anttimicrob. Agents, 17 : 527-529.

DOI.org/10.1016/S0924-8579(01)00326-0.

12. Ekalu A. E. and J. D. Habila. 2020. Flavonoids: isolation, characterization, and health benefits. Beni-Suef University Journal of Basic and Applied Sciences. Volume 9, Article number: 45.DOI.org/10.1186/s43088-020-00065-9.

13. Feliciano R.P.; S. Pritzel ; C. Heiss and A. Rodriguez-Mateos. 2015. Flavonoid intake and cardiovascular disease risk. Curr. Opin. Food Sci. 2:92–99. DOI.org/10.101p /j. cofs. 2015.02.006

14. Ghedira K.; P. Goetz. 2015. Géranium rosat: Pelargonium graveolens L'Hér. (Géraniaceae). Phytotherapie. 13(3). DOI:10.1007/s10298-015-0955-x.

15. Hammood E. K.; M. N. Khalaf and J. M. Naser. 2024. Effect of Adding *Malv a neglecta* 

L. Leaves Powder on the Sensory Properties of Laboratory Biscuits. IOP Conf. Series: Earth and Environmental Science1371 (2024) 062031 IOP Publishing. doi:10.1088/1755-1315/1371/6/062031.

16. Hammood E. K.; A. C. Saddam and J. M. Naser. 2024. Improving nutritional and qualitative properties of wheat bread by using mallow (*malva neglecta* L.) leaves powder. Iraqi Journal of Agricultural Sciences. 55 (1): 560-568. <u>https://doi.org/10.36103/8p73pr77</u>

17. Hsouna A. B. and N. Hamdi .2012. Phytochemical composition and antimicrobial activities of the essential oils and organic extracts from *pelargonium graveolens* growing in Tunisia. Lipids in Health and Disease. 11: 167-172. doi: 10.1186/1476-511X-11-167.

18. Jasim A. S. and J. M. Nasser. 2024. Antidiabetic effect of green -synthesized silver nanoparticles using beta-glucan extract in a streptozotocin-injected rat model. Bulgarian J. of Veterinary Med. 51 (3): 777-788. DOI:10.15547/bjvm.2024-0030.

19. Kuppusamya P.; K. D. Lee; C. E. Songc; S. Ilavenil; S. S. Mariadhas and K. C. Choi. 2018 . Quantification of major phenolic and flavonoid markers in forage crop Lolium multiflorum using HPLC-DAD. Revista Brasileira de Farmacognosia. V. 28, Issue 3, pp. 282-288.

https://doi.org/10.1016/j.bjp.2018.03.006

20. Kumar S. and A. K. Pandey.2013. Chemistry and Biological Activities of Flavonoids: An Overview. First published: 29 December. doi. org/ 10.1155 /2013 / 162750.

21.Lalli J. Y. Y. ; R.L. Van Zyl ; S.F. Van Vuuren and A.M. Viljoen. 2008. In vitro biological activities of South African Pelargonium (Geraniaceae) species . South African J. of Botany.V.74, Issue1, pp. 153-157.

https://doi.org/10.1016/j.sajb.2007.08.011

22. Laohakungit N.; O. Kerdchoechuen ; R. Kaprasob and F.B. Matta. 2017. Volatile flavor, antioxidant activity and physicochemical properties of enzymatic defatted sesame hydrolysates. Journal of Food Processing and Preservation, 41(4) : 13075. DOI.org/10.1111/jfpp.13075

23. Marie S.; A. A. Magid and L. V. Nazabadioko. 2020. Investigation of

Antioxidant and Elastase Inhibitory Activities of Geum urbanum Aerial Parts, Chemical Characterization of Extracts Guided by Chemical and Biological Assays. Natural Product Communications. V.15(3): pp. 1–9 https://doi.org/10.1177/1934578X20915307.

24. Mativandlela S. P. N.; N. Lall and J. J. M. Meyer . 2006 . Antibacterial , antifungal and antitubercular activity of the roots of Pelargonium reniforme (CURT) and Pelargonium sidoides (DC) (Geraniaceae) root extracts. South African Journal of Botany. Volume 72, Issue 2, Pp 232-237.

DOI.org/10.1016/j.sajb.2005.08.002.

25.Mizzi L.; C. Chatzitzika ; R. Gatt and V. Valdramidis.2020. HPLC Analysis of Phenolic Compounds and Flavonoids with Overlapping Peaks. Food Technol Biotechnol. Mar; 58(1) :12-19 . doi: 10.17113/ftb.58.01.20.6395.

26. Mnif W.; W. Dhifi ; N. Jelali and H. Baaziz. 2013. Characterization of Leaves Essential oil of Pelargonium graveolens Originating from Tunisia: Chemical Composition, Antioxidant and Biological Activities. Journal of Essential Oil Bearing Plants 14(6):761-769.

DOI:10.1080/0972060X.2011.10644001.

27. Mutha R. E. ; A. U. Tatiya and S. J. Surana. 2021. Flavonoids as natural phenolic compounds and their role in therapeutics: an overview. Future Journal of Pharmaceutical Sciences 7(1). pp.25-36.

DOI:10.1186/s43094-020-00161-8

28. Nawaz H.; M. A. Shad; N. Rehman and H. Andaleeb. 2020. Effect of solvent polarity on extraction yield and antioxidant properties of phytochemicals from bean (Phaseolus vulgaris) seeds. Braz. J. Pharm. Sci. 56. DOI:10.1590/s2175-97902019000417129.

29. Pradeepa M; V. Kalidas and N. Geetha. 2016. Qualitative and quantitative phytochemical analysis and bactericidal activity of Pelargonium graveolens L'Her. Int J App Pharm. 8(3):7-11. 30. Radovanović, B.; J. Mladenović; A. Radovanović; R. Pavlović and V. Nikolić. 2015. Phenolic composition, antioxidant, antimicrobial and cytotoxic activates of Allium porrum L. (Serbia) extracts. J. of Food and Nutrition Research.3 (9): 564-569. DOI:10.12691/JFNR-3-9-1.

31. Sanguri S.; S. Kapil.; P. Gopinathan; F. K. Pandey and T. Bhatnagar. 2012. Comparative screening of antibacterial and antifungal activities of some Weeds and medicinal plants leaf extracts: An in-vitro study. Elixir Appl. Botany 47 : 8903-8905.

32. Santhi R.; G. Lakshmi ; A. Priyadharshini and L. Anandargj. 2011. Phytochemical screening Nerium oleander leaves and Momordica Charentais leaves . Inter. Res. J. Pharm., 2 (1): 131 – 135.

33. Saraswathi J.; K. Venkatesh; N. Baburao; M. H. Hilal1 and A. R. Rani. 2011. Phytopharmacological importance of Pelargonium species. Journal of Medicinal Plants Research . 5(13): 2587-2598.

34. Sompaga S.; B. A. Jyothi ; S. Chekuri ; N. Baburao and R. R. Anupalli . 2016. Organic extracts of *Pelargonium graveolens*: Phenol content, anti-oxidant and anti-bacterial activities. Eur J Med Plants; 17(1): 1-8. DOI: 10.9734/EJMP/2016/29040

35. Ullah A.; M. Sidra ; L. B. Syed ; K. Noreen ; G. Lubna ; G. P. Benjamin ; E. Abdul-Hamid and J. Mariusz .2020. Important flavonoids and their role as a therapeutic agent. Molecules .Nov; 25(22): 5243.

doi: 10.3390/molecules25225243.