

Vol. 3, No.2, 2025

The effect of Chlorella vulgaris and Spirulina platensis on enhancing the active compounds in basil (Ocimum basilicum L.) seeds and their role in supporting human health

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Received: 10 June (2025), Accepted: 22 June. 2025. Published: 30 June. 2025

ABSTRACT

In light of the global trend towards organic and sustainable agriculture, there has been an increasing interest in using microalgae as bio-stimulants and natural fertilizers as alternatives to chemical fertilizers. Algae such as Chlorella vulgaris and Spirulina platensis are considered rich sources of bioactive compounds that promote plant growth and stimulate the production of active substances. This study aims to evaluate the effect of adding microalgal extracts (Chlorella vulgaris and Spirulina platensis) on the growth of basil plants (Ocimum basilicum L.) and to improve their content of active compounds. Taking place in 2024 in Alhindia city, Kerbala, Iraq, The aforementioned experiment was set up using a Randomized Complete Block Design (RCBD) with three replications and a factorial arrangement. Two components were used in the study: Spirulina platensis algae (control, 100, 200, 250 mL.L⁻¹) and Chlorella vulgaris (control, 100, 200, 250 mL.L⁻¹ 1). After the active chemical components were measured, the plants were harvested when they reached full maturity. The results show that the effects of the spray treatments with the extracts of Spirulina platensis and Chlorella vulgaris mL.L-1 resulted in notable variations in the active component content. Camphor, Linalool, Pinene, and Myercene mg g⁻¹ at concentration (250 mL. L⁻¹ ¹) had the greatest averages of 2.97, 3.31 mg g⁻¹, 23.22, 24.97 mg g⁻¹, 0.23, 0.27 mg g⁻¹, 8.29, and 8.53 mg g⁻¹, respectively.

Keywords: Chlorella vulgaris, Spirulina platensis, Basil, supporting human health.

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Introduction

lgae are microscopic organisms that play an important role in many marine and freshwater ecosystems, and they have wide applications in various fields, including sustainable agriculture. Among these algae, spirulina and chlorella are considered wellknown species that contain a rich array of nutrients and active compounds, making them excellent natural sources for stimulating plant growth and improving soil fertility [1] Basil (Ocimum basilicum) is one of the famous aromatic plants used in many culinary and medicinal applications [2] Basil is considered an aromatic plant rich in bioactive chemical compounds with diverse biological activities. Its medicinal importance lies in its content of essential oils, which constitute the main element of its therapeutic properties, in addition to a wide range of phenolic compounds and flavonoids [3]. Essential oils are among the most important components of basil, and they include compounds such as Eugenol It has anti-inflammatory and pain-relieving properties [4], and is used in dentistry. Linalool Known for its calming and anti-anxiety effects. Methyl chavicol: It is characterized by antifungal and antibacterial properties [5]. Basil also contains compounds with high antioxidant capacity such Rosmarinic acid, Quercetin and Apigenin [6], In addition to its active compounds, basil contains: Vitamins like A, C, and K Minerals like calcium, iron, and magnesium Essential amino acid [7] Studies indicate that basil extracts contain a high percentage of antioxidants [8] that combat free radicals and reduce the risk of chronic diseases such as cancer and heart disease [9,10], which contribute to its antibacterial, antifungal, and antiviral properties. However, environmental conditions and agricultural practices can significantly affect the concentrations of these active compounds[11,12]. It focus on studying how the use of algae as plant nutrients affects the growth of basil and increases the production of effective aromatic compounds that contribute to improving its nutritional and medicinal quality [13]. It also algae affect the overall health of the soil and the environmental balance. Spirulina algae (Spirulina platensis) has emerged as one of the natural sources rich in proteins, vitamins, minerals, and essential amino acids, making it an important nutritional element in promoting overall health [14]. Spirulina is widely used as a dietary supplement due to its proven health benefits, such as boosting immunity and improving overall health. Additionally, recent studies have shown that the benefits of spirulina algae are not limited to humans but also extend to plants, where they have demonstrated positive effects on plant growth and increased production of active compounds in some medicinal and aromatic plants. The use of spirulina in sustainable agriculture may improve the quality of agricultural crops and increase Copyright 2025.





the concentration of active compounds in plants like basil, enhancing their medicinal and aromatic value [15]. This study aimed to highlight the environmental and economic benefits of using algae in sustainable agriculture, with a focus on improving the quality of aromatic crops such as basil using these natural ingredients.

METHODS

Taking place in 2024 in Alhindia city, Kerbala, Iraq, the current study sought to ascertain the effects of fertilizer including *Spirulina platensis* and *Chlorella vulgaris* algae on the secondary metabolism compounds of basil (*Ocimum basilicum* L.). The aforementioned experiment was set up using a Randomized Complete Block Design (RCBD) with three replications and a factorial arrangement. Two components were used in the study: *Spirulina platensis* algae (control, 100, 200, 250 mL.L⁻¹) and Chlorella vulgaris (control, 100, 200, 250 mL.L⁻¹). After the active chemical components were measured, the plants were harvested when they reached full maturity [16]. Twelve kilograms of soil were placed in plastic pots measuring 32 cm in diameter and 50 cm in height, using soil samples from the city of Karbala. On March 19, 2024, 40 basil seeds were sown in plastic pots, with each seed being buried 2 cm below the surface.

Essential oil extraction method

The basil seeds were extracted after the flower heads dried and turned brown Using a Soxhlet apparatus, Put the ground basil seeds inside the thimble, with the organic solvent hexane at a concentration of 70% so that it covers the plant material (basil seeds). Heat the solvent using a heat source, allowing the solvent to extract the essential oil from the seeds. This repetition (cycle) continues for several hours until the maximum amount of oil is extracted from the basil seeds .Separating the oil from the solvent, the oil is easily separated from the solvent by using evaporation apparatuses [17]. The extracted oil is stored in dark-colored bottles to protect it from light and heat. It is preferable to store it in a cool, dry place to maintain its quality, After the oil was stored in the refrigerator, it was taken to the Center for Science and Technology to examine the active compounds (Camphor, Linalool, Pinene, and Myercene mg. g⁻¹) using Gas chromatography mass-spectrometer (GC-MS) [18].



Statistical analysis

All the recorded data were analyzed according to randomized complete block design with factorial arrangement and three replications. The Least Significant difference (LSD_{0.05}) test was also used for comparison and separation of means [19].

RESULTS AND DISCUSSION

Camphor mg. g⁻¹

The results Indicate in Table (1) that there are significant differences in the content of the active compound Camphor (mg g⁻¹)due to the effect of the spray treatments with the extract *Chlorella vulgaris* mL.L⁻¹. The spray treatment (250 mL.L⁻¹) achieved the highest average of 3.31 mg g⁻¹, followed by the spray treatments 200 and 100, which had averages of 2.87and 2.28 mg g⁻¹, respectively. Meanwhile, the control plants achieved the lowest average of 2.14 mg g⁻¹. And observe from the same table that the spray treatments with fertilizer *Spirulina platensis* mL.L⁻¹ significantly outperformed the control, with a concentration of (250 mL. L⁻¹) achieving the highest average for the trait at 2.97 mg g⁻¹, while the 200 and 100 mL .L⁻¹ treatments recorded the lowest average in the leaf content of Camphor at 2.73, 2.51 mg g⁻¹, compared to the control treatment 2.38 mg g⁻¹, respectively. As observe from the results, there is a significant interaction. The plants treated with the (*Spirulina platensis* + *Chlorella vulgaris* 250 mL. L⁻¹) interaction recorded the highest average of 3.93 mg g⁻¹ compared to the plants treated with the (*Spirulina platensis* + *Chlorella vulgaris* control mL.L⁻¹) interaction, which recorded the lowest average of 2.02 mg g⁻¹.

Table 1. Effects Chlorella vulgaris and Spirulina platensis algae on the Camphor mg g⁻¹.

Chlorella vulgaris mL.L ⁻¹	Spirulina platensis mL.L ⁻¹				Means
	0	100	200	250	Means
Control	2.02	2.12	2.18	2.24	2.14
100	2.13	2.19	2.29	2.51	2.28
200	2.54	2.75	2.99	3.23	2.87
250	2.86	2.99	3.47	3.93	3.31
Means	2.38	2.51	2.73	2.97	

 $LSD_{0.05}$ Chlorella vulgaris = 1.76, $LSD_{0.05}$ Spirulina platensis = 1.53, $LSD_{0.05}$ Interaction =1.58



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Linalool mg. g⁻¹

Based on the results in Table (2), that there are significant differences in the content of the active compound Linalool (mg. g⁻¹) due to the effect of the spray treatments with the extract *Chlorella vulgaris* mL.L⁻¹. The spray treatment (250 mL. L⁻¹) achieved the highest average of 24.97 mg g⁻¹, followed by the spray treatments 200 and 100 mL. L⁻¹, which had averages of 21.93 and 20.42 mg g⁻¹, respectively. Meanwhile, the control plants achieved the lowest average of 18.09 mg g⁻¹. And observe from the same table that the spray treatments with fertilizer *Spirulina platensis* mL.L⁻¹ significantly outperformed the control, with a concentration of (250 mL.L⁻¹) achieving the highest average for the trait at 23.22 mg g⁻¹, while the 200 and 100 mL. L⁻¹ treatments recorded the lowest average in the leaf content of Linalool at 21.93, 20.89 mg g⁻¹, compared to the control treatment 19.40 mg g⁻¹, respectively. As observe from the results, there is a significant interaction. The plants treated with the (*Spirulina platensis* + *Chlorella vulgaris* 250 mL. L⁻¹) interaction recorded the highest average of 26.84 mg g⁻¹ compared to the plants treated with the (*Spirulina platensis* + *Chlorella vulgaris* control mL.L⁻¹) interaction, which recorded the lowest average of 16.75 mg. g⁻¹.

Table 2. Effects Chlorella vulgaris and Spirulina platensis algae on the Linalool mg g⁻¹

Chlorella vulgaris mL.L ⁻¹	Spirulina platensis ml.L ⁻¹				
	control	100	200	250	Means
Control	16.75	17.48	18.58	19.56	18.09
100	18.64	19.56	20.74	22.74	20.42
200	19.56	21.76	22.67	23.74	21.93
250	22.56	24.76	25.73	26.84	24.97
Means	19.40	20.89	21.93	23.22	

 $LSD_{0.05}$ *Chlorella vulgaris* =1.75, $LSD_{0.05}$ *Spirulina platensis* 1.48=, $LSD_{0.05}$ Interaction =1.94

Pinene mg. g⁻¹

Based on the results in Table (3), that there are significant differences in the content of the active compound Pinene mg. g⁻¹ due to the effect of the spray treatments with the extract *Chlorella vulgaris* mL.L⁻¹. The spray treatment (250 mL. L⁻¹) achieved the highest average of 0.27 mg g⁻¹, followed by the spray treatments 200 and 100, which had averages of 0.24 and 0.19 mg g⁻¹, respectively. Meanwhile, the control plants achieved the lowest average of 0.15 mg g⁻¹, and Copyright



observe from the same table that the spray treatments with fertilizer *Spirulina platensis* mL.L⁻¹ significantly outperformed the control, with a concentration of (250 mL. L⁻¹) achieving the highest average for the trait at 0.26 mg g⁻¹, while the 200 and 100 treatments recorded the lowest average in the leaf content of pinene at 0.23, 0.20 mg g⁻¹, compared to the control treatment 0.17 mg g⁻¹, respectively. As observe from the results, there is a significant interaction. The plants treated with the (*Spirulina platensis* + *Chlorella vulgaris* 250 mL. L⁻¹) interaction recorded the highest average of 0.31mg .g⁻¹ compared to the plants treated with the (*Spirulina platensis* + *Chlorella vulgaris* control mL.L⁻¹) interaction, which recorded the lowest average of 0.13 mg g⁻¹.

Table 3. Effects Chlorella vulgaris and Spirulina platensis algae on the pinene mg g⁻¹.

Chlorella vulgaris mL.L ⁻¹	Spirulina platensis mL.L ⁻¹				
	control	100	200	250	Means
Control	0.13	0.14	0.16	0.19	0.15
100	0.15	0.17	0.20	0.25	0.19
200	0.19	0.22	0.27	0.29	0.24
250	0.23	0.28	0.29	0.31	0.27
Means	0.17	0.20	0.23	0.26	

LSD_{0.05} *Chlorella vulgaris* =2.76, LSD_{0.05} *Spirulina platensis* 2.86=, LSD_{0.05} Interaction =0.73

Myercene mg g⁻¹

Based on the results in Table (4), that there are significant differences in the content of the active compound Myercene mg .g⁻¹ due to the effect of the spray treatments with the extract *Chlorella vulgaris* mL.L⁻¹. The spray treatment (250 mL.L⁻¹) achieved the highest average of 8.53 mg g⁻¹, followed by the spray treatments 200 and 100 mL. L⁻¹, which had averages of 7.57and 6.19 mg g⁻¹, respectively. Meanwhile, the control plants achieved the lowest average of 4.44 mg g⁻¹. And observe from the same table that the spray treatments with fertilizer *Spirulina platensis* mL.L⁻¹ significantly outperformed the control, with a concentration of (250 mL. L⁻¹) achieving the highest average for the trait at 8.29 mg g⁻¹, while the 200 and 100 mL. L⁻¹ treatments recorded the lowest average in the leaf content of Myercene at 7.15, 6.13 mg g⁻¹, compared to the control treatment 5.17

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mg g⁻¹, respectively. Based on from the results, there is a significant interaction. The plants treated with the (*Spirulina platensis* mL.L⁻¹ + *Chlorella vulgaris* mL.L⁻¹ 250 mL. L⁻¹) interaction recorded the highest average of 10.12mg .g⁻¹ compared to the plants treated with the (*Spirulina platensis* mL.L⁻¹ + *Chlorella vulgaris* control) interaction, which recorded the lowest average of 3.75 mg g⁻¹.

Table 4. Effects Chlorella vulgaris and Spirulina platensis algae on the Myercene mg g⁻¹.

Chlorella vulgaris mL.L ⁻¹	Spirulina platensis mL.L ⁻¹				Maana
	control	100	200	250	Means
Control	3.75	3.96	4.23	5.84	4.44
100	4.85	5.38	6.83	7.73	6.19
200	5.76	6.74	8.34	9.45	7.57
250	6.34	8.43	9.23	10.12	8.53
Means	5.17	6.13	7.15	8.29	

 $LSD_{0.05}$ Chlorella vulgaris = 2.87, $LSD_{0.05}$ Spirulina platensis 2.67=, $LSD_{0.05}$ Interaction = 1.65

DISCUSSION

Microalgae such as *Chlorella vulgaris* and *Spirulina platensis* are considered rich sources of active compounds that contribute to enhancing plant health. When added to the soil or used as fertilizer, these algae may affect plant growth and the formation of their active compounds [20]. *Chlorella vulgaris* shows the presence of active biological compounds such as phenols, terpenoids, and alkaloids, which are known for their antioxidant and antibacterial properties. For example, Chlorella contains the compound β -1,3-glucan, which is considered a strong immunostimulant and has anti-tumor and cholesterol-lowering effects [21]. As for *Spirulina platensis*, it contains active compounds such as phenols and flavonoids, which exhibit antioxidant and anti-diabetic effects. For example, Spirulina extracts have shown inhibitory activity against the enzyme α -glucosidase, which contributes to reducing blood sugar levels [22]. When these algae are applied to basil plants, the active compounds present in the algae may contribute to enhancing the plant's growth and improving the quality of its active compounds. For example, the content of basil in antioxidant compounds such as phenols and flavonoids may be improved, enhancing its medicinal and nutritional properties [23]. These studies show that the use of *Spirulina platensis* may be more



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effective than Chlorella vulgaris in enhancing the active compounds in basil, especially concerning phenolic and flavonoid compounds and antioxidant activity. However, it is recommended to conduct practical experiments to determine the appropriate doses and the best application methods to ensure optimal results [24,25].

CONCLUSIONS

Studies show that using *Chlorella vulgaris* and *Spirulina platensis* as agricultural additives may improve the quality of active compounds in basil, contributing to enhancing its health benefits. However, it is recommended to conduct practical experiments to determine the appropriate dosages and the best application methods to ensure optimal results.

Acknowledgments

The authors wish to thank to all persons which supported this work in the College of Education for Pure Sciences, College of Nursing and, College of Agriculture Science staff at the University of kerbala.

REFERENCES

- 1. El-Kheir, A.F., El-Ansary, M.A., El-Sayed, S.M. and Khalifa, S.M., 2023. *Effect of Spirulina platensis extract on the growth and antioxidant activity of Ocimum basilicum L. under greenhouse conditions*. Journal of Applied Research on Medicinal and Aromatic Plants, 30, p.100434. Available at: https://doi.org/10.1016/j.jarmap.2023.100434.
- 2. Al-Yasssiry, A.S., Aljenaby, H.K.A., Al-Masoody, I.H. and Al-Ibrahemi, N., 2024. *Biofertilizers effect on the active compounds of sweet basil (Ocimum basilicum L.)*. SABRAO Journal of Breeding and Genetics, 56(1), pp.0–0.
- 3. Al-Ibrahemi N, Al-Yassiry A, AL-Laith ZN, Al-Musawi BH (2023). Phytochemical study of volatile oils for the Ocimum basilicum L. and mentha spicata by gas chromatography technique. IOP Conf. Ser.: Earth Environ. Sci. 1158 (1):1-8.
- 4. Murarikova A A, Tazk J, Neugebauerova A, Plankova J, Jampilek P, Muicaji, Mikus P (2017). Characterization of essential oil composition in \ 140 different basil species and pot cultures by a GC-MS method. Molecules, 22, 1221.



- 5. Larimi S B, Mohammadreza S, Adel DM, Mohammad M V (2014). Changes in nitrogen and chlorophyll density and leaf area of sweet basil (Ocimum basilicum L.) affected by biofertilizer and nitrogen application. Int. J. Biosciences, 5(9),256-265.
- 6. Banchio E, Xie X, Zhang H, Par PW (2009). Soil bacteria elevate essential oil accumulation and emissions in sweet. J. Agric. Food. Chem. 57: 653–657.
- 7. Abdel-Hamid AN (2020). Effect of benzyl adenine, indole acetic acid, and gibberellic acid on vegetative growth, chemical constituents, and volatile oil attributes of sweet basil plants. Egyptian J. Hortic. 47(1): 41-56.
- 8. Adigzel A.; G. L L.CE M., Peng.L, M, H, Ú.TC. H., Fikrettin Pahun, Karaman. Ü.(2005). Antimicrobial Effects of Ocimum basilicum (Labiatae) Extract .Turk. J. Biol., 29:155-160.
- 9. Ababutaim . I.M. (2019). Antimicrobial Activity and Gas Chromatography-Mass Spectrometry (GCMS) Analysis of Saudi Arabian Ocimum basilicum Leaves Extracts .J Pure Appl Microbiol, 13(2), 823-833 .
- 10. Nyarko, A. K.(2002). Extract of Ocimum canum lowers blood glucose and facilitates insulin release by isolated pancreatic beta islet cells. Phytomedicin. 9 (4): 346-351.
- 11. Alakashy, Z.H. and Al-Bedairi, N., 2020a. *Growth and yield response of three basil (Ocimum basilicum L.) varieties to different nitrogen sources*. Plant Archives, 20(1), pp.398–401.
- 12. Alakashy, Z.H. and Al-Bedairi, N., 2020b. *Effect of three varieties of basil and the nitrogen source in essential oil contents*. European Journal of Agricultural Sciences, 12(1), pp.1–6.
- 13. Abd El-Monem, A.A., El-Fallal, A.A., Ghoname, A.A. and Ragab, M.E., 2022. *Biostimulant potential of Chlorella vulgaris on the physiological parameters and essential oil content of sweet basil (Ocimum basilicum L.)*. Egyptian Journal of Agronomy, 44(2), pp.207–217.
- 14. Safi, C., Zebib, B., Merah, O., Pontalier, P.Y. and Vaca-Garcia, C., 2014. Morphology, composition, and antioxidant capacity of Spirulina platensis powders prepared using different drying methods. Food Chemistry, 153, pp.366–373. Available at: https://doi.org/10.1016/j.foodchem.2013.12.066.
- 15. Khalifa, A.Y., El-Awady, M.E. and Youssef, S.A., 2021. Evaluation of Chlorella vulgaris and Spirulina platensis as biofertilizers and their impact on basil plant (Ocimum basilicum L.) growth and phytochemicals. Environmental and Experimental Biology, 19(3), pp.455–462. Available at: https://doi.org/10.22364/eeb.19.34.



- 16. Al-Masaoodi, N.H., Al-Ibrahemi, N., Abdulammar, S.H. and Al-Yassiry, A.S., 2025. Fenugreek (Trigonella graecum L.) response to Azotobacter and orange peels in metabolism, growth, and yield traits. SABRAO Journal of Breeding and Genetics, 57(1), pp.0–0.
- 17. Al-Ibrahemi, N., Al-Laith, Z.N., Al-Yasssiry, A.S. and Al-Masaoodi, N.N., 2022. *Chemical analysis of phytochemicals for the Anethum graveolens L. fresh and commercial dry by gas chromatography mass-spectrometer*. IOP Conference Series: Earth and Environmental Science, 1069(1), pp.1–10.
- 18. Al-Ibrahemi, N., Al-Yassiry, A., Al-Laith, Z.N. and Al-Musawi, B.H., 2023. *Phytochemical study of volatile oils for the Ocimum basilicum L. and Mentha spicata by gas chromatography technique*. IOP Conference Series: Earth and Environmental Science, 1158(5), pp.1–8.
- 19. Al-Ibrahemi, N., Al-Asadi, Q.T.H.Y., Hassan, S.F., Hamid, B.A. and Jawad, N.N., 2024. *Response of flax (Linum usitatissimum) to Nano-NPK and EMG-1 in growth, oil content, and active compounds.* SABRAO Journal of Breeding and Genetics, 56(6), pp.2481–2487. Available at: https://doi.org/10.54910/sabrao2024.56.6.29.
- 20. Torres-Duran, P.V., Ferreira-Hermosillo, A. and Juarez-Oropeza, M.A., 2007. *Antihyperlipemic and antihypertensive effects of Spirulina maxima in an open sample of Mexican population: a preliminary report*. Lipids in Health and Disease, 6(1), p.33. Available at: https://doi.org/10.1186/1476-511x-6-33.
- 21. Antelo, F.S., Costa, J.A.V. and Kalil, S.J., 2008. *Thermal degradation kinetics of the phycocyanin from Spirulina platensis*. Biochemical Engineering Journal, 41(1), pp.43–47. Available at: https://doi.org/10.1016/j.bej.2008.03.012.
- 22. Koyande, A.K., Chew, K.W., Rambabu, K., Tao, Y., Chu, D.T. and Show, P.L., 2019. *Microalgae: A potential alternative to health supplementation for humans*. Food Science and Human Wellness, 8(1), pp.16–24. Available at: https://doi.org/10.1016/j.fshw.2019.03.001.
- 23. Neyrinck, A.M., Taminiau, B., Walgrave, H., Daube, G., Cani, P.D., Bindels, L.B. and Delzenne, N.M., 2017. *Spirulina protects against hepatic inflammation in aging: an effect related to the modulation of the gut microbiota?* Nutrients, 9(6), p.633. Available at: https://doi.org/10.3390/nu9060633.
- 24. Fallah, A.A., Sarmast, E., Dehkordi, S.H., Engardeh, J., Mahmoodnia, L., Khaledifar, A. and Jafari, T., 2018. *Effect of Chlorella supplementation on cardiovascular risk factors: a meta-*





- *analysis of randomized controlled trials*. Clinical Nutrition, 37(6), pp.1892–1901. Available at: https://doi.org/10.1016/j.clnu.2017.09.019.
- 25. Udayan, A., Arumugam, M. and Pandey, A., 2017. *Nutraceuticals from algae and cyanobacteria*. In: Rastogi, R., Madamwar, D. and Pandey, A. (eds.) Algal Green Chemistry. Amsterdam: Elsevier, pp.65–89.

