

9. Armitage GC. Clinical evaluation of periodontal disease. *Periodontol* 2000 1995;7:39-53.
10. Tantivanich S, Laohapand P, Thawe - boon S. et al. Prevalence of cytomegalovirus, human herpesvirus-6, and Epstein-Barr v - rus in periodontitis patients and healthy su - jects in the Thai population. *Southeast Asian J Trop Med Public Health* 2004; 35(3):635-40.
11. Imbronito AV, Okuda OS, Maria de Freitas et al. Detection of herpesviruses and periodontal pathogens in subgingival plaque of patients with chronic periodontitid, ge - neralized aggressive periodontitis, or gingiv - tis. *J Periodontol*. 2008; 79: 2313-2321.
12. Bilder L, Elemelech R, Szwarcwort-Cohen M, Kra-Oz Z, Machtei EE. The prev - lence of human herpes viruses in the saliva of chronic periodontitis patients compared to oral health providers and healthy controls. *Arch Virol*. 2013; 158(6):1221-6.
13. Wu Y M, Yan J, Ojcius D M, Chen L L, Gu Z Y, Pan J P. Correlation between i - fections with different genotypes of human cytomegalovirus and Epstein-Barr virus in sub gingival samples and periodontal status of patients. *J Clin Microbiol*. 2007; 45: 3665-3670.
14. Sunde TP, Olsen I, Enersen M, Beiske K and Grinde B. Human Cytomegalovirus and Epstein-Barr Virus in Apical and Marginal Periodontitis: A Role in Pathology?. *Journal of Medical Virology*. 2008; 80:1007-1011.
15. Ling L J, Chuan-Chen H, Wu CY, et al. Association between human herpesviruses and severity of periodontitis. *J Periodontol* 2004; 75(11): 1479-85.
16. Iigeh S. EBV and CMV in chronic periodontitis: a prevalence study. *Archives of Virology* 2008; 153(10):1917-9.
17. Saygun I, Kubar A, Özdemir A, Slots J. Periodontitis lesions are a source of salivary cytomegalovirus and Epstein-Barr virus. *J Periodontol Res* 2005; 40: 187-191.
18. Cohen J. I. Epstein-Bar virus infe - tion. *N. J. Med*. 2000; 343: 481-492.
19. Reeves M B, MacAry P A, Lehner PJ, Sissons JG, and Sinclair JH. Latency, chr - matin remodeling, and reactivation of h - man cytomegalovirus in the dendritic cells of healthy carriers. *Proc. Natl. Acad. Sci*. 2005; 102: 4140-4145.
20. Slots J, Contreras A. Herpesviruses: a unifying causative factor in periodontitis? *Oral Microbiol Immunol* 2000; 15: 276-279.
21. Slots J. Human viruses in periodont - tis. *Periodontol* 2000. 2010; 53:89-110.

tion between the presence of CMV and BOP. Saygun et al [17] found that there was no statistical correlation between HCMV counts in periodontal pockets and saliva. Tantivaich et al [10] revealed that PPD not associated with the presence of HCMV. HCMV represent some of the most successful virus in humans, infecting over 90% of humans and persisting for the lifetime of the individuals [18, 19]. This data suggest that HCMV is usually present in the body in inactive state. Reactivation of it in periodontal sites comprises an important pathogenic event in the development of periodontitis [20]. A solid understanding of the etiology of periodontitis is critical for developing clinically relevant classification systems and therapies that can ensure long-lasting disease control. Research during the past 15 years has implied that herpesviruses are involved in the etiopathogeny of destructive periodontal disease. It appears that latent herpesviruses infections are found in chronic periodontitis. Also, specific genotypes of herpesvirus species may exhibit increased periodontopathic potential. Herpesviruses especially HCMV are probably not stand-alone periodontopathic agents, but cooperate with specific bacteria in periodontal tissue breakdown. A co-infection of active herpesviruses and periodontopathic bacteria may constitute a major cause of periodontitis and explain a number of the clinical characteristics of the disease [21].

References:

1. Novak JM and Novak FK. Chronic periodontitis In: NEWMAN, TAKEI, KLOKEVOLD, CARRANZA (eds.) CLINICAL PERIODONTOLOGY 10th ed. ELSEVIER, China. 2009; 494.
2. Kinane FD, Lindhe J, Trombelli L. Chronic Periodontitis In: Jan L, Thorkild K, Niklaus P. L. (eds.) Clinical Periodontology and Implant Dentistry. 5th ed. Munksgaard, Copenhagen. 2008 ; 420-426.
3. Contreras A, Zadeh HH, Nowzari H, Slots J. Herpesviruses in human periodontal disease. J Periodontal Res 2000; 35:3-16.
4. Slots J. Herpesviruses in periodontal diseases. Periodontol 2000; 2005; 38: 33-62.
5. Saygun I, Yapar M, Özdemir A, Kubar A, Slots S. Human cytomegalovirus and Epstein-Bar virus type I in periodontal abscesses. Oral Microbiol Immunol 2004; 19: 1437-1443.
6. Slots J. Interactions between herpesviruses and bacteria in human periodontal disease. In: Brogden KA, Guthmiller JM, editors. Polymicrobial Diseases. Washington, DC: ASM Press. 2002; 317-331.
7. Slots J. Herpesviral-bacterial interactions in periodontal diseases. Periodontol 2000 2010; 52:117-140.
8. Thylstrup, A. and Fejerskov, O. Textbook of Clinical Cariology. 2nd ed., Munksgaard, 1996. Vol. 17. Pp: 17-43.

ment onto the root surfaces accompanied by loss of connective tissue and alveolar bone [9]. Concerning PCR results, the percentage of HCMV DNA positive of chronic

periodontitis patients was 31.2% while in control subjects was 13.3%. Tantivanich et al [10] detected HCMV in 34% of chronic periodontitis patients and 3.3% in healthy group. Imbronito et al found HCMV in 75% of chronic periodontitis patients [11]. Another study done by Bilder et al [12] where they detected HCMV in 15% of subjects with chronic periodontitis and 0% in healthy group. The reasons for variation in HCMV occurrence among studies may include difference in detection technique, samples and population. In this study there was no significant difference between chronic periodontitis and healthy subjects regarding the presence of HCMV DNA. This is agree with Wu et al [13] who found that there was no significant difference in presence of HCMV DNA between chronic periodontitis and healthy subjects and disagree with Tantivanich et al [10] who found the percentage of HCMV DNA occurrence higher in chronic periodontitis than in healthy groups. Most herpesviruses are ubiquitous agents that often are acquired early in life and infect individuals from diverse geographic areas and economic backgrounds [4]. Transmission of HCMV can happen vertically either prenatally or perinatally, from mother

to infant, or horizontally in children or adults by direct or indirect person-to-person contact [4]. HCMV are present latently in the vast majority of the adult population. HCMV is present in a greater variety of cells including various epithelial cells, endothelial cells, and leukocytes and it is active periodically and shed viral particles to the saliva [14]. The present study revealed that there is no significant difference in the presence of HCMV DNA in study groups between males and females. Tantivanich et al [10] found that there was no association between gender and HCMV infection among periodontitis patients and the healthy control group and females had higher rate of HCMV than males in the same study [10]. Ling et al [15] showed that the presence of HCMV equally distributed between women and men. In this study there was no relation found between the presence of HCMV DNA and periodontal parameters (PLI, GI, BOP, and PPD) in chronic periodontitis patients. Regarding CAL there was a significant relation between the presence of HCMV DNA and CAL scale 3. Ling et al [15] found the presence of HCMV was significantly higher in periodontitis group that had lower PLI. Also they showed that coinfection of HCMV was significantly associated with higher GI and BOP, coinfection of HCMV was also associated with higher PPD and CAL [15]. Ligehe [16] found that there was no associ-

Table 5: Results of PCR for CMV in chronic periodontitis & control groups regarding gender.

CMV PCR results Chronic Periodontitis Controls						
	Male	Female	No	Male	Female	No
	No	%		%	No	%
No						
1	Positive		5	35.7	5	27.8
	12.5	1	14.3			
7	Negative		9	64.3	13	72.2
	87.5	6	85.7			
	P value		0.631	0.919		

×Non- significant (P≥0.05).

30.9±32.2			
P.D. Scale 0	10	80.2±17.5	22
88.4±12.1			
P.D. Scale 1	10	12.0±10.1	22
8.4±5.0			
P.D. Scale 2	10	1.8±3.3	22
0.7±1.5			
C.A.L. Scale 0	10	70.0±23.6	22
83.0±14.7			
C.A.L. Scale 1	10	5.9±7.6	22
6.6±7.4			
C.A.L. Scale 2	10	12.1±12.7	22
6.5±4.7			
C.A.L. Scale 3	10	6.1×±10.0	22
1.5±2.5			
No. of sites	10	94.0±10.9	22
97.5±12.7			

×Significant(P<0.05)

Table 6: The relationship between the presence of CMV DNA and clinical periodontal parameters; PI, GI, BOP, PPD, CAL & No. of sites in chronic periodontitis.

Chronic Periodontitis				
CMV PCR positive CMV PCR negative				
No	Mean±SD	No	Mean±SD	
Mean P.I	10 1.5±0.4	22		
1.5±0.3				
Mean G.I	10 1.2±0.3	22		
1.3±0.4				
B.O.P. Score 0	10 73.4±26.8	22		
66.6±27.7				
B.O.P. Score 1	10 20.6±24.7	22		

Discussion:

In this study, a conventional PCR method was designed to detect HCMV DNA in saliva samples. This method provided a sensitivity sufficient to allow meaningful data to be obtained from a small volume of saliva taken from study groups. In this study there was highly significant difference in mean of PLI and GI between chronic periodontitis and control groups, this is clear because plaque induced periodontitis is the presence of gingival inflammation at sites where there has been apical migration of the epithelial attac -

groups (mean & SD) regarding; PLI, GI, BOP, PPD, CAL & No. of sites.

	Chronic Periodontitis	Controls
P value		
Mean PLI	1.5±0.3 (1.00-2.26)	0.7 ± 0.4 (0.08-1.10)
	0.0001*	
Mean G.I	1.3±0.4 (1.02-3.00)	0.8 ± 0.4 (0.05-1.04)
	0.0001*	
B.O.P. Score 0	68.7±27.2 (0-105)	-
B.O.P. Score 1	27.7±30.0 (2-112)	-
P.D. Scale 0	85.8±14.2 (42-106)	-
P.D. Scale 1	9.5±7.1 (3-33)	-
P.D. Scale 2	1.0±2.2 (0-9)	-
C.A.L. Scale 0	79.0±18.6 (30-106)	-
C.A.L. Scale 1	6.4±7.3 (0-26)	-
C.A.L. Scale 2	8.3±8.3 (0-32)	-
C.A.L. Scale 3	2.9±6.2 (0-27)	-
No. of site	96.4±12.1 (76-112)	106.1 ± 8.1 (84-112)
	0.007*	

* * Highly significant (<0.01)

	Chronic Periodontitis		Controls	
P value				
	No	%	No	%
CMV PCR results	Positive		10	31.2
2	13.3	0.189		

Negative 22 68.8 13 86.7

*Non-significant (P>0.05).

Table 4: Results of PCR for CMV in chronic periodontitis & control groups.

Figure 1: Forty – seven of DNA samples were extracted from saliva of study groups using Noragen extraction method. (M) means 6µl of Kapa ladder and the other samples 9µl of (1-47) Saliva DNA samples were loaded in 1% Agarose gel and electrophoresed for 45 minutes on 70 Volt.

Figure 2: Electrophoresis gel of 2% agarose showing amplification product (287bp) of G protein-coupled receptor (US28) gene using conventional PCR under electrophoresis condition(100volt) and 50 A with 0.5 X of TAE buffer for 45 minutes. Twelve PCR products of HCMV-US28 gene were generated by conventional PCR.

Figure-3: Alignments of primers with US28 gene of HCMV showing amplification size 287bp in this genetic map for the PCR purpose.

number and percentage of chronic periodontitis group who were positive for HCMV DNA were 10 (31.2%) respectively while in control group were 2 (13.3%) respectively. There was no significant difference between chronic periodontitis and control groups ($P=0.05$) regarding the presence of HCMV DNA. Figure 2 shows the results of conventional PCR of detection of HCMV DNA Lane (A) Kapa DNA ladder 6 μ l; Lanes 4, 8, 13, 17, 18, 23, 25, 28, 32, 34, 41, 43, 46 HCMV DNA positive saliva sample from study groups. Table 5 shows the results of PCR for HCMV DNA in chronic periodontitis & control groups regarding gender, there was no significant difference between male and female ($P=0.05$). Table 6 describes the relationship between the presence of CMV DNA and clinical periodontal parameters; PLI, GI, BOP, PPD, CAL & No. of sites in chronic periodontitis group. As seen in this table there was no significant difference between PLI, GI, BOP, PPD and CAL scales 0, 1, and 2 ($P=0.05$) and a significant difference was found between the presence of HCMV DNA and CAL scale 3 ($P<0.05$).

Table 1: Optimal condition for PCR and primers used in PCR for HCMV detection.

Temperature	Time	Cycle
Initial Denaturation	95 °C	3min
Denaturation	95 °C	18 sec

Annealing 54°C 20 sec 35

Polymerization 72°C 20sec

Final Extension 72°C 10min

Infinity 4°C

Forward primer A A C A G G C C T G -
CTTTTCAGT

Reverse primer A A A G A C A A G C A -
GACCGCTA

Table 2: General distribution of the study groups (no. percentage, mean & SD) regarding age & gender

	Chronic Periodontitis		Controls	
P value	No	%	No	%
Age (years)	20---29	7	21.9	5
	30---39	10	31.3	8
	=>40	15	46.9	2
	Mean \pm SD(Range)		36.4 \pm 7.2 (25-45)	
	31.6 \pm 6.3 (20-45)			
Gender	Male	14	43.8	8
			53.3	
	Female	18	56.3	7
			46.7	

×Non-significant ($P=0.05$)

Table 3: General distribution of the study

in HCMV genome. Amplification of DNA was performed with a total reaction volume (25 μ l). The reaction mixture was submitted to the PCR device with optimal conditions that are illustrated in table 1. The products were analyzed by using 1.5% agarose gel. 0.2 μ g Ethidium bromide and electrophoresed for 45 minutes. The gel was photographed under UV illumination.

Statistical Analysis of data was carried out using the available statistical package of SPSS-22 (Statistical Packages for Social Sciences- version 22). (Data were presented in simple measures of frequency, percentage, mean, standard deviation, and range (minimum-maximum values). The significance of difference of different means (quantitative data) were tested using Students-t-test for difference between two independent means or Paired-t-test for difference of paired observations (or two dependent means), or ANOVA test for difference among more than two independent means. The significance of difference of different percentages (qualitative data) were tested using Pearson Chi-square test with application of Yate's correction or Fisher Exact test whenever applicable. Pearson correlation was calculated for the correlation between two quantitative variables with its t-test for testing the significance of correlation. Statistical significance was considered whenever the P value for the test of significance was equal or less than 0.05.

Results.

Table 2 shows the distribution of study group by basic characteristics, according to age and gender. Also this table describes the mean, standard deviation (SD) and the range of plaque index (PLI), gingival index (GI) and number of sites in the study group. The mean and SD of age among periodontitis patients were (36.4 \pm 7.2) respectively, while the mean and SD of age in control group were (31.6 \pm 6.3) respectively. Table 3 shows the general distribution of the study groups (mean & SD) regarding; PLI, GI, BOP, PPD, CAL & No. of sites. As seen in this table the mean, SD and the range of PLI in chronic periodontitis were 1.5 \pm 0.3 (1.00-2.26) respectively while in control group were 0.7 \pm 0.4 (0.08-1.10) respectively. The mean, SD and the range of GI in chronic periodontitis patients were 1.3 \pm 0.4 (1.02-3.00) respectively while in control group were 0.8 \pm 0.4 (0.05-1.04) respectively. The mean, SD and the range of number of sites that measured in chronic periodontitis patients and control group were 96.4 \pm 12.1 (76-112) 106.1 \pm 8.1 (84-112) respectively. There was highly significant difference in mean PLI, GI and no. of sites between chronic periodontitis and healthy subjects ($P < 0.01$) with increase of them in chronic periodontitis patients. The results of conventional PCR for HCMV DNA was shown in table 4. As seen in this table the

was taking from the center (doctors, dentist, dental hygienists and dental assistants) (8 males and 7 females). they were free of any signs and symptoms of periodontal disease. All patients in this research were with no history of any systemic disease, had not received previous periodontal treatment. Subjects were excluded if pregnant and/or smokers.

Clinical examination.

Periodontal examination consisted of plaque index (PLI), gingival index (GI), bleeding on probing (BOP), probing pocket depth (PPD) and clinical attachment level (CAL) at 4 sites for all teeth except 3rd molar on (mesial, midvestibular, distal, midlingual), using a calibrated periodontal probe (Michigan O probe). Patients with chronic periodontitis had periodontal pockets equal or greater than 4mm with clinical attachment loss. For ease of estimation, a scale was designed to measure the PPD & it included the following scores: score 0 = 1-3 mm, score 1 = 4-5 mm, score 2 \geq 6 mm. Also another scale was designed to measure CAL as follows: score 0 = no attachment loss, score 1 = 1-2 mm, score 2 = 3-4 mm & score 3 \geq 5.

Collection of saliva samples

All participants were instructed not to eat or drink (except water) at least 1 hour prior to donation of saliva, the subject should sit in a relaxed position and samples containing

blood should be discarded. Saliva was collected between 9-12 am. After the subject rinse his mouth several times by sterilized water and then wait for 1-2 minutes for water clearance. 5ml of whole unstimulated mixed saliva was collected into polyethylene tubes using a standardized method [8]. Saliva then stored frozen at (-20 °C) until they were assayed.

Nucleic acid extraction

All samples were analyzed at the Department of Molecular biology, Iraqi Center for Cancer and Medical Genetics Research, Al-Mustansyriah University. Frozen saliva samples were allowed to thaw and come to room temperature. Therefore, they were subjected to DNA extraction. DNA was extracted from the saliva samples by using (Norgen's Saliva DNA Isolation Kit / Canada-RU45400).

PCR procedure.

A conventional PCR method was used to detect viral DNA from CMV. 2X robust 2G ready mix DNA polymerase kit was purchased from Kapa biosystem, South Africa. PCR primers were synthesized in Bioneer Company, South Korea as mentioned in table 1. DNA that was extracted from saliva samples by digesting with proteinase K solution and Dnase-free Rnase was used as template in PCR amplification for US 28 gene (G protein-coupled receptor) 287bp that is present

The clinical feature that distinguishes periodontitis from gingivitis is the presence of clinically detectable attachment loss [1]. Chronic periodontitis is the most common form of periodontitis. It is associated with the accumulation of dental plaque and calculus and generally has a slow-to-moderate rate of disease progression, but periods of more rapid destruction may be observed. Increases in the rate of disease progression may be caused by the impact of local, systemic, or environmental factors that may influence the normal host-bacteria interaction [2]. Even though specific infectious agents are of key importance in the development of periodontitis, it is unlikely that a single agent or even a small group of pathogens are the sole cause or modulator of this heterogeneous disease [3]. Since the mid-1990s, herpesviruses in particular, human cytomegalovirus (HCMV) have emerged as putative pathogens in various types of periodontal disease [4]. Genomes of HCMV occur at high frequency in chronic periodontitis [5, 6]. HCMV is the largest of genomic size, it possesses a high protein coding capacity, with estimates ranging from 160 to more than 200 open reading frames. The sequence of the HCMV genome has been known for over a decade [4]. The herpesviral-bacterial hypothesis of periodontitis proposes that an active herpesvirus infection initiates periodontal tissue breakdown and that host

immune responses against the herpesvirus infection are an important component of the etiopathogeny of the periodontitis [7]. Contemporary studies of periodontal viruses particularly HCMV have been employed high-performance polymerase chain reaction (PCR) techniques to determine the frequency of the viral genome. PCR-based studies of periodontal herpesviruses have targeted different genomic regions and used methods of different efficiency to extract the target nucleic acid [7]. Polymerase chain reaction (PCR) offers a rapid and relatively inexpensive method of identifying viral nucleic acids in clinical specimens [4]. The aim of this study is to detect HCMV in saliva of both chronic periodontitis and healthy subjects by conventional PCR.

Material and methods.

Human Sample.

Sample population consisted of forty seven males and females, age ranged from 20 to 45 years. Samples collection was started at 1st of September 2014 till February 2015. Subjects participating in the present study were informed about the purpose of the study & they agreed to participate. Patients with chronic periodontitis (14 males and 18 females) were screened at the Clinic of the Department of Dentistry in Primary health care center in AL-Saydia region in Al-Karkh district of Baghdad city. The control group

The present study observed that there was no relation between PLI and GI with HCMV DNA. Concerning BOP and PPD there was no relation found between BOP and PPD with HCMV DNA. For CAL there was a significant relation found between CAL scale 3 and HCMV DNA.

Keyword: HCMV, Chronic periodontitis, G protein-coupled receptor gene, Conventional PCR.

التحري عن الحامض النووي للراشح المضخم للخلايا في لعاب المرضى المصابين بالنساع المزمن والاشخاص الاصحاء بواسطة سلسلة تفاعل البلمرة التقليدية

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الخلاصة:

النساع المزمن هو من الاصابات الشائعة التي تسببها الفيروسية الجرثومية والذي يمتد الى الانسجة الساندة للاسنان. ويعتبر السبب الرئيسي لفقدان الاسنان في العالم. الاصابات الفيروسية ربما تسهل تدمير الانسجة الساندة للاسنان عن طريق النشاط التحللي ضد خلايا الانسجة الساندة للاسنان، تثبيط المناعة والذي يزيد من قابلية المضيف لهجوم البكتيريا. الهدف من هذه الدراسة هو التحري عن الحامض النووي للراشح المضخم للخلايا البشرية في لعاب المرضى المصابين بالنساع المزمن وشريحة ضابطة من الاصحاء بواسطة اختبار سلسلة تفاعل

وتحليل العلاقة (Conventional PCR) البلمرة التقليدية مع وضع الانسجة الساندة للاسنان. تكونت عينة الدراسة من اثنين وثلاثين مصابا بالنساع المزمن من كلي الجنسين (14 ذكرا و 18 انثى) وخمسة عشر شخصا من الاصحاء كعينة مسيطرة (8 ذكور و 7 اناث) وبمدى عمري يتراوح بين 25-45 سنة. كل المشاركين لايشكون من اي مرض جهازى. المعلومات السريرية لما حول الاسنان المذكورة في هذه الدراسة كانت، (مؤشر الصفيحة الجرثومية، المؤشر اللثوي، النزف بالمسبر الطبي، عمق الجيب المسبري والمستوى السريري الرابط). جمعت عينات اللعاب غير المحفز من كل الاشخاص المشاركين في البحث وفحصت باختبار سلسلة تفاعل البلمرة التقليدية للتحري عن الحامض النووي للراشح المضخم للخلايا البشرية. نتائج هذه الدراسة اظهرت ان نسبة المصابين بالنساع المزمن الذين اظهروا تفاعل ايجابي للحامض النووي للراشح المضخم للخلايا البشرية كانت (31.25%) وعند الاصحاء كانت (13.33%). هذه الدراسة بينت بانه لا توجد علاقة معنوية بين وجود الراشح عند المرضى المصابين بالنساع المزمن والاشخاص الاصحاء وانه لا يوجد فرق معني بين الذكور والاناث وبين وجود الراشح وانه لا توجد علاقة بين مؤشر الصفيحة الجرثومي، المؤشر اللثوي، النزف بالمسبر الطبي وعمق الجيب المسبري وبين المستوى العلابي للحامض النووي للراشح. بالنسبة الى المستوى السريري الرابط، وجدت هناك علاقة معنوية ايجابية بين مستوى الحامض النووي للراشح وبين المستوى السريري الرابط مقياس 3

Introduction:

Periodontitis is defined as “an inflammatory disease of the supporting tissues of the teeth caused by specific microorganisms or group of specific microorganisms, resulting in progressive destruction of the periodontal ligament and alveolar bone with increased probing depth formation, recession, or both.”

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Detection of HCMV DNA in Saliva of Chronic Periodontitis Patients and Healthy Subjects by Conventional PCR

Abstract:

Chronic periodontitis is a common plaque-induced periodontal infection that is a major cause of teeth loss throughout the world. Viral infections may facilitate the destruction of periodontal tissue by lytic activity against periodontal cells and immune suppression, which increase the susceptibility of the host to bacterial attacks. The aim of this study is to detect HCMV DNA in saliva of chronic periodontitis patients and healthy subjects by conventional PCR test and to analyze the relation with periodontal status. The study sample consisted of thirty-two chronic periodontitis patients of both gender (14 males and 18 females) and fifteen healthy subjects of both gender (8 males and 7 females) with age ranged from 25 to 45 years. All the pa-

ticipants in this study without any systemic disease. Periodontal parameters used in this study were plaque index, gingival index, bleeding on probing, probing pocket depth and clinical attachment level. Unstimulated saliva samples were collected from all subjects and examined by conventional PCR test for HCMV DNA detection. The results of this study observed that the percentage of chronic periodontitis group who were positive for HCMV DNA was (31.25%) and of control group was (13.33%). The present study observed that there was no significant difference between chronic periodontitis and control groups ($P \geq 0.05$) regarding the presence of HCMV DNA. There was no significant difference between male and female ($P \geq 0.05$) in the presence of HCMV DNA.