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Employing Biogenic Reduced Graphene Oxide as an Antibacterial Agent Against Some MDR Pathogenic Bacterial Isolates

Abtisam Jasim Abbas¹

Hazim Abed Walli²

Ali A. Ghazzay³

Environment Department, Science College, University of AL-Qadisiyah, Iraq^{1,2}

Biology Department, Science College, University of AL-Qadisiyah, Iraq³

Corresponding author: Abtisam Jasim Abbas

abtisam.jasim@qu.edu.iq

Abstract:

Antibiotic resistance in pathogenic microbes has become a severe health issue worldwide. Consequently, hard work has been made to progress novel materials with antimicrobial effectiveness. Current research was accomplished to investigate the efficiency of biogenic reduced graphene oxide nanoparticles, which were reduced for the first time by two bacterial strains, *Heyndirckxia coagulans* (accession no. PP425868.1) and *Streptococcus thermophilus* (accession no. PP425862.1). The biosynthesized rGO NPs appeared to have antibacterial activity against (*Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Klebsiella pneumonia*, and *Pantoea agglomerans*), which were isolated from burns, wounds, and urinary tract infections. Different concentrations of rGO NPs (0.05, 0.1, and 0.2 mg/ml) have been applied against the above MDR bacterial isolates. Results revealed that the development of inhibition areas was augmented with the increase of rGO NPs concentration compared to the ampicillin antibiotic. Using rGO NPs as antibacterial agents for curing infections associated with MDR bacteria is developing owing to exceptional physicochemical features such as high surface area, shape, and biocompatibility of rGO NPs.

Key Words: Biogenic reduced graphene oxide, *Heyndirckxia coagulans*, *Streptococcus thermophilus*, Antibacterial Activity, MDR bacteria,

Introduction

Recently, the random use of antibiotics has become a developing issue (1) and lead to the presence of multi-drug-resistance bacterial isolates (2). When antibiotics discharge into the environment they destroy the bacteria realm, aquatic and soil organisms, as well as plants. As a result, bacteria particularly progress resistance or even MDR. These bacteria can produce deactivating enzymes and block drugs by changing the permeability of the cell membrane (3). The noticeable development of antimicrobial resistance and the lack of new antimicrobial medicine improvement has gradually decreased the treatment elections for bacterial infection (4).

The current concept proposes that these bacteria develop resistance to numerous drugs by obtaining antimicrobial resistance genetic factors via horizontal transfer, facilitated by moveable genetic components like integrons, plasmids, and transposons among and across diverse bacterial strains is a significant factor that can be involved in the rise of multi-drug resistant bacteria (5). Therefore, it is crucial to discover unconventional resources as an alternative of antibiotics to perform an antibacterial function (6). One of the potential techniques used to overcome the great prevalence of MDR bacterial strains is Nanotechnology (7). The Nano-scale materials that have showed antibacterial activity like Ag, AgO, Au, TiO₂, Cu, Zn, ZnO and CuO (8,9) have shown noteworthy beneficial antimicrobial action.

Most recently, graphene-materials (GMs) have been widely studied for numerous applications for instance, environmental and medical fields (10,11). Owing to the physicochemical features of GMs, researchers have studied their potential application in several biotechnologies (12-14). Many prior studies have informed that rGO NPs has antimicrobial efficacy (15-17). Altogether, the antibacterial efficacy of the graphene-based nanomaterials could be attributed to the synergy of oxidation stress and physical damages to the membrane (18). The sharp edges of graphene oxide (GO) and rGO resulted in physical damages to the bacteria's cell wall and membrane. Moreover, graphene sheets could promote cellular oxidative stress, which contributed to the disruption of particular microbial processes (19,20).

The high NPs (SSA) allow particles to bind somewhat a great concentration of functional groups or serves as transporters for other active matters, improving their contact

with bacteria (21). In general, small size nanoparticles show better antibacterial activity, because decreasing of the volume will cause increasing of the surface area, and hence increase the antibacterial activity. Furthermore, the cell membrane has nanosized pores which allow the smaller nanoparticles easily to penetrate the cell and reach to the nuclear content of bacteria (reverse osmosis) (22). Eventually, the antibacterial properties of NPs are difficult and include diverse routes. Consequently, the mechanism of antibacterial action is yet vague (23).

Experiment parts

1- Bacterial Isolates

Five bacterial isolates were used in the current research for the detection of rGO NPs biological activity, i.e., (*Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Klebsiella pneumonia*, and *Pantoea agglomerans*), which were isolated from different sources like burns, wounds, and urinary infections, diagnosed by VITEK-2 system.

2- Antibiotics sensitivity test of pathogenic isolates

The susceptibility of antibiotics test (AST) was conduct utilizing VITEK2 system.

3-Antibacterial Activity of rGO NPs

The antibacterial efficacy of rGO NPs were evaluated by using an agar-well diffusion technique against five bacterial pathogens i.e. (*Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Klebsiella pneumonia*, and *Pantoea agglomerans*).

A free culture of pure five isolate colonies was suspended in brain heart infusion broth (5ml), to get an optical density equal to the 0.5 McFarland criteria (equivalent to 1.5×10^8 cells/ml). The mixture was incubated overnight at 37 °C. Moreover, the turbid growth culture was calibrated using sterile broth (24). A sterilized cotton swab was dipped into the solution and used to streak it across the whole surface of a Mueller-Hinton agar plate. Stock concentrations of rGO nanoparticles and antibiotic standard drug ampicillin were prepared at three different concentrations (0.05, 0.1, and 0.2 mg/ml). After that, 4 wells were produced using a sterile cork porer in each plate and 100 μ L of both rGO nanoparticles and ampicillin were poured into wells that had been prepared on agar plates. Distilled water served as the negative control. The pore diameter measure was 7 millimeters.

Finally, the Petri dishes were incubated at 37 °C overnight in an incubator to assess the antibacterial effect of rGO nanoparticles in comparison with antibiotic drug. The diameter of the growth inhibition zones was measured in millimeters. All the test plates were prepared in triplicate and incubated for 24 h at 37°C. After incubation, transparent ruler was used to measure the zone of inhibition (ZOI) diameter around the well (24,25). Ampicillin represents the positive control, while distilled water acts as the negative control (26).

Results and Discussion

Antibacterial Activity of rGO NPs

The antibacterial activity of different concentrations (0.05, 0.1 and 0.2 mg/ml) of Heyndirckxia coagulans-reduced graphene oxide (HC-rGO) and Streptococcus thermophilus-reduced graphene oxide (ST-rGO) nanoparticles against clinical pathogenic isolates Gram-positive isolate (*Staphylococcus aureus*) and Gram-negative isolates (*Pseudomonas aeruginosa*, *Escherichia coli*, *Klebsiella pneumoniae*, and *Pantoea agglomerans*) were studied by agar well diffusion method. The inoculated plates were incubated at 37°C for overnight for detection of antibacterial activity.

The results showed that both nanoparticles had potent inhibitory effects against all the tested clinical pathogenic isolates, paradoxically ampicillin has slight inhibitory effects against all the tested clinical pathogenic isolates. The growth inhibition zones clearly revealed that both nanoparticles inhibited the growth of tested clinical pathogenic isolates where the highest concentration of both nanoparticles have the strongest antibacterial activity and their inhibited zone ranged from 10.0 ± 1.73 to 27.0 ± 1.73 mm, whereas ampicillin revealed inhibition zone ranged from 0 to 13.0 ± 1.0 mm.

The obtained results revealed a significantly high zone of inhibition against Gram-positive isolate and Gram negative isolates. HC-rGO revealed the highest zone of inhibition (27.0 ± 1.73 mm) at concentration (0.2 mg/ml) against *Escherichia coli* and *Pantoea agglomerans* in comparison to positive control (ampicillin) with a zone of inhibition of (11.33 ± 0.58 and 12.33 ± 0.58), respectively ($p \leq 0.05$), while ST-rGO revealed the largest zone of inhibition (25.0 ± 1.0 mm) at the same concentration against *Escherichia coli* in comparison to ampicillin with a zone of inhibition of (11.33 ± 0.58) (p

≤ 0.05). On the other hand, the negative control (distilled water) has showed no inhibition zone with 0.0 mm. As shown in Figure 1.

The obtained results are in agreement with those found by several investigators (27-29) who stated that the antibacterial activity of Bio-reduced graphene oxide (B-rGO) indeed demonstrated potent growth inhibition zones was increased with increasing concentrations of B-rGO from *Escherichia coli*, *Bacillus clausii* and *Lysinibacillus sphaericum*, respectively.

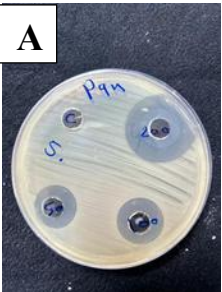




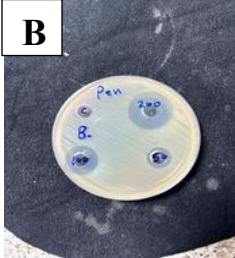



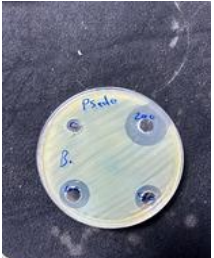
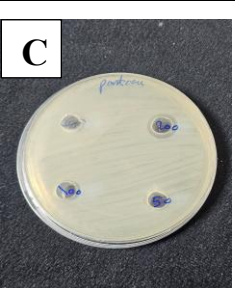




A 				
<i>Pantoea agglomerans</i>	<i>Klebsiella pneumoniae</i>	<i>Staphylococcus aureus</i>	<i>Escherichia coli</i>	<i>Pseudomonas aeruginosa</i>
B 				
<i>Pantoea agglomerans</i>	<i>Klebsiella pneumoniae</i>	<i>Staphylococcus aureus</i>	<i>Escherichia coli</i>	<i>Pseudomonas aeruginosa</i>
C 				
<i>Pantoea agglomerans</i>	<i>Klebsiella pneumoniae</i>	<i>Staphylococcus aureus</i>	<i>Escherichia coli</i>	<i>Pseudomonas aeruginosa</i>

Figure 1: Illustrates the antibacterial activity of (A) ST-rGO, (B) HC-rGO Nanoparticles and (C) ampicillin against MDR bacterial isolates

Initially, B-rGO can provide a unique isolation barrier between two different regions and encapsulate microorganisms (30). In addition, due to the lamellar structure of reduced graphene oxide (rGO), its sharp edges can cut and rupture cell membranes during contact with cells, thereby destroying the integrity of cell structure and being toxic to the cells (31,32), furthermore, B-rGO leads to oxidative stress by directly forming Reactive Oxygen Species (ROS) or consuming cellular antioxidants (33,34). As a result, B-rGO shows steady biocompatibility compared to that commercial reduced graphene oxide (C-rGO) (35).

Table 1: Evaluation of antibacterial activity of HC-rGO nanoparticles against MDR pathogenic isolates as compared to ampicillin.

Microorganisms	Zone of inhibitions (mm), mean \pm SD, n = 3					
	Concentration of Ampicillin (μ g/ml)			Concentration of HC-rGO (μ g/ml)		
	50	100	200	50	100	200
<i>Pseudomonas aeruginosa</i>	-	9.33 \pm 0.58	10.33 \pm 0.58	15.67 \pm 1.15	18.33 \pm 1.5	25.33 \pm 3.06
<i>Staphylococcus aureus</i>	-	9.67 \pm 0.58	11.0 \pm 1.0	15.67 \pm 0.58	20.0 \pm 2.0	26.67 \pm 3.06
<i>Pantoea agglomerans</i>	-	10.67 \pm 0.58	12.33 \pm 0.58	16.67 \pm 1.53	22.0 \pm 0.0	27.0 \pm 1.73
<i>Escherichia coli</i>	-	9.33 \pm 0.58	11.33 \pm 0.58	16.33 \pm 0.58	19.0 \pm 1.0	27.0 \pm 1.0
<i>Klebsiella pneumoniae</i>	-	10.33 \pm 0.58	13.0 \pm 1.0	16.0 \pm 1.73	19.0 \pm 1.0	25.0 \pm 1.0

(-) indicates that the absence of inhibited zone as no ampicillin and antibacterial activities.

Values are means of 3 replicates \pm standard deviations.

Table 2: Evaluation of antibacterial activity of ST-rGO nanoparticles against MDR pathogenic isolates as compared to ampicillin.

Microorganisms	Zone of inhibitions (mm), mean \pm SD, n = 3					
	Concentration of Ampicillin (μ g/ml)			Concentration of ST-rGO (μ g/ml)		
	50	100	200	50	100	200
<i>Pseudomonas aeruginosa</i>	-	9.33 \pm 0.58	10.33 \pm 0.58	17.0 \pm 1.73	20.33 \pm 2.89	24.0 \pm 0.0
<i>Staphylococcus aureus</i>	-	9.67 \pm 0.58	11.0 \pm 1.0	16.33 \pm 5.13	20.33 \pm 4.93	24.67 \pm 5.03
<i>Pantoea agglomerans</i>	-	10.67 \pm 0.58	12.33 \pm 0.58	14.67 \pm 3.06	18.67 \pm 1.15	24.67 \pm 4.62
<i>Escherichia coli</i>	-	9.33 \pm 0.58	11.33 \pm 0.58	10.0 \pm 1.73	18.0 \pm 1.0	25.0 \pm 1.0

<i>Klebsiella pneumoniae</i>	-	10.33±0.58	13.0± 1.0	12.0±2.0	19.0±1.0	23.0±1.73
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(-) indicates that the absence of inhibited zone as no ampicillin and antibacterial activities.

Values are means of 3 replicates ± standard deviations.

Finally, from the current results above, it can be concluded that biogenic rGO NPs demonstrated significant antibacterial activity against the tested MDR bacterial isolates, and the efficiency increased with the increase in biogenic rGO NPs concentration.

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