



**Article**

**Preparation and Evaluation of the polyelectrolyte complex of Chitosan/Carrageenan Cross-Linked with Glutaraldehyde**

**Zahraa T. Hano, Tahseen A. Saki<sup>a</sup>, Bushra A.A. Al Salem<sup>b</sup>**

Department of Chemistry, College of Science, University of Basrah

<sup>a</sup> Corresponding author: email: [Tahseen.saki@uobasrah.edu.iq](mailto:Tahseen.saki@uobasrah.edu.iq)  
<https://orcid.org/0000-0003-3155-5810>

<sup>b</sup> email: [bushra.abdulaziz@uobasrah.edu.iq](mailto:bushra.abdulaziz@uobasrah.edu.iq)  
<https://orcid.org/0000-0003-2370-0416>

**ABSTRACT**

This research work deals with the preparation hydrogel based on Chitosan (CS) and Carrageenan (CG) have been studied. Polyelectrolyte complexes (PECs) hydrogels were prepared using Chitosan and Carrageenan with glutaraldehyde as the crosslinking agent. The ratio of glutaraldehyde as crosslinking agents was investigated. Upon cross-linking with glutaraldehyde was varied as 0, 20,50 and 80% (w/w) of the total quantity of chitosan and carrageenan. Prepared hydrogels were characterized for swelling ability in (pH = 1.2 acidic, 8.2 basic, 7.4 neutral), FTIR, mechanical and thermal properties. Results showed that swelling or water absorbency decreased with increased concentration of glutaraldehyde. Prepared hydrogels maintained high swelling in acidic media as compared to neutral and basic. FTIR spectroscopy indicated formation of a Schiff base C=N bond due to reaction between NH<sub>2</sub> of chitosan and the aldehyde group of glutaraldehyde. The mechanical properties assay findings prove that physically crosslinked chitosan/carrageenan are more biocompatible and enhance more the adhesion with addition chemical crosslinked by glutaraldehyde. The addition of glutaraldehyde improves strengthens of the chitosan/carrageenan and its thermal properties; the

resulting was found to be more efficient for thermal properties than their non-crosslinking blend.

**Keywords:** Chitosan, Carrageenan, Hydrogel films, Mechanical Properties, Thermal properties

## **1. Introduction**

Hydrogels are cross-linked polymeric materials that can absorb large volumes of water. The Hydrogels ability to absorb water is due to hydrophilic groups in polymers chemical structure<sup>(1)</sup>. Hydrogels exhibit a distinctive swelling tendency without dissolving in an aqueous environment because of their high-water absorption capacity and the presence of crosslinks in their structure<sup>(2)</sup>. In hydrogel formulations, various cross-linking techniques have been employed. Hydrogel compositions often employ physical and chemical techniques. Hydrogen bonds, covalent bonds, van der Waals interactions or physical entanglements are the sources of cross-links in polymeric networks<sup>(3)</sup>. Polyelectrolyte complexes (PECs) are macromolecules with a lot of charged or easily charged groups occurred when oppositely charged polyelectrolyte solutions are mixed without the need of solvents<sup>(4)</sup>. PECs are being studied more and more in academia and industry because of their easy manufacture and excellent delivery system for macromolecules and pharmaceuticals that are prone to deterioration<sup>(5,6)</sup>. The hydrophilic-hydrophobic balance, molar mass, charge density, ionic strength and the polyion mixing mode of the complimentary polymers, can all be used to adjust PEC properties. Secondary bonds such as van der Waals interactions and hydrogen bonds play a major role in the formation and integrity of PECs. However, pH values are important values in the formation PEC between polyelectrolytes<sup>(7)</sup>. Chitosan (CS) is a naturally polysaccharide with varying molecular weights and degrees of deacetylation that are deacetylated derivatives of the natural polysaccharide chitin. It is made up of deacetylated units called  $\beta$ -1,4-linked glucosamine and acetylated units called N-acetyl-d-glucosamine, with molecular weights ranging from 10 to 1,000 kDa and typical deacetylation levels between 70-95% and presence of reactive functional groups, such as  $\text{NH}_2$  and  $\text{OH}$ <sup>(8)</sup>. Chitosan considers one of naturally occurring cationic polysaccharide functions well as a bioadhesive substance. Furthermore, because of its hydrophilic nature, high availability, great biocompatibility, and biodegradability, chitosan-based PECs have been widely used in drug delivery<sup>(9)</sup>. PECs based on chitosan and related

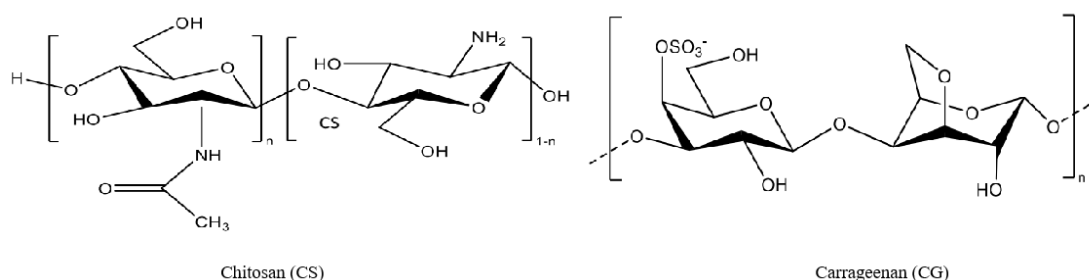
polymers have been utilized in medication delivery for the past few decades. Controlled release of proteins and vaccines. However, the cationic amino group at the C2 position of the repeating glucopyranose unit of chitosan can electrostatically interact with the anionic groups of other polyions to form PECs many different polyanions of natural origin such as hypromellose phthalate<sup>(10)</sup> and carboxymethyl cellulose sodium<sup>(11)</sup> Sodium alginate<sup>(12)</sup> alginate, carrageenan<sup>(13)</sup>. Carrageenan (CG) is an anionic polymer naturally occurring, high molecular weight biopolymer that is ecofriendly and non-irritating, allowing for its usage in biomedical applications. CG has alternating  $\alpha(1-3)$ -D-galactose-4-sulfated and  $\beta(1-4)$ -3,6-anhydro-D-galactose units, containing one sulfate group per disaccharide unit<sup>(14)</sup>. However, the scientific papers and application of using glutaraldehyde to synthesis a crosslinked chitosan/carrageenan is little<sup>(15-17)</sup>. Therefore, this article using glutaraldehyde as a crosslinking agent to create a novel resin material based on chitosan/carrageenan chitosan hydrogel.

Chitosan/Carrageenan (CS/CG) hydrogel blend were prepared by PECs of oppositely charged CS (cation) and CG (anion). Also, prepared CS/CG hydrogel blend with glutaraldehyde as the cross-linking agent.

## 2. Materials and Method

### 2.1. Chemicals

Chitosan (CS) 75% (CAS Number 9012-76-4) from HIMEDIA, Carrageenan (CG) (CAS Number 9000-07-1) from SIGMA-ALDRICH. Glycerol from Fluka, glutaraldehyde and acetic acid from PDH.



### 2.2. Preparation Polyelectrolyte Complexes

Bio-polymeric films were prepared by dissolving each of polymers separately, where 1g of CS was dissolved in 1% acetic acid with continuous stirring (500 rpm) for 15 minutes, then leave it for 15 minutes until the bubbles disappear.

CG was dissolved by distilled water with continuous stirring. After that, added drop by drop to CS by stirring to obtain a homogeneous solution. Before the last addition, 0.1 ml of glycerol was added to give elasticity to the prepared films.

Three ratios were used 20, 50, and 80% of the crosslinking agent, glutaraldehyde was added to the total solution. The homogenized solution was transferred to a Petri dish and allowed to dry at laboratory temperature. After drying, the films were washed with distilled water to get rid of the acid.

### **2.3. Swelling index**

Swelling indicators were recorded for the prepared polymeric films (PECs) by immersing samples of similar weight in different acidic solutions (pH) (1.2, 7, 8.2) at room temperature. Then removing them from the solutions at regular time intervals and recording the weight of the sample after drying it. For each entanglement rate, three samples were taken and immersed in the media mentioned above. The swell ability was determined by the following equation:

$$Q\% = (W_s - W_d) / W_d * 100$$

Q represents the swelling ratio,  $W_s$  represents the weight of the swollen film,  $W_d$  represent the weight of the dry film.

## **3. Results and Discussion**

### **3.1. Fourier transform infrared spectroscopy (FTIR)**

FTIR spectra obtained for the Chitosan/Carrageenan hydrogels containing 80% glutaraldehyde (cross-linking agent) are collected together for better comparison are shown in Figure 3. The peaks at 1648 to amide I ( $\text{CONH}_2$ ) and 1545  $\text{cm}^{-1}$  corresponded to amide II ( $\text{NH}_2$ ) was dedicated in the chitosan spectrum (Figure 1). In the spectrum ((Figure 2) of the carrageenan, the peak at 1221  $\text{cm}^{-1}$  was belonging to the band of  $\text{S}=\text{O}$  ( $\text{SO}_4$ ). In comparison, the FTIR spectrum (Figure 3) of the Chitosan/Carrageenan cross-linked with glutaraldehyde (80%) showed a decrease in peak intensity of  $-\text{NH}_3^+$  of chitosan at 1450  $\text{cm}^{-1}$  and appeared a new peak at 1699  $\text{cm}^{-1}$  indicated the formation of linkage ( $\text{C}=\text{N}$  imine) from reaction  $\text{NH}_2$  groups of chitosan and the  $\text{C}=\text{O}$  groups of glutaraldehyde.

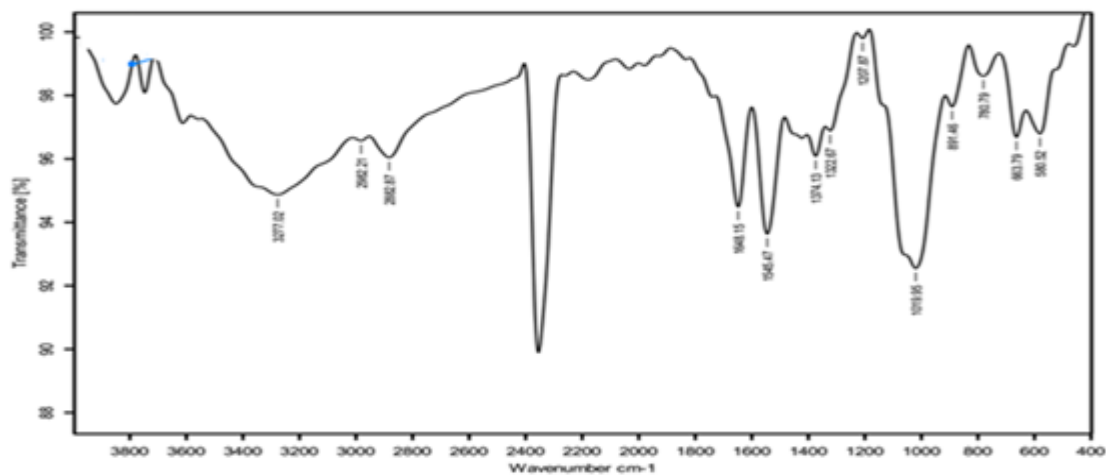


Figure 1. FTIR spectroscopy of the Chitosan

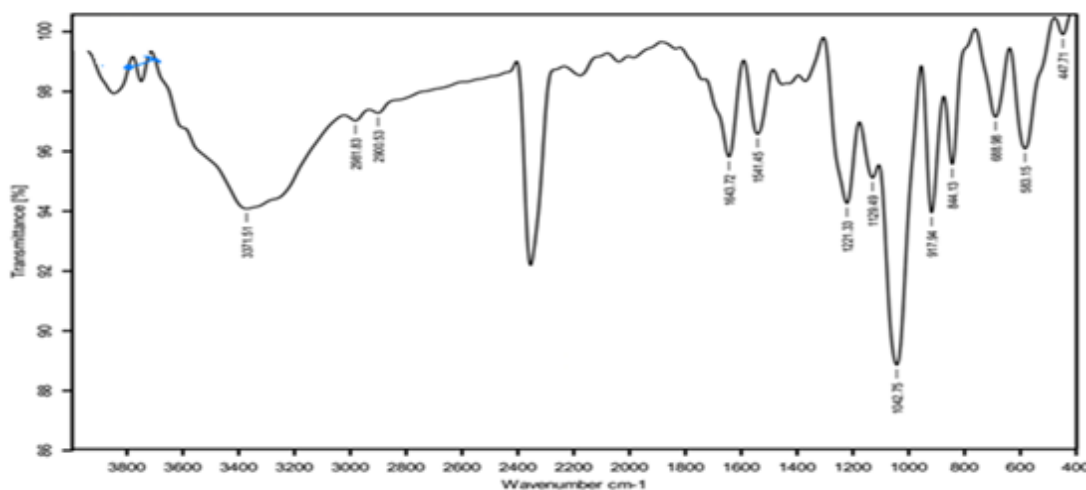


Figure 2. FTIR spectroscopy of the Carrageenan

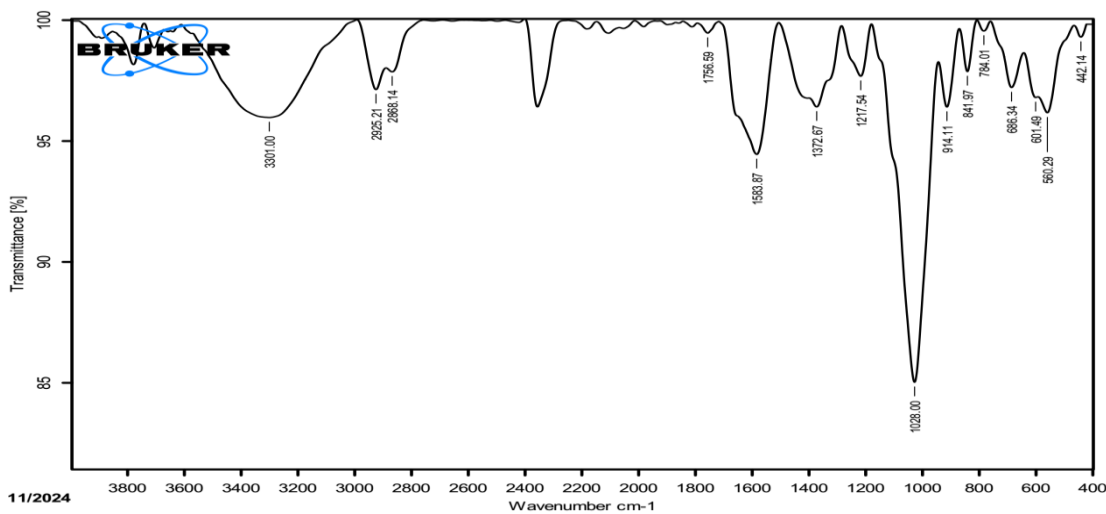
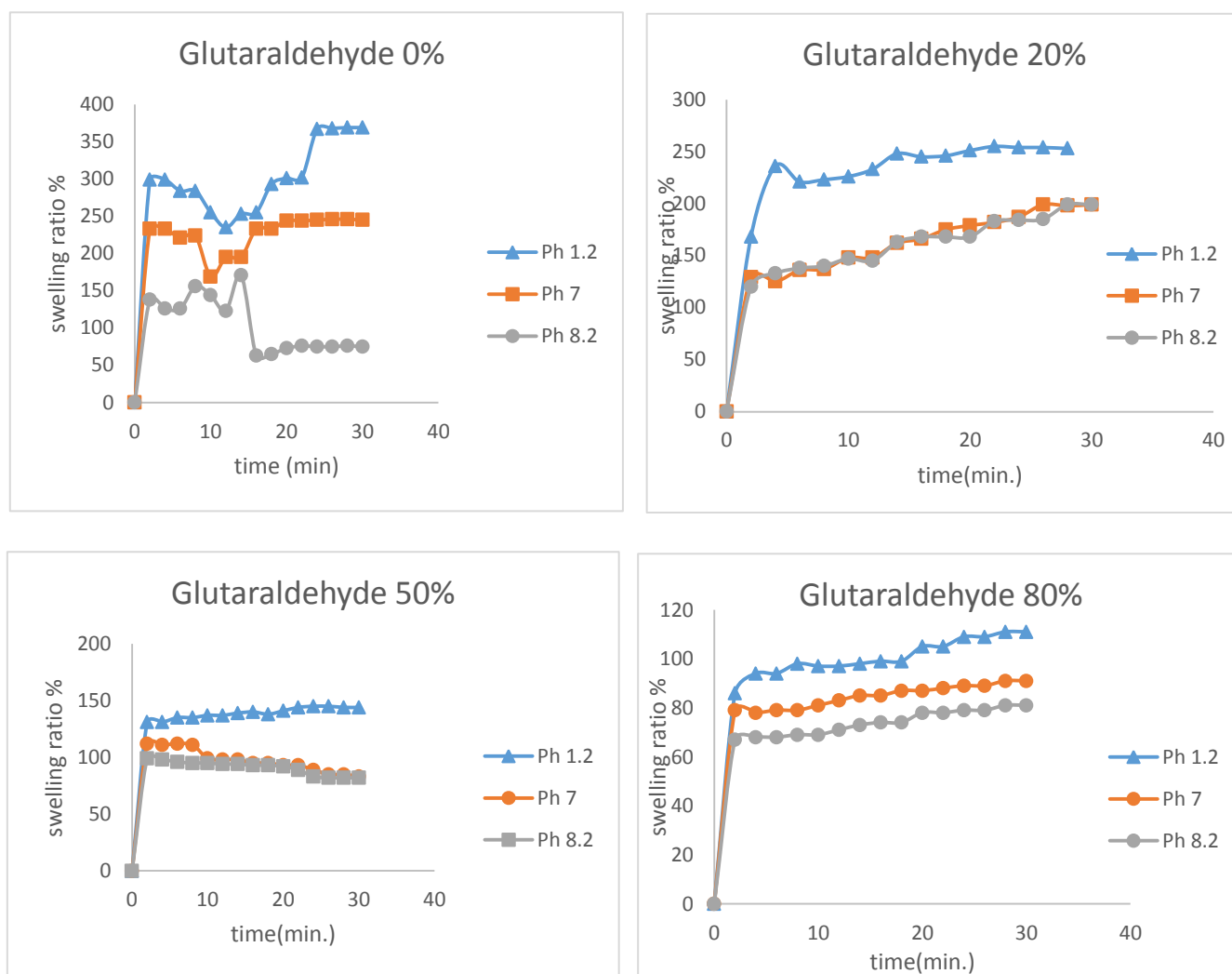


Figure 3. FTIR spectroscopy of the CS/CG/glutaraldehyde

### 3.2. Swelling Study

The swelling ratio study of the CS/CG was performed in 3 dissolution systems by the pH media 1.2; 7.4 and 8.2. The hydrogel blend prepared without any addition of crosslinking agent had highest swelling ratio. However, swelling ratio of the blend decreased with increased addition of crosslinking agent (glutaraldehyde). On the other hand, the highest value of swelling was detected in acidic pH media. Polymeric chains of chitosan and carrageenan had complete freedom of movement, when was not added with crosslinking agent. In other words, increasing the crosslinker concentration forms a strong structure that reduce the swelling due to decrease in the movement of the polymer chains besides decreased the number of hydrophilic groups present in them, lower their ability to absorb water, see Figure 4.



**Figure 4.** Swelling ratio obtained for the prepared Chitosan/Carrageenan blend hydrogels

### 3.3. Mechanical Properties

As shown in Table 1 the CS/CG gel films showed good mechanical properties, with  $\delta_B$ ,  $\epsilon_B$ , and E being results superior to the values of CS/CG gel without crosslinking agent (glutaraldehyde). As expected, weight ratio of glutaraldehyde significantly influenced the mechanical performance of the gels. The gel film with a weight ratio 80% had the maximum  $\delta_B$  and  $\delta_M$ , whereas the CS/CG gel film with a weight ratio 50% possessed the lowest values. However, it was interesting that the CS/CG with slight excess of glutaraldehyde had the highest values of mechanical properties data. Crosslinking bridge bonds formed when added glutaraldehyde, resulting in a rigid and brittle physical CS/CG gel. It was remarkable that the CS/CG gel possessed good mechanical performances over a wide range of weight ratio. This characteristic occurred due to multiple interactions and the crosslinking network of the gels<sup>(7)</sup>.

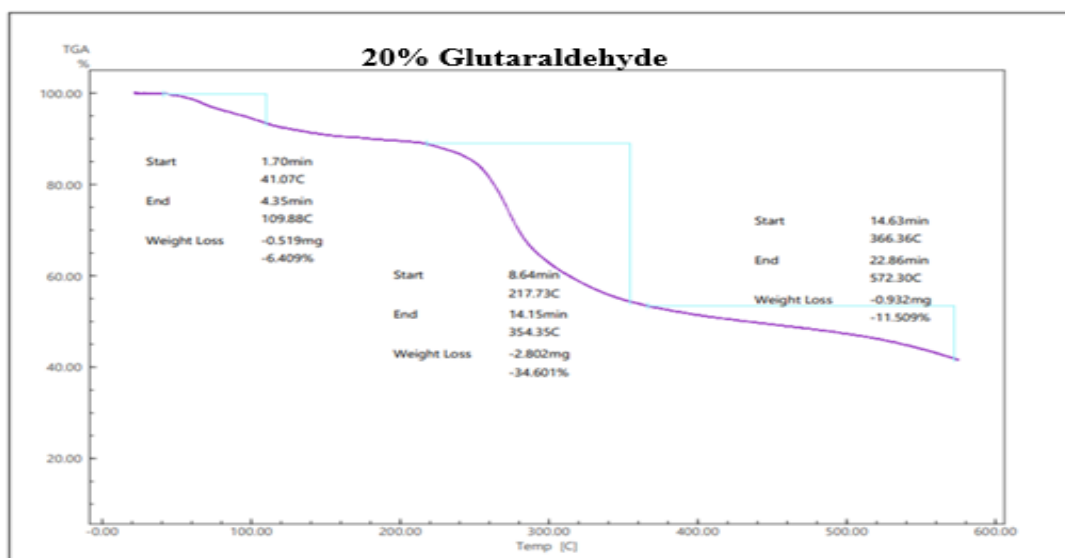
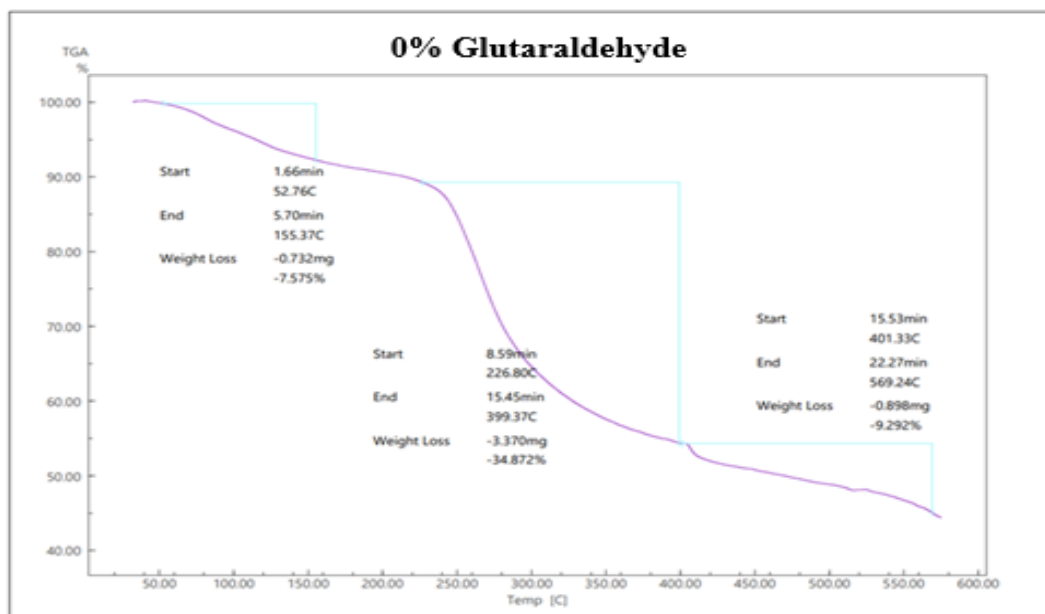
**Table.1.** Mechanical properties data of the Chitosan/Carrageenan blend with different ratio of glutaraldehyde

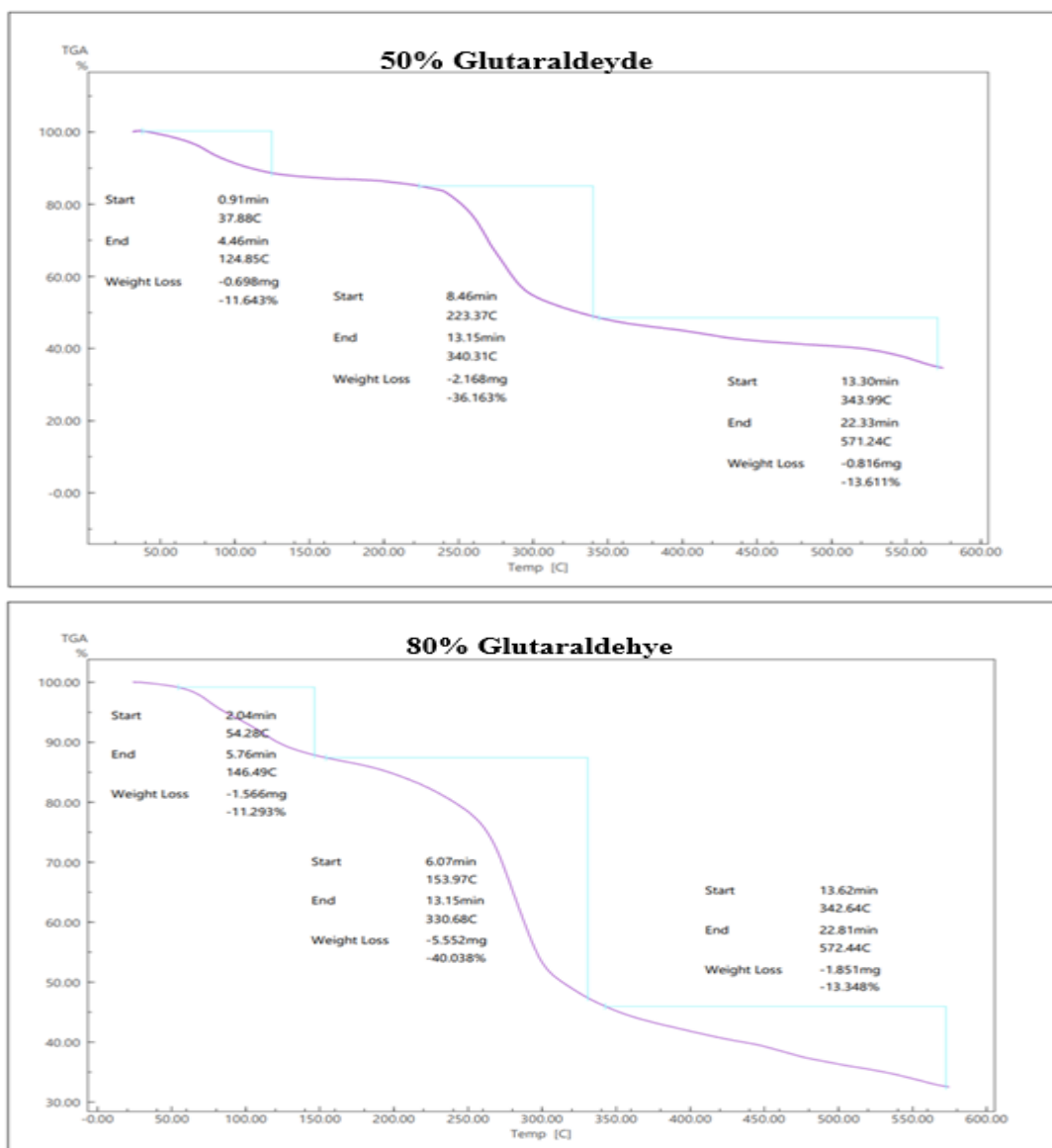
Glutaraldehyde Ratio %	$\delta_M$ (Mpa)	$\epsilon_M$ %	$\delta_B$ (Mpa)	$\epsilon_B$ %	Young Modulus E (Mpa)
0	5.48	1.0	5.46	1.0	5.48
20	7.32	0.9	7.32	0.9	8.13
50	2.73	0.6	2.6	0.7	4.55
80	10.1	1.1	9.6	1.1	9.18

### 3.4. Thermogravimetric analysis TGA

Thermo gravimetric analysis TGA was measure the thermal stability of the CS/CG blends with different percentages of glutaraldehyde with heating rate 20 °C.min<sup>-1</sup> in inert atmosphere (Figure 5). The weight losses at different temperatures were analyzed. It is obvious, that the weight loss (%) of CS/CG blends crosslinked glutaraldehyde was greater compared to that of blends without crosslinking agent. Ultimate degradation temperature of blends without glutaraldehyde is 265 °C. However, added glutaraldehyde to the blends in several % ratio, which enhance the thermal stability compared with non-crosslinking blend. However, degradation

temperature of blend cross-linked with 20, 70 and 80% glutaraldehyde were at 267, 269 and 280 °C respectively. Also, from the char content results we are able to conclude that the prepared CS/CG blends showed relatively high thermal stability as shown in Table 2.





**Figure 5.** Thermogravimetric analysis curves of CS/CG blends with different ratio of Glutaraldehyde

**Table.2.** Thermal properties data of the CS/CG blend with different ratio of Glutaraldehyde

Glutaraldehyde Ratio %	Ultimate Decomposition Temperature °C	Weight Loss%	Temperature of Weight Loss °C	Rate of Decomposition (min <sup>-1</sup> . %)	Char %Content At 550 °C
0	265	25	275	1.5	48.26
		50	485		
		75	-		
20	267	25	292	1.54	51.3
		50	488		
		75	-		
50	269	25	270	1.36	38.5
		50	330		
		75	-		
80	280	25	268	1.41	35.32
		50	310		
		75	-		

#### 4. Conclusion

A series of hydrogel blends were prepared using chitosan and carrageenan with glutaraldehyde as the crosslinking agent. Prepared hydrogels were characterized for water absorbency or swelling ability in acidic and basic pH and neutral media, FTIR spectroscopy, mechanical and thermal properties. It was found that swelling capacity decreased with increased concentration of glutaraldehyde in the blends. However, prepared hydrogel had higher water absorbency in acidic media as compared to basic and neutral PH. Prepared hydrogels exhibited good mechanical properties and thermal stability.

#### References

- [1]Pugar D, Haramina T, Leskovic M and Curković L [2024]: Preparation and Characterization of Poly(vinyl-alcohol)/Chitosan Polymer Blend Films Chemically Crosslinked with Glutaraldehyde: Mechanical and Thermal Investigations. *Molecules*. 29, pp. 5914 (1-17).
- [2]Abed MA, Haddad AM, Hassen AJ and Sultan SM [2006]: Preparation and evaluation of new hydrogels as new fertilizer delivery system. *Basrah Journal of Scienc (C)*, 24(1), pp. 103-114.

- [3] Fodor CS, Domjan A and Ivan B. [2013]: Unprecedented scissor effect of macromolecular cross-linkers on the glass transition temperature of poly(N-vinylimidazole), crystallinity suppression of poly(tetrahydrofuran) and molecular mobility by solid state NMR in poly(N-vinylimidazole)-*l*-poly(tetrahydrofuran) conetworks. *Polym. Chem.* 4, pp. 3714–3724.
- [4] Wu QX, Wang DD, Su T, Cheng X, Du D, Xu X and Chen Y [2017]: Self-assembly of polyelectrolyte complexes microcapsules with natural polysaccharides for sustained drug release. *Cellulose.* 24, pp. 4949–4962.
- [5] Bourganis V, Karamanidou T, Kammona O and Kiparissides C [2017]: Polyelectrolyte complexes as prospective carriers for the oral delivery of protein therapeutics. *Eur. J. Pharm. Biopharm.*, 111, pp. 44–60.
- [6] Wu QX, Wang DD, Su T, Cheng XD, Xu X and Chen Y [2017]: Self-assembly of polyelectrolyte complexes microcapsules with natural polysaccharides for sustained drug release. *Cellulose*, 24, pp. 4949–4962.
- [7] Hai CY, He Z, Kefeng R, Zhimin Y, Fengbo Z, Jin Q, Jian J, Zi LW and Qiang Z. [2018]: Ultrathin  $\kappa$ -Carrageenan/Chitosan Hydrogel Films with High Toughness and Antiadhesion Property, *ACS Appl. Mater. Interfaces*, 10, pp.9002-9009.
- [8] Pellá M.; Lima-Tenório MK, Tenório-Neto E, Guilherme MR, Muniz EC and Rubira AF. [2018] Chitosan-based hydrogels: From preparation to biomedical applications. *Carbohydr. Polym.*, 196, pp. 233–245.
- [9] Al-Assady NA, Hussein WAK and Jabir HA. [2011]: Preparation, Characterization, and Diltiazem HCl Release Study of Chitosan/ poly(vinyl alcohol) Microspheres. *National Journal of Chemistry*, 41, pp.113-126.
- [10] deSouza Chaves MP, de Mattos NH, Pedreiro LN, Boni FI, dos Santos Ramos MA, Bauab TM, Daflon GMP and Chorilli M. [2020]: Design of Mucoadhesive Nanostructured Polyelectrolyte Complexes Based on Chitosan and Hypromellose Phthalate for Metronidazole Delivery Intended to the Treatment of Helicobacter pylori Infections. *Pharmaceutics*, 12, pp. 1211 (1-22).
- [11] Roy JC, Ferri A, Giraud S, Jinping G and Salaün F. [2018]: Chitosan – carboxymethylcellulose-based polyelectrolyte complexation and microcapsule shell formulation. *Int. J. Mol. Sci.*, 19, pp. 2521 (1-19).
- [12] Tomida H, Nakamura C and Kiryu S. [1994]: A novel method for the preparation of controlled-release theophylline capsules coated with a polyelectrolyte complex of  $\kappa$ -carrageenan and chitosan. *Chem Pharm Bull (Tokyo)*, 42, pp. 979-981.
- [13] Olea AF, Carrasco H, Santana F, Navarro L, Guajardo-Maturana R,

- Linares-Flores C and Alvarado N [2024]: Formation and Characterization of Chitosan-Based Polyelectrolyte Complex Containing Antifungal Phenylpropanoids. *Polymers*. 16, pp.3348 (1-16).
- [14] Gupta VK, Hariharan M, Wheatley TA and Price JC. [2001]: Controlled-release tablets from carrageenans: effect of formulation, storage and dissolution factors. *Eur J Pharm Biopharm*. 51, pp.241-248.
- [15] Al-Sokanee ZN, Al-Chezzy KW, Saki TA and Al-Amber LJ. [2007]: Drug Release Study of Nalidixic acid from the Chitosan-Carrageenan Ionic Matrix. *Basrah Journal of Science (C)*, 25(1), pp.104-118.
- [16] Tapia C, Escobar Z, Costa E, Sapag-Hagar J, Valenzuela F, Basualto C, Gai MN and Yazdani-Pedram M.[2004]: Comparative studies on polyelectrolyte complexes and mixture of chitosan-alginate and chitosan-carrageenan as prolonged diltiazem clorhydrate release system. *Eur J Pharm Biopharm*, 57, pp.65-75.
- [17] Genta I, Constantini M, Asti A, Conti B and Montanari L. [1998]: Influence of glutaraldehyde on drug release and mucoadhesive properties of chitosan microspheres. *Carbohydr Polym*. 36, pp.81-88.