



Embryonic Groundwork of Histological Architecture and Implication on Physiology: Developmental Physiological Integration: A Review

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KEYWORDS	ABSTRACT
Embryogenesis; Tissue architecture; Germ layers; Morphogenesis; Organogenesis; Epigenetics.	During embryogenesis, the structural organization of tissues in the human body can be essentially specified, with the use of the most coordinated cell specification, morphogenesis, and spatial patterning. The three germ layers are ectoderm, mesoderm and endoderm that make up the developmental blueprint of all tissues and organ systems. At the microscopic and macroscopic level, the tissue architecture is shaped by early molecular gradients, epigenetic programming and biomechanical forces, which are structural prerequisites for physiological specialization. This review discusses in detail the developmental basis of tissue architecture and critically examines the role of developmental patterning in shaping adult physiological function. Specific attention is given to specification of germ layer, signaling networks, epithelial such as mesenchymal interactions, organogenesis, vascularization, and neurofunctional integration. Moreover, the translational uses of developmental biology in congenital pathology, regenerative medicine, and systems physiology are addressed. Knowledge of the developmental determinants of the tissue structure provides a coherent structure that connects embryology, histology, physiology and clinical medicine.

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1. INTRODUCTION

The embryonic development constitutes one of the most advanced biological processes as the initial fertilized cell develops into a multicellular organism that is structure-organized and functionally differentiated into tissues [1]. This change is controlled by genetic, molecular and biomechanical processes that are strictly gated by factors that, as a whole, dictate the spatial distribution and the identity of cells [2]. Adult tissue architecture is consequently not random; instead, it is the resultant product of genetically coordinated developmental processes initiated during fertilization and progressing during gastrulation, neurulation and organogenesis [3].

One major tenet of integrative biology is that structure defines function. But this principle can obtain its real explanatory force only when studied in a developmental perspective [4]. The architecture of tissues is determined by cell polarity, composition of the extracellular matrix, vascularization, innervation, and three-dimensional organization, and begins in the embryo during embryogenesis [5]. The early physiological decisions are therefore expressed in the physiological functioning in adulthood [6].

The latter has been supported by recent progress in molecular embryology, stem cell biology, and systems physiology to suggest that adult disease vulnerability and capacity are developmentally determined [7]. This functional-developmental continuum emphasizes the fact that the physiological performance and pathological conditions can only be explained in terms of embryological underpinnings [8].

2. EARLY EMBRYOGENESIS: FROM FERTILIZATION AND TOTIPOTENCY TO THE DEVELOPMENTAL BLUEPRINT

The process of embryogenesis starts at fertilization; the zygote, a totipotent cell, is produced, which can produce all embryonic and extraembryonic tissues [9]. The divisions form a cleavage and then a blastocyst, which consists of the inner cell mass (ICM) and the trophoblast [10]. The pluripotent cells of the ICM will produce the embryo proper [11].

The first separation is the first axis of differentiation and is responsible for defining lineage commitment and cellular competence [12]. The geometrical arrangement of the blastocyst is a prelude to subsequent tissue structure, with stress on structural asymmetry preceding functional specialization [13].

3. GASTRULATION AND GERM LAYER FORMATION

The last step in embryonic patterning is called gastrulation, during which cells from the epiblast pass through the primitive streak to give rise to the trilaminar embryonic disc consisting of three germ layers: ectoderm, mesoderm, and endoderm [14]. This is a key step in the organization of the main body plan, and precisely establishes the anterior-posterior axis, dorsal-ventral and left-right axis of the developing embryo [15]. Complex morphogen gradients and networks of transcription factors control this spatial organisation at the molecular level, and coordinate spatiotemporal gene expression and cell fate specification [16]. Overall, these three germ layers provide the initial scaffolding, that is, a direct connection between early embryo patterning and the specialized architecture and functional complexity of adult tissues, as shown in Figure 1[17].

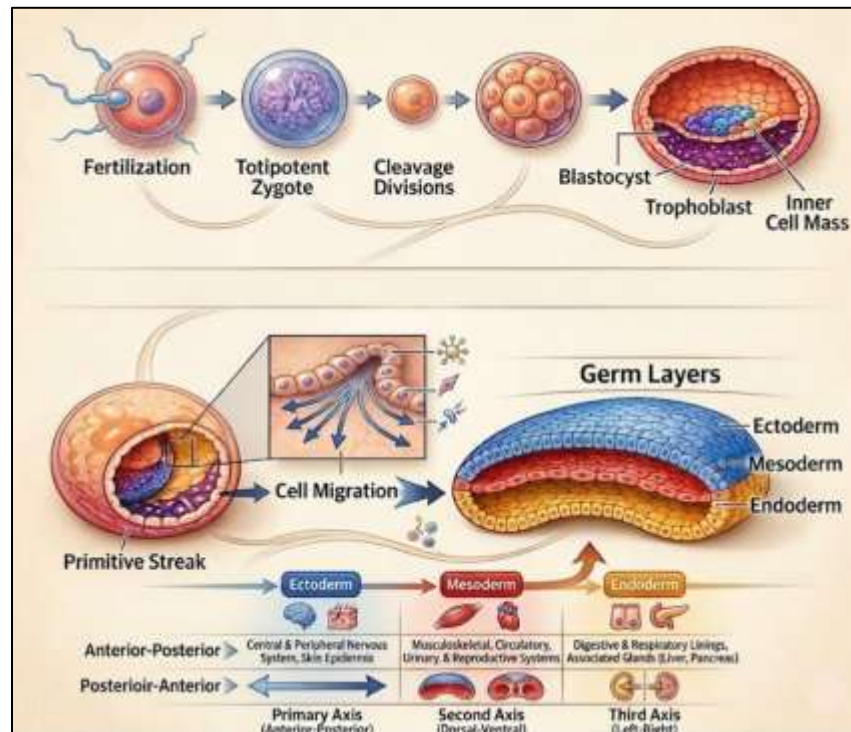


Fig. 1. Timeline of Early Embryogenesis and Gastrulation.

The upper panel shows the early stages of development from fertilization to cleavage divisions and forming the inner cell mass (ICM) of the blastocyst. The lower panel shows the process of gastrulation with cells migrating through the primitive streak to form the trilaminar embryonic disc (ectoderm, mesoderm, endoderm). The diagram illustrates the particular derivatives of the adult tissues and the formation of the major axes of the body (front-rear, top-bottom, and left-right). Created using BioRender.com, integrating its built-in Artificial Intelligence (AI) generation tools.

4. ECTODERMAL DERIVATIVES AND NEUROEPITHELIAL SPECIALIZATION: NEURAL INDUCTION AND CENTRAL NERVOUS SYSTEM FORMATION

A key step in this process is the commitment of the neuroectoderm, and further, this will be followed by the formation of the neural plate, which will then fold and close to form the neural tube [18]. The additional regionalization of this tube leads to an increase in the morphogenesis of the brain vesicles and lengthening of the spinal cord [19]. The basic structure of the C.N.S is based on the intrinsic organization of the neuroepithelial cells, which are characterized by an apical-basal polarity, characteristic of the organization of the ventricular zones and the layered cortices [20]. The radial glial scaffold further defines this spatial plan as it provides the scaffolding necessary for the direction of migration of newly born neurons and is crucial to the laminar architecture that underlies precise synaptic integration [21]. Lastly, this structural organization holds physiological implications: functional differentiation of the various regions of the cortex critically relies on the arrangement of connections among the various layers; developmental specification of the voltage-gated ion channels configuration is important for the timing when the membrane becomes excitable, and the brain starts sending information to each other through synapses. Myelination is also a functional maturation which greatly increases the conduction velocity of axons and allows for the synchronized coordination of the activities of complex neural network systems [23].

Simultaneously, neurulation produces neural crest cells, which are a transient multipotent cell population that comes from the dorsal edge of the closing neural tube [24]. These cells become highly migratory upon undergoing an epithelial-mesenchymal transition (EMT) and spread according to precise embryonic routes directed by a complex network of molecular signals such as chemotactic gradients, cell adhesion molecules and interactions with the extracellular matrix [25,26]. The neural crest cells take part in a broad and fascinating distribution, which has led to their derivatives being involved in the formation of a remarkable spectrum of peripheral sensory and autonomic ganglia, Schwann cells, melanocytes, the adrenal medulla, and the cartilage and bone of craniofacial structures, demonstrating their predominant role in establishing the structural complexity of numerous organ systems [28, 27]. Their functional roles are enormous as they contribute to the development of functional sensory-motor reflex loops and efficient impulse conduction in peripheral nervous system through the role in peripheral ganglia and Schwann cells [29]. In addition, they are incorporated into the enteric nervous system to coordinate intestinal motility and metabolic functions autonomously, and they are differentiated into melanocytes that give pigment to the skin to protect from UV-induced photodamage [30]. From the structural point of view their craniofacial skeletal derivatives offer mechanical support for respiration, mastication and speech [31]. Therefore, failures in neural crest specification, migration or differentiation cause serious congenital neurocristopathies, including pigment abnormalities, intestinal aganglionosis (Hirschsprung disease), and craniofacial defects highlighting the far-reaching physiological consequences of this lineage [32].

Together, the rest of the surface ectoderm differentiates along a different route to form the stratified squamous epithelium that will make up the epidermis of the skin [33]. This morphogenetic process involves the progressive differentiation of cells from the basal proliferating layer to the superficial keratinized layers and generates a multilayered epithelial covering that is well suited to deal with environmental exposure [34, 35]. The barrier is architecturally highly stratified, and the keratinocyte cells are tightly connected to one another by strong desmosomal and tight junctions [36]. The sequential assembly of keratinized proteins into the intercellular matrix and the cornification of the terminal keratin at the end of each unit leads to a thick barrier, which is selectively permeable and mechanically resistant, forming the outer barrier [37]. This special tissue structure has important physiological significance and provides strong mechanical protection against external mechanical and microbial stresses [38]. In addition, it plays an active regulatory role in thermoregulation by controlling sweat gland function and peripheral vascular reactions [39] and it also inhibits excessive sweating to prevent loss of fluid and electrolytes from the body and therefore prevents disturbance of systemic fluid and electrolyte balance [40].

5. MESODERMAL PATTERNING AND SYSTEMIC INTEGRATION: PARAXIAL MESODERM AND SOMITOGENESIS

The paraxial mesoderm is responsible for the formation of a ventro-dorsal series of segments, called somite's, along the cranio-caudal axis, which are the basic segments of embryonic segmentation [41]. Within each of the somites, there are three different lineages of cells: the sclerotome, myotome, and dermatome [42] that together form the basic skeletal plan for the axial musculoskeletal organization [43]. This concept of segmental patterning has been adopted for the precise alignment of the vertebral bodies together with their associated skeletal muscles and overlying dermis, from an architectural point of view [44]. They are spatially coordinated to produce structural symmetry and biomechanical stability, resulting in a highly synchronized musculoskeletal system of biomechanical forces that are conducive to functional movement along the skeleton axis [45]. Such somite-derived muscles are

innervated in segments, which gives rise to the physiologic organization of voluntary motor activity [46]. The exact anatomical layout of this localization enables the fine proprioceptive information to be transmitted via sensorimotor pathways, which are coordinated [47], and the early developmental segmentation is an anatomical basis for adult locomotion and postural control [48].

Simultaneously, the formation of cardiac morphogenesis begins in the cardiogenic mesoderm, and during primitive organogenesis, the primary heart tube loops, forms chambers, and is divided into two by septation [49]. They are complex morphogenetic movements which establish the spatial orientation and specific identity of cardiac chambers [50] and contribute to the structural remodeling of the immature heart into an organ that pumps systemic blood circulation [51]. The architectural features during early development include myocardial trabeculation that increases the internal surface area and also raises the contractile capacity [52]. The heart is further subdivided by means of subsequent septation, which ensures that oxygenated and deoxygenated blood remain physically apart [53], and the coordinated contraction of the heart is structurally supported by the coordinated alignment of myocardial fibers [54]. The physiological results of this design are dramatic: the hemodynamic workload of the heart depends completely on the chamber and valve shapes determined structurally [55]. Moreover, the cardiac conduction system differentiation is critical for the production and propagation of rhythmic electrical signals [56] and showcases that cardiac architecture is a direct cause of the circulatory competence [57].

Concurrently, a system of homeostasis and adaptive defenses is formed: First, hematopoietic stem cells are produced in the hemogenic endothelium of the embryo; later, the organs of definitive hematopoiesis develop and are colonized and activated by these cells [58]. These undifferentiated cells have the ability for unlimited self-renewal and immunogenicity and can expand into different cell lineages [58]. These lineages differentiate to form erythrocytes, leukocytes and platelets, which have structural and morphological specializations that enable them to carry out extremely specific systemic functions, namely oxygenation, immune protection and hemostasis, respectively [59]. This cellular heterogeneity gives rise to the framework needed to maintain systemic homeostasis and adaptive immune responses throughout life [59].

Finally, the intermediate mesoderm induces the development of the excretory system by inducing and reciprocating inductive signals from the metanephric mesenchyme to the epithelial ureteric bud, the latter inducing nephrogenesis and spatial cell organization [60]. This reciprocal cross-talk sets the defining structural plan for the mature kidney, which involves the organization of the segmented nephron (glomerulus, proximal tubule, loop of Henle, and distal tubule) into a functional continuum [61]. This fine structural compartmentalization allows to selectively regulate filtration and reabsorption processes at different nephron segments [61]. From a functional perspective, the unique configuration enables the countercurrent exchange mechanism required for the effective concentration of urine, and also performs acid-base balance, fluid volume and continuous maintenance of systemic hypertension [62].

6. ENDODERMAL DIFFERENTIATION AND METABOLIC REGULATION: GASTROINTESTINAL MORPHOGENESIS

Reciprocal interactions between the epithelial and mesenchymal components are required to achieve regionalization and regulation of the differentiation of the primitive endoderm to give rise to the specialized lining epithelium of the digestive tract in a manner that results in a digestive tract identity along the anterior-posterior axis [63]. The large surface area of absorption can be achieved from the complex villus morphogenesis on an architectural level, while the crypt stem cell niches guarantee a continuous renewal of the epithelial surface [64]. This dynamic tissue architecture combines good high-capacity nutrient absorption with good structural barrier properties. Physiologically, proper nutrient intake will rely entirely on the large surface area of this epithelial sheet and the specific transport mechanisms; at the same time, the integrity of the barrier and the ability to keep luminal pathogens out of the body are maintained by local mucosal immunity and by maintaining the tight junction structure intact [65].

The hepatocytes start to differentiate in an inductive process between the foregut endoderm and the septum transversum mesenchyme, and form the liver bud, which matures and vascularizes into a highly organized parenchymal organ [66]. Structurally, the hepatic lobules are organized around the central vein-portal triad axis, with a microanatomical structure that has been optimized for efficient metabolic exchange, coordinated blood flow, and bile drainage [67]. The unique spatial organization of hepatocytes enables the continuous processing of nutrients, production of plasma proteins, and maintenance of glycogen stores through efficient nutrient processing and systemic distribution, whereas the organized vascular sinusoids facilitate continuous detoxification [68]. Together, pancreatic organogenesis promotes the differentiation of endodermal progenitors into specific endocrine islets and exocrine acinar units, with differential morphogen signaling that regulates this compartmentalization

[69]. The physiological importance of this strict compartmentalization of architecture is enormous because the two separate functions, endocrine and exocrine, enable the independent but coordinated secretion of digestive enzymes and systemic hormones to maintain global glucose homeostasis [70].

The upper panel shows gastrointestinal morphogenesis and the development of villi and crypt stem cell niches for the continuous renewal of epithelial cells and for increasing the absorptive surface area, as well as important physiological features such as tight junctions and mucosal immunity. The lower panel illustrates the cell differentiation of the liver and pancreas; the left one displays the structural organization of a hepatic lobule, optimized for metabolic exchange and excretion of bile; the right one represents the liver hepatocyte functional compartmentalization (detoxification, protein synthesis, glucose metabolism), and the dual endocrine-exocrine architecture of the pancreas. Created using BioRender.com, integrating its built-in Artificial Intelligence (AI) generation tools.

7. MOLECULAR REGULATION OF TISSUE ARCHITECTURE: SIGNALING PATHWAYS

Developmental signaling pathways such as the Wnt, Hedgehog, BMP, FGF and Notch pathways comprise a core conserved group that is integral to orchestrating the lineage commitment and spatial patterning of developing tissues [71]. These pathways work through the formation of accurate concentration gradients of morphogens that provide positional information and indicate different structural domains in the embryo [71]. In morphogenesis, fidelity of the spatial signal and the timing of pathways are essential, and any imbalance or disruption of these pathways can cause very serious organ architecture deformation, which can be responsible for congenital birth defects or trigger the onset of neoplastic cell adhesion [72]. It is therefore essential that these signally networks are precisely spatiotemporally regulated to guarantee normal, predictable morphogenesis in all the embryonic lineages [72].

The epigenetic mechanisms by which the transcriptional programs that create the identity of a cell are complex and may involve DNA methylation and covalent modifications of histones [73] that stabilize these developmental decisions over time. Such structural changes in the chromatin structure provide the molecular memory to store the differentiated state for long-term periods, causing specialized cells to retain their functional identity [73]. On the other hand, abnormal epigenetic regulation can completely break the integrity of tissues, initiate developmental syndromes, or cause severe metabolic diseases [74]. Therefore, chromatin dynamics should be regarded as a high-level organization of chromatin structure that is involved in protecting the stability of the architectural plan of the embryo [74].

In addition to chemical and genetic signals, the physical inputs that are generated during tissue growth can also significantly impact the organization of the cytoskeleton and the nuclear signaling pathways involved in the cell's response to the extracellular environment, a process known as mechanotransduction [75]. These biomechanical stimuli are actively sensed and responded to by cells, with the cells undergoing remodeling of their internal and external structures in order to optimize the spatial organization and enable correct differentiation [75]. This mechanical feedback loop is very important during several morphogenetic events such as cardiac tube looping, pulmonary branching and musculoskeletal development [76]. In the end, this suggests that maturation of tissue architecture is not purely a genetic process but is instead the result of an ongoing interplay of epigenetic stabilization, genetic programs and mechanical determinants.

8. DEVELOPMENTAL ORIGINS OF ADULT PHYSIOLOGY AND DISEASE

The Developmental Origins of Health and Disease (DOHaD) paradigm is a strong theoretical concept that states that the embryonic/fetal environment is a critical determinant for long-term physiological outcomes in adulthood [77]. Specific developmental structural deficits provide a striking illustration of this paradigm: A normal or limited nephron development during the early stages of nephrogenesis is directly associated with the development of chronic hypertension in later years of life [77]. In the same way, failure to develop a developmental disability or a developmental disruption in the pancreas beta-cells affects lifelong insulin secretion and causes the onset of diabetes, and a disorganized cardiac septum during early primitive organogenesis causes permanent congenital heart defects [77]. Therefore, these developmental and structural determinants are physiologically expressed in the vulnerability to disease and the capacity of an individual in adulthood [77].

9. TRANSLATIONAL IMPLICATIONS: REGENERATIVE MEDICINE, ORGANOID TECHNOLOGY, AND TISSUE ENGINEERING

To exploit these embryological principles, regenerative medicine directly incorporates these factors by trying to mimic the intricate signaling patterns of the early embryo to direct stem cell differentiation towards particular, functional adult tissues [78]. It is hoped that these well-preserved developmental processes can be rejuvenated to provide structurally organized and physiologically competent functional tissues that can be used to replace damaged or congenitally absent organs [78]. Three-dimensional, organoid technologies serve as a priceless in vitro model that helps to reflect significant parts of embryonic morphogenesis, and thus enhance this translational journey [79]. These are engineered organisms that render a true-to-life spatial structure and organ-specific physiology of tissues in development and thus offer an unprecedented platform for the study of real-time human development, elucidation of disease pathophysiology and testing of individualized therapeutic responses [79, 80]. Simultaneously, tissue engineering combines these cell models with advanced biomaterials which replicate the structure of the embryonic extracellular matrix and the critical mechanical cues [80]. This discipline allows optimal scaffold design and strict control of the microenvironment so as to achieve seamless structural integration and complete physiological functional recovery in vivo.

10. INTEGRATIVE PERSPECTIVE: FROM EMBRYO TO SYSTEM PHYSIOLOGY

Translation of genetic and mechanical blueprints into a living organism is a very coordinated, hierarchical developmental process. The sequential process can be condensed into a number of successive steps, starting with the initial cell differentiation, through to complex morphogenesis, ultimately to system integration to form an entire, mature tissue, a cascade that is essential to the development of a mature tissue's complex structure [81]. In this hierarchy, the functional potential of each anatomical level is determined by the organization. Thus, for the mature human, physiology should not be seen as a condition but the dynamic functional manifestation of early embryonic, structural programming [81].

11. CONCLUSION

In the highly orchestrated process of embryogenesis, a slew of structural and molecular patterns are set up to systematically demarcate tissue architecture and functional physiological specialization. Germ layer specification, morphogenetic signaling networks, epigenetic changes and biomechanical forces all function in a defined continuum in order to dictate specific functions of the definitive organs. This is a key challenge to completely integrate classical embryology with current systems biology and clinical medicine, and an even stronger imperative to do so given the profound, unbreakable association between adult physiology and developmental origin. Finally, innovations in developmental science not only advance our basic understanding of normal physiology but also provide game-changing and transformative insights into the prevention of congenital disorders, the pathogenesis of chronic diseases, and the development of new regenerative therapeutic strategies.

Conflict of interest

The authors state that they do not have any complaints about financial or personal relationships that could have biased the work reported in this paper.

Consent for publications

All authors read and approved the final manuscript for publication.

Availability of data and material

All the data collected and analyzed in the process of this research work is included and embedded in this published article.

Authors' contributions

Zainab and Rajaa. I jointly conceived the original idea, designed the conceptual framework, and analysed the integrated developmental data of this study. Z.M. conducted the primary literature research, compiled the embryological and physiological findings, and drafted the initial manuscript, while R.I. evaluated the molecular signaling pathways, structured the translational implications, and critically revised the intellectual content of the text. Both authors contributed equally to the final synthesis of the article, and they have read and approved the completed manuscript for publication.

Funding

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

Acknowledgement

The authors would like to express their sincere gratitude and appreciation to the Department of Biology, College of Education for Pure Science, Kirkuk University, Kirkuk, Iraq, for the required institutional support and scientific environment that enabled the successful completion and writing of this article.

12. REFERENCES

- [1] Denker, H. W. (2024). Embryoids, models, embryos? We need to take a new look at legal norms concerning the beginning of organismic development. *Molecular Human Reproduction*, 30(1), gaad047. <https://doi.org/10.1093/molehr/gaad047>
- [2] Huang, H., Gao, S., & Bao, M. (2024). Exploring mechanical forces shaping self-organization and morphogenesis during early embryo development. *Annual Review of Cell and Developmental Biology*, 40(1), 75-96. <https://doi.org/10.1146/annurev-cellbio-120123-105748>
- [3] Arrangoiz, R., Cordera, F., Caba, D., Muñoz, M., Moreno, E., de León, E. L., ... & Muñoz, M. (2018). Comprehensive review of thyroid embryology, anatomy, histology, and physiology for surgeons. *International Journal of Otolaryngology and Head & Neck Surgery*, 7(4), 160-188. <https://doi.org/10.4236/ijohns.2018.74019>
- [4] Nadolski, E. M., & Moczek, A. P. (2023). Promises and limits of an agency perspective in evolutionary developmental biology. *Evolution & Development*, 25(6), 371-392. <https://doi.org/10.1111/ede.12432>
- [5] Walma, D. A. C., & Yamada, K. M. (2020). The extracellular matrix in development. *Development*, 147(10), dev175596. <https://doi.org/10.1242/dev.175596>
- [6] Strough, J., & Bruine de Bruin, W. (2020). Decision making across adulthood. *Annual Review of Developmental Psychology*, 2(1), 345-363. <https://doi.org/10.1146/annurev-devpsych-051120-010038>
- [7] Soczyńska, J., Gawelczyk, W., Obrycka, P., Żołyński, M., Muzyka, A., Majcherczyk, K., ... & Woźniak, S. (2025). Medical embryology and regenerative medicine: research and applications in clinical practice. *Frontiers in Cell and Developmental Biology*, 13, 1619036. <https://doi.org/10.3389/fcell.2025.1619036>
- [8] Lázár, E., & Lundberg, J. (2026). Spatial architecture of development and disease. *Nature Reviews Genetics*, 27(2), 118-136. <https://doi.org/10.1038/s41576-025-00892-5>
- [9] Pandey, D., Sharma, S., & Sharma, U. (2025). Embryogenesis. In *Advances in Growth Regulation of Fruit Crops* (pp. 115-127). CRC Press. <https://doi.org/10.1201/9781003354055-11>
- [10] Kai, Y., Mei, H., Kawano, H., Nakajima, N., Takai, A., Kumon, M., ... & Yamashita, N. (2022). Transcriptomic signatures in trophectoderm and inner cell mass of human blastocysts classified according to developmental potential, maternal age and morphology. *PLoS One*, 17(12), e0278663. <https://doi.org/10.1371/journal.pone.0278663>
- [11] Hassani, S. N., Moradi, S., Taleahmad, S., Braun, T., & Baharvand, H. (2019). Transition of inner cell mass to embryonic stem cells: mechanisms, facts, and hypotheses. *Cellular and molecular life sciences*, 76(5), 873-892. <https://doi.org/10.1007/s00018-018-2965-y>
- [12] Tam, P. P. L. (2024). Cell Lineages, Fate Determination, and Differentiation Pathways. In *Developmental Biology and Cancer* (pp. 187-209). CRC Press. <https://doi.org/10.1201/9781003575153-10>
- [13] Ghelman, R. (2026). Development Biology and Morphogenetic Fields. In *Information Fields Theory and Applications: Quantum Communication in Physics and Biology* (pp. 139-162). Singapore: Springer Nature Singapore. https://doi.org/10.1007/978-981-95-1742-8_7
- [14] Ghimire, S., Mantziou, V., Moris, N., & Arias, A. M. (2021). Human gastrulation: The embryo and its models. *Developmental biology*, 474, 100-108. <https://doi.org/10.1016/j.ydbio.2021.01.006>
- [15] Srinivas, S., & Watanabe, T. (2025). Establishment of early embryonic lineages and the basic body plan. In *Kaufman's Atlas of Mouse Development Supplement* (pp. 67-77). Academic Press. <https://doi.org/10.1016/B978-0-443-23739-3.00004-3>
- [16] Kühn, S., Magno, V., Freudenberg, U., & Werner, C. (2025). Engineering Approaches to Control Morphogen Gradients in Tissue Models. In *Advanced Polymer Life Science* (pp. 457-489). https://doi.org/10.1142/9789819806782_0012
- [17] Tickle, C. (2024). Patterning and Morphogenesis and the Development of Organized Tissues. In *Developmental Biology and Cancer* (pp. 129-160). CRC Press. <https://doi.org/10.1201/9781003575153-8>
- [18] Gupta, N., & Ross, M. E. (2026). Disorders of neural tube development. In *Swaian's Pediatric Neurology* (pp. 331-340). Elsevier. <https://doi.org/10.1016/B978-0-443-10944-7.00040-8>

- [19] Saade, M., & Martí, E. (2025). Early spinal cord development: from neural tube formation to neurogenesis. *Nature Reviews Neuroscience*, 26(4), 195-213. <https://doi.org/10.1038/s41583-025-00906-5>
- [20] Akeret, K., Weller, M., & Krayenbühl, N. (2023). The anatomy of neuroepithelial tumours. *Brain*, 146(8), 3133-3145. <https://doi.org/10.1093/brain/awad138>
- [21] Mubuchi, A., Takechi, M., Nishio, S., Matsuda, T., Itoh, Y., Sato, C., ... & Miyata, S. (2024). Assembly of neuron-and radial glial-cell-derived extracellular matrix molecules promotes radial migration of developing cortical neurons. *Elife*, 12, RP92342. <https://doi.org/10.7554/eLife.92342.3>
- [22] Eichler, A., Kruse, P., Schob, C., & Lenz, M. (2025). Synaptic transmission in supragranular layers of the human cortex-comparative review of structure, function, and plasticity. *Frontiers in Synaptic Neuroscience*, 17, 1724377. <https://doi.org/10.3389/fnsyn.2025.1724377>
- [23] Montgomery, R. (2024). The Role and Impact of Myelination in the Adult Brain: Cognitive Functions, Neurological Health, Brain Efficiency, and Comparisons to Deep Learning Perceptrons. <https://doi.org/10.20944/preprints202408.0317.v1>
- [24] Motohashi, T., & Kunisada, T. (2026). Neural Crest Cells: Their Multipotency and Plasticity. *Stem Cells and Development*, 35(1-2), 1-12. <https://doi.org/10.1177/15473287251405289>
- [25] Yang, J., Antin, P., Berx, G., Blanpain, C., Brabletz, T., Bronner, M., ... & EMT International Association (TEMIA). (2020). Guidelines and definitions for research on epithelial-mesenchymal transition. *Nature reviews Molecular cell biology*, 21(6), 341-352. <https://doi.org/10.1038/s41580-020-0237-9>
- [26] Fortunato, I. C., & Sunyer, R. (2022). The forces behind directed cell migration. *Biophysica*, 2(4), 548-563. <https://doi.org/10.3390/biophysica2040046>
- [27] Kalcheim, C. (2025). The association between neural crest-derived glia and melanocyte lineages throughout development and disease. *Developmental Dynamics*. <https://doi.org/10.1002/dvdy.70098>
- [28] Andrews, T. G., & Priya, R. (2025). The mechanics of building functional organs. *Cold Spring Harbor perspectives in biology*, 17(3), a041520. <https://doi.org/10.1101/cshperspect.a041520>
- [29] Wawrzyniak, A., Krawczyk-Marć, I., Żuryń, A., Kłosowicz, M., Walocha, J., Wysiadecki, G., & Balawender, K. (2025). Unveiling significance of peripheral nervous system glia: implications for nervous system disorders and therapeutic interventions. *Folia Morphologica*, 84(3), 493-500. <https://doi.org/10.5603/fm.102401>
- [30] Ascsillán, A. A., & Kemény, L. V. (2024). The skin-brain Axis: From UV and pigmentation to behaviour modulation. *International Journal of Molecular Sciences*, 25(11), 6199. <https://doi.org/10.3390/ijms25116199>
- [31] Hosomichi, J., & Ono, T. (2026). A functional axis of mastication and respiration in cognitive function: A narrative review. *Archives of Oral Biology*, 106517. <https://doi.org/10.1016/j.archoralbio.2026.106517>
- [32] Vermeij-Keers, C., Mathijssen, I. M., Trainor, P., & ten Donkelaar, H. J. (2023). The Neural Crest and Craniofacial Malformations. In *Clinical Neuroembryology: Development and Developmental Disorders of the Human Central Nervous System* (pp. 313-378). Cham: Springer International Publishing. https://doi.org/10.1007/978-3-031-26098-8_5
- [33] Yin, H., Hu, M., & Li, D. (2023). Regulation of epidermal stratification and development by basal keratinocytes. *Journal of cellular physiology*, 238(4), 742-748. <https://doi.org/10.1002/jcp.30978>
- [34] Nosrati, H., Fallah Tafti, M., Aghamollaei, H., Bonakdar, S., & Moosazadeh Moghaddam, M. (2024). Directed differentiation of adipose-derived stem cells using imprinted cell-like topographies as a growth factor-free approach. *Stem Cell Reviews and Reports*, 20(7), 1752-1781. <https://doi.org/10.1007/s12015-024-10767-7>
- [35] Jedel, E., Schator, D., Kumar, N. G., Sullivan, A. B., Rietsch, A., Evans, D. J., & Fleiszig, S. M. (2025). The *Pseudomonas aeruginosa* T3SS can contribute to traversal of an in situ epithelial multilayer independently of the T3SS needle. *Mbio*, 16(4), e00266-25. <https://doi.org/10.1128/mbio.00266-25>
- [36] Leprince, C., & Simon, M. (2025). Epidermal lamellar bodies, essential organelles for the skin barrier. *Frontiers in Cell and Developmental Biology*, 13, 1597884. <https://doi.org/10.3389/fcell.2025.1597884>
- [37] Fernandes, M. G., da Silva, L. P., & Marques, A. P. (2019). Skin mechanobiology and biomechanics: From homeostasis to wound healing. *Advances in Biomechanics and Tissue Regeneration*, 343-360. <https://doi.org/10.1016/B978-0-12-816390-0.00017-0>
- [38] Jia, B., Du, X., Wang, W., Qu, Y., Liu, X., Zhao, M., ... & Li, Y. Q. (2022). Nanophysical antimicrobial strategies: a rational deployment of nanomaterials and physical stimulations in combating bacterial infections. *Advanced Science*, 9(10), 2105252. <https://doi.org/10.1002/advs.202105252>
- [39] Cramer, M. N., Gagnon, D., Laitano, O., & Crandall, C. G. (2022). Human temperature regulation under heat stress in health, disease, and injury. *Physiological reviews*. <https://doi.org/10.1152/physrev.00047.2021>
- [40] Kitada, K., & Nishiyama, A. (2025). Water and sodium metabolisms in kidney-skin crosstalk. *Physiology*. <https://doi.org/10.1152/physiol.00038.2025>

- [41] Toulouse, G. (2024). Development and Growth of Limb and Epaxial Musculature in Amniotes (Doctoral dissertation, Lyon 1). <https://doi.org/10.70675/6d3199a9z5bdcz4cd6z87e0z9c11a4dc70c6>
- [42] Della Gaspera, B., & Chanoine, C. (2023). The lateral somitic frontier: The source of multipotent somitic cells in *Xenopus*. *Medicine Sciences: M/S*, 39(12), 967-974. <https://doi.org/10.1051/medsci/2023181>
- [43] Mallo, M. (2025). The axial musculoskeletal system. In *Kaufman's atlas of mouse development supplement* (pp. 281-296). Academic Press. <https://doi.org/10.1016/B978-0-443-23739-3.00027-4>
- [44] Rajendran, B., Yunos, M. A., Nagalinggam, H., Aziz, M. L. A., & Abdullah, J. M. (2025). Surface Anatomy and Sensory Evaluation of Dermatomes: A Guide for Residents. *The Malaysian Journal of Medical Sciences: MJMS*, 32(1), 169. <https://doi.org/10.21315/mjms-09-2024-738>
- [45] Herzog, W. (2017). Skeletal muscle mechanics: questions, problems and possible solutions. *Journal of neuroengineering and rehabilitation*, 14(1), 98. <https://doi.org/10.1186/s12984-017-0310-6>
- [46] Sasikumar, S., Bhan, A., & Rajendra, T. K. (2018). Stem Cells for Nerve and Muscle Repair: Harnessing Developmental Dynamics in Therapeutics. In *Stem Cells for Cancer and Genetic Disease Treatment* (pp. 149-186). Cham: Springer International Publishing. https://doi.org/10.1007/978-3-319-98065-2_10
- [47] Florio, T. M. (2025). Emergent aspects of the integration of sensory and motor functions. *Brain Sciences*, 15(2), 162. <https://doi.org/10.3390/brainsci15020162>
- [48] Grillner, S., & El Manira, A. (2020). Current principles of motor control, with special reference to vertebrate locomotion. *Physiological reviews*, 100(1), 271-320. <https://doi.org/10.1152/physrev.00015.2019>
- [49] Paredes-Espinosa, M. B., & Paluh, J. L. (2025). Synthetic embryology of the human heart. *Frontiers in Cell and Developmental Biology*, 12, 1478549. <https://doi.org/10.3389/fcell.2024.1478549>
- [50] Liu, Y., Xue, X., Sun, S., Kobayashi, N., Kim, Y. S., & Fu, J. (2024). Morphogenesis beyond in vivo. *Nature Reviews Physics*, 6(1), 28-44. <https://doi.org/10.1038/s42254-023-00669-x>
- [51] Rabadia, J. P., Thite, V. S., Desai, B. K., Bera, R. G., & Patel, S. (2024). Cardiovascular system, its functions and disorders. In *Cardioprotective Plants* (pp. 1-34). Singapore: Springer Nature Singapore. https://doi.org/10.1007/978-981-97-4627-9_1
- [52] Petersen, S. E., Jensen, B., Aung, N., Friedrich, M. G., McMahon, C. J., Mohiddin, S. A., ... & Bluemke, D. A. (2023). Excessive trabeculation of the left ventricle: JACC: cardiovascular imaging expert panel paper. *Cardiovascular Imaging*, 16(3), 408-425. <https://doi.org/10.1016/j.jcmg.2022.12.026>
- [53] Arts, T., Delhaas, T., Bovendeerd, P., Verbeek, X., & Prinzen, F. W. (2005). Adaptation to mechanical load determines shape and properties of heart and circulation: the CircAdapt model. *American Journal of Physiology-Heart and Circulatory Physiology*, 288(4), H1943-H1954. <https://doi.org/10.1152/ajpheart.00444.2004>
- [54] Liu, L., Xu, F., Jin, H., Qiu, B., Yang, J., Zhang, W., ... & Sun, D. (2023). Integrated manufacturing of suspended and aligned nanofibrous scaffold for structural maturation and synchronous contraction of HiPSC-derived cardiomyocytes. *Bioengineering*, 10(6), 702. <https://doi.org/10.3390/bioengineering10060702>
- [55] El-Nashar, H., Sabry, M., Tseng, Y. T., Francis, N., Latif, N., Parker, K. H., ... & Yacoub, M. H. (2024). Multiscale structure and function of the aortic valve apparatus. *Physiological Reviews*, 104(4), 1487-1532. <https://doi.org/10.1152/physrev.00038.2022>
- [56] Kléber, A. G., & Jin, Q. (2021). Coupling between cardiac cells-An important determinant of electrical impulse propagation and arrhythmogenesis. *Biophysics Reviews*, 2(3). <https://doi.org/10.1063/5.0050192>
- [57] Sanz, J., Sánchez-Quintana, D., Bossone, E., Bogaard, H. J., & Naeije, R. (2019). Anatomy, function, and dysfunction of the right ventricle: JACC state-of-the-art review. *Journal of the American College of Cardiology*, 73(12), 1463-1482. <https://doi.org/10.1016/j.jacc.2018.12.076>
- [58] Calvanese, V., & Mikkola, H. K. (2023). The genesis of human hematopoietic stem cells. *Blood*, 142(6), 519-532. <https://doi.org/10.1182/blood.2022017934>
- [59] Menter, D. G., Fowlkes, N. W., Honn, K. V., & Sood, A. K. (2025). The Evolution and Role of Platelets in Cancer Biology and Metastasis. In *Platelets in Disease: Thrombotic Disorders and Disorders not Involving Hemorrhage or Thrombosis: Volume 4* (pp. 1789-1806). Cham: Springer Nature Switzerland. https://doi.org/10.1007/978-3-031-96352-0_19
- [60] Schnell, J., Achieng, M., & Lindström, N. O. (2022). Principles of human and mouse nephron development. *Nature Reviews Nephrology*, 18(10), 628-642. <https://doi.org/10.1038/s41581-022-00598-5>
- [61] Herold, L. (2016). The renal system. Monitoring and Intervention for the Critically Ill Small Animal: The Rule of 20, 225-245. <https://doi.org/10.1002/9781118923870.ch13>
- [62] Đambić, V., & Kibel, A. (2025). Pressure Natriuresis in Blood Pressure Control. <https://doi.org/10.5772/intechopen.1010604>

- [63] Loe, A. K. H., Rao-Bhatia, A., Kim, J. E., & Kim, T. H. (2021). Mesenchymal niches for digestive organ development, homeostasis, and disease. *Trends in Cell Biology*, 31(3), 152-165. <https://doi.org/10.1016/j.tcb.2020.11.010>
- [64] Bonis, V., Rossell, C., & Gehart, H. (2021). The intestinal epithelium-fluid fate and rigid structure from crypt bottom to villus tip. *Frontiers in cell and developmental biology*, 9, 661931. <https://doi.org/10.3389/fcell.2021.661931>
- [65] Zang, J., Xiao, L., Shi, Y., Kou, Y., Ma, K., Zhang, C., ... & Li, W. (2026). Advances in Dietary Modulation of the Intestinal Barrier: Mechanistic, Structural, and Functional Insights. *Comprehensive Reviews in Food Science and Food Safety*, 25(1), e70383. <https://doi.org/10.1111/1541-4337.70383>
- [66] Lotto, J., Stephan, T. L., & Hoodless, P. A. (2023). Fetal liver development and implications for liver disease pathogenesis. *Nature Reviews Gastroenterology & Hepatology*, 20(9), 561-581. <https://doi.org/10.1038/s41575-023-00775-2>
- [67] Crawford, J. M. (2025). Anatomy and histopathology of the liver. In *Hepatology* (pp. 1-25). Academic Press. <https://doi.org/10.1016/B978-0-443-26710-9.00001-8>
- [68] Chesney, R. W., Han, X., & Patters, A. B. (2010). Taurine and the renal system. *Journal of biomedical science*, 17(Suppl 1), S4. <https://doi.org/10.1186/1423-0127-17-S1-S4>
- [69] Dutta, S., Mishra, S. P., Sahu, A. K., Mishra, K., Kashyap, P., & Sahu, B. (2021). Hepatocytes and their role in metabolism. In *Drug Metabolism*. IntechOpen . <https://doi.org/10.5772/intechopen.99083>
- [70] Hu, C., Chen, Y., Yin, X., Xu, R., Yin, C., Wang, C., & Zhao, Y. (2025). Pancreatic endocrine and exocrine signaling and crosstalk in physiological and pathological status. *Signal transduction and targeted therapy*, 10(1), 39. <https://doi.org/10.1038/s41392-024-02098-3>
- [71] Kicheva, A., & Briscoe, J. (2023). Control of tissue development by morphogens. *Annual review of cell and developmental biology*, 39(1), 91-121. <https://doi.org/10.1146/annurev-cellbio-020823-011522>
- [72] Fourikou, M., & Dotis, J. (2025). From Genes to Malformations: Molecular Mechanisms Driving the Pathogenesis of Congenital Anomalies of the Kidney and Urinary Tract. *International Journal of Molecular Sciences*, 27(1), 17. <https://doi.org/10.3390/ijms27010017>
- [73] Gagnidze, K., & Pfaff, D. W. (2022). Epigenetic mechanisms: DNA methylation and histone protein modification. In *Neuroscience in the 21st century: from basic to clinical* (pp. 2677-2716). Cham: Springer International Publishing. https://doi.org/10.1007/978-3-030-88832-9_69
- [74] Wu, Y. L., Lin, Z. J., Li, C. C., Lin, X., Shan, S. K., Guo, B., ... & Li, Z. H. (2023). Epigenetic regulation in metabolic diseases: mechanisms and advances in clinical study. *Signal Transduction and Targeted Therapy*, 8(1), 98. <https://doi.org/10.1038/s41392-023-01333-7>
- [75] Zhang, X., Zhang, S., & Wang, T. (2022). How the mechanical microenvironment of stem cell growth affects their differentiation: A review. *Stem cell research & therapy*, 13(1), 415. <https://doi.org/10.1186/s13287-022-03070-0>
- [76] Pesce, M., Duda, G. N., Forte, G., Girao, H., Raya, A., Roca-Cusachs, P., ... & Van Linthout, S. (2023). Cardiac fibroblasts and mechanosensation in heart development, health and disease. *Nature Reviews Cardiology*, 20(5), 309-324. <https://doi.org/10.1038/s41569-022-00799-2>
- [77] Gluckman, P. D., & Hanson, M. A. (2006). The developmental origins of health and disease. *Early life origins of health and disease*, 1-7. https://doi.org/10.1007/0-387-32632-4_1
- [78] Ding, S. (2025). Therapeutic reprogramming toward regenerative medicine. *Chemical Reviews*, 125(4), 1805-1822. <https://doi.org/10.1021/acs.chemrev.4c00332>
- [79] Bhattacharya, R., Bose, D., Kaur, T., Patel, R., Renuka, O., & Rodriguez, R. V. (2025). Model organoids: integrated frameworks for the next frontier of healthcare advancements. *Stem Cell Reviews and Reports*, 21(2), 319-336. <https://doi.org/10.1007/s12015-024-10814-3>
- [80] Mangani, S., Vetoulas, M., Mineschou, K., Spanopoulos, K., Vivanco, M. D., Piperigkou, Z., & Karamanos, N. K. (2025). Design and applications of extracellular matrix scaffolds in tissue engineering and regeneration. *Cells*, 14(14), 1076. <https://doi.org/10.3390/cells14141076>
- [81] Ghelman, R. (2026). Development Biology and Morphogenetic Fields. In *Information Fields Theory and Applications: Quantum Communication in Physics and Biology* (pp. 139-162). Singapore: Springer Nature Singapore. https://doi.org/10.1007/978-981-95-1742-8_7



الأساس الجنيني للبنية النسيجية وأثره على الوظائف الحيوية: التكامل الفسيولوجي التطوري: مراجعة

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الكلمات المفتاحية	الملخص
التخليق الجنيني؛ بنية الأنسجة؛ الطبقات الجرثومية؛ التخليق الشكلي؛ تخلق الأعضاء.	أثناء التخليق الجنيني، يتم تحديد التنظيم البنيوي لأنسجة جسم الإنسان بشكل أساسي من خلال التنسيق الدقيق للتمايز الخلوي، والتخليق الشكلي، والتنظيم المكاني. وتتكون المخططات التطورية لجميع الأنسجة والأجهزة العضوية من الطبقات الجرثومية الثلاث الرئيسية: الأديم الظاهر، والأديم المتوسط، والأديم الباطن. تتشكل بنية الأنسجة على المستويين المجهرية والجهري تحت تأثير التدرجات الجزيئية المبكرة، والبرمجة فوق الجينية، والقوى الميكانيكية الحيوية، مما يضع الشروط البنيوية المسبقة للتخصص الوظيفي (الفسيولوجي). تناقش هذه المراجعة العلمية بالتفصيل الأسس التطورية لبنية الأنسجة، وتقدم تحليلاً نقدياً لكيفية تحديد النمذجة التطورية للوظائف الفسيولوجية لدى البالغين. ويسلط البحث اهتماماً خاصاً على تمايز الطبقات الجرثومية، وشبكات الإشارات الخلوية، والتفاعلات الظهارية-المتوسطة، وتخلق الأعضاء، والتروية الوعائية، والتكامل العصبي الوظيفي. علاوة على ذلك، تتطرق المراجعة إلى التطبيقات التحويلية لبيولوجيا التطور في الأمراض الخلقية، والطب التجديدي، وفيزيولوجيا الأنظمة. إن فهم المحددات التطورية لبنية الأنسجة يوفر إطاراً مترابطاً يربط بين علم الأجنة، وعلم الأنسجة، وعلم وظائف الأعضاء، والطب السريري.

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